

Association between Methylenetetrahydrofolate Reductase C677T Polymorphisms, Plasma Homocysteine Levels, and Primary Open-Angle Glaucoma in Jordanian Patients

Mahmoud Kaswal¹, Mohammad Abu-Jeyyab², Khalid Alzubi³,
Fawaz Alsarireh⁴, Hala Fouad⁵ and Samir Mahgoub^{6*}

¹Hematology-Oncology Department, ST Bartholomew Hospital, Barts Health NHS Trust, London, UK.

²Anesthesia and Intensive Care Department, Istishari hospital, Amman, Jordan.

³Special Surgeries Department, Faculty of Medicine, Mutah University, Al-Karak, Jordan.

⁴Ophthalmology Department, AlQassim Hospital, AlQassim Saudi Arabia.

⁵Department of Anatomy, Histology and Embryology, Faculty of Medicine, Mutah University, Al-Karak, Jordan.

⁶Department of Biochemistry and Molecular Biology, Faculty of Medicine, Mutah University, Al-Karak, Jordan.

*Corresponding Author E-mail:samir_mhgb@yahoo.com

<https://dx.doi.org/10.13005/bpj/3315>

(Received: 18 September 2025; accepted: 10 November 2025)

Primary open-angle glaucoma (POAG) is a chronic, progressive optic neuropathy, recognized as the second most common cause of irreversible blindness globally. It is marked by the gradual degeneration of retinal ganglion cells, leading to optic nerve head cupping, optic disc excavation, and progressive visual field loss. The study aimed to investigate the relationship between POAG, methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism, and levels of plasma homocysteine, a well-known vascular risk factor amongst POAG Jordanian patients. This case-control study included 183 participants: 89 POAG patients and 94 non-glaucomatous controls. In total, blood samples from all participants were analyzed for assessing plasma homocysteine levels and MTHFR C677T polymorphisms. Homocysteine levels were measured using ELISA, while the MTHFR C677T polymorphisms were identified through polymerase chain reaction restriction fragment length polymorphism (PCR-RFLP) analysis. Our findings revealed that plasma homocysteine levels and hyperhomocysteinemia rate were significantly higher in POAG patients versus the controls ($p < 0.05$). In contrast, no significant differences were identified in allelic frequency and MTHFR C677T polymorphisms distribution between POAG patients and controls. Elevated homocysteine levels and a higher hyperhomocysteinemia prevalence have been observed in POAG patients, suggesting a potential role in the disease's pathogenesis. Furthermore, a statistically significant association and increased risk linked to the CT and TT polymorphisms, as well as T allele, indicate that MTHFR C677T polymorphisms may contribute to POAG development in the Jordanian population.

Keywords: Alleles; Homocysteine; MTHFR C677T; Polymorphisms; POAG.

Glaucoma is an irreversible optic neuropathy that can cause permanent loss of the visual field.¹In 2010, an estimated 4.5 million and 3.9 million individuals with POAG and primary

angle-closure glaucoma (PACG), were bilaterally blind, furthermore by the year 2020, these figures had increased to 5.9 million and 5.3 million, respectively.²⁻⁴

Higher intraocular pressure (IOP) is widely recognized as the most significant modifiable risk factor for glaucoma onset and its progress,⁵ particularly POAG. Among patients with primary glaucoma, IOP typically peaks in the morning and gradually decreases through the day in non-clinical settings.⁶

Primary glaucoma is broadly categorized into two main types: POAG, the most common one and PACG.² The main risk factors for primary open-angle glaucoma (POAG) are elevated intraocular pressure (commonly above 21 mmHg), a thin central corneal thickness (under 500 μ m), a family history of the disease, black ethnicity, and high levels of myopia.⁷

Various genetic studies have revealed genetic polymorphisms of different genes with uncertain effects that are linked to POAG including C677T polymorphisms of MTHFR gene,⁸ some variants of genes encoding for membrane palmitoylated protein⁷ (MPP7), tumor protein p53,⁹ GRIN2B gene 421C/A, Arg72Pro,¹⁰ and soluble guanylate cyclase sGC α 1.¹¹

MTHFR enzyme plays a vital role in regulating folate, homocysteine (HCY) and methionine metabolism and in maintaining normal plasma HCY levels.¹² HCY is a key intermediate in the metabolism of methionine and cysteine, it is generated through the hydrolysis of S-adenosylhomocysteine in the cycle of methionine. MTHFR catalyzes 5,10-methylenetetrahydrofolate reduction to 5-methylenetetrahydrofolate which is the main circulating form of folate and the donor of methyl groups for remethylating HCY to methionine in a reaction catalyzed by a B12-dependent methionine synthase enzyme.¹³

A 677C/T mutation at position 222 in exon 4 of MTHFR gene results in converting alanine to valine codon. As a consequence of this mutation, MTHFR enzymatic activity is reduced comparable to the wild type (CC) enzyme¹⁴ and the enzyme's thermal stability increases,¹⁵ thus, mutations of MTHFR gene are associated with higher levels of HCY.¹⁶

Elevated plasma levels of HCY have been observed in glaucoma patients.¹⁷ HCY has been implicated in vascular injury¹⁸ alterations of the extracellular matrix,¹⁹ and neuronal cell death due to excitotoxicity and apoptosis,²⁰ also, hyperhomocysteinemia has also been associated

with structural remodeling of connective tissues²¹ and various neurodegenerative disorders.²²

The aim of this study was to examine the potential link between MTHFR C677T polymorphisms and plasma HCY levels in individuals with POAG, with the objective of enhancing our understanding of the disease's underlying pathogenic mechanisms.

MATERIALS AND METHODS

183 subjects participated in this study, among them 89 POAG patients and 94 non-glaucomatous individuals who served as the control group. All participants were recruited from the Outpatient Clinics of Special Surgeries Department/Ophthalmology Division at Al-Karak Governmental Hospital, Al-Karak, Jordan. The exclusion criteria from the study included patients who had any ocular diseases other than glaucoma that could elevate IOP, or if they had cancer, diabetes mellitus, and chronic kidney disease, or patients who received immunosuppressants, antimicrobials, hormonal therapies, lipid-lowering drugs, vitamin supplements, anticonvulsants and antidepressants treatment within six months prior to the study. All participants (both patients and controls) were subjected to complete ophthalmic examination, including testing of visual acuity with the Humphrey 24-2 protocol, slit-lamp biomicroscopy, measurement of IOP using a Goldmann applanation tonometer, assessment of central corneal thickness via pachymetry, gonioscopy with a Goldmann 3-mirror lens, optic nerve imaging by OCT (optical coherence tomography). Patients were diagnosed with POAG based on optic nerve damage confirmed by OCT, along with an elevated IOP greater than 21 mmHg, and an open anterior chamber angle on gonioscopy according to Shaffer angle grading. The POAG group included 89 patients (51 males and 38 females), aged between 40 and 74 years, with mean \pm SD 58.73 \pm 9.47 years. Control group consisted of individuals with healthy-appearing optic discs, IOP below 21 mmHg, a cup-to-disc ratio less than 0.4, and normal visual field assessments in both eyes. None of the controls had a history of using IOP-lowering medications or a family history of glaucoma, the group included 94 participants (55 males and 39 females), with ages

between 42 and 76 years with mean \pm SD 59.26 \pm 10.39 years.

Biochemical Analyses

Plasma HCY assay

12 hours fasting blood sample was collected from each participant in the study in EDTA-treated tubes for plasma HCY estimation, then, immediately centrifuged at 4°C, plasma fraction was withdrawn for storage at -20°C until further analysis while, cellular portion was preserved for extraction of DNA.

Plasma HCY levels were measured using ELISA, according to the method adopted by Engvall *et al.*²³. Hyperhomocysteinemia was defined as a HCY concentration exceeding 15 μ mol/L.²⁴

MTHFR polymorphisms analysis

Based on Lahiri and Nurnberger,²⁵ method, genomic DNA was extracted from leukocytes, then, MTHFR C677T polymorphisms in the gene were analyzed using PCR-RFLP technique, as reported in a previous study.²⁶ The sequences of the primers used for amplification of a 198 bp fragment of the targeted DNA were: Forward: 5'-TGA AGG AGA AGG TGT CTG CGG GA-3' Reverse: 5'-AGG ACG GTG CGG TGA GAG TG-3'.

PCR was initiated with denaturation at 94°C for 2 minutes, followed by 35 cycles of denaturation at 94°C for 30 seconds, annealing at 62°C for 30 seconds, and extension at 72°C for 30 seconds. A final extension was performed at 72°C for 7 minutes.

5 units of *Hinf*I enzyme were used for digesting PCR products for 12 hours at 37°C. Electrophoresis was performed on a 2% agarose gel. The digestion yielded a single 198 bp band for CC polymorphism (wild-type), two bands of 175 bp and 23 bp for CT polymorphism (the heterozygous-type), and a 175 bp and 23 bp for TT polymorphism (homozygous mutant). The 23 bp fragment cannot be seen because of its too small size.

Statistical analysis

SPSS software package version 25.0 was used for data analysis which were presented as mean \pm SD, while, variables in categories were expressed as percentages, student's t-test was applied for the assessment of the differences between continuous variables, whereas Chi-square (χ^2) test was used to evaluate the differences between categorical variables and to test Hardy-Weinberg equilibrium by comparing the observed and expected polymorphisms frequencies. A p-value less than 0.05 was regarded as statistically significant.

RESULTS

Table 1 shows insignificant differences between POAG patients and controls in terms of age, hypertension, diabetes mellitus, or cardiovascular disorders. However, the differences in plasma HCY levels and hyperhomocysteinemia rates were significant.

Table 1. The biological and clinical characteristics of participants in POAG and control groups

Variables	POAG patients (no. 89)	Controls (no. 94)
Gender	Male/female (51/38)	Male/female (55/39)
Age (range) (mean \pm SD)	40-74 (58.73 \pm 9.47)*	42-76 (59.26 \pm 10.39)
Hypertension	53 (60.1%)*	55 (58.6%)
Diabetes mellitus	40 (44.9%)*	45 (47.4%)
Cardiovascular disorder	22 (24.4%)*	13 (13.5%)
Homocysteine (μ mol/l) (mean \pm SD)	29.59 \pm 1.49**	10.14 \pm 0.96
Hyperhomocysteinemia	33 (37%)**	19 (20.3%)

* P > 0.05 is insignificant versus the controls

** P < 0.05 is significant when compared to the controls

Table 2. Distribution of polymorphisms of MTHFR C677T and allelic frequencies in POAG patients and control groups

Groups	C677T Polymorphism			Allele	
	**CC (no.) %	**CT (no.) %	**TT (no.) %	**C (no.) %	**T (no.) %
POAG group (no. 89)	43 (48.8%)	28 (31.5%)	18 (19.7%)	114 (64.0 %)	64 (36%)
Controls (no. 94)	54 (57.9%)	25 (26.3%)	15 (15.8%)	189 (70.7%)	55 (29.%)
Odds ratio (95% CI)	1.00	1.38 (0.98-1.95)	1.21(0.87-1.88)	1.00	1.53 (0.97-2.49)
P*	0.461	0.633	0.662	0.316	0.405

* χ^2 test

**CC (Cytosine-Cytosine), TT (Thymine-Thymine), CT (Cytosine-Thymine), C (Cytosine), T (Thymine).

Table 2 presents C677T polymorphisms' distribution and frequencies of alleles in POAG patients and controls. Among POAG patients, the polymorphisms distribution was 48.8% CC, 31.5% CT, and 19.7% TT. In the control group, the distribution was 57.9% CC, 26.3% CT, and 15.8% TT. Also, the results revealed significant association and positive risk ratio between CT, TT polymorphisms and T allele in POAG patients (OR=1.38, 95%CI=0.98-1.95), (OR=1.21, 95%CI=0.87-1.88) (OR=1.53, 95%CI=0.97-2.49), respectively, while, no significant risk ratio regarding CC polymorphism and C allele was detected.

DISCUSSION

Glaucoma represents a group of chronic eye disorders distinguished by the progressive loss of retinal ganglion cells (RGCs), thinning of the retinal nerve fiber layer, and a gradual decline in visual field. Among these conditions, POAG is the most common type and has been the focus of extensive research. The etiology of POAG is complex and multifactorial, encompassing interactions between environmental influences and significant genetic predisposition. Family linkage analyses have revealed at least 17 genetic loci associated with the disease. Within this framework, MTHFR gene emerges as a compelling candidate due to its critical function in one-carbon

metabolism and its regulatory role in maintaining HCY levels.^{9, 27}

Our findings demonstrated that plasma homocysteine (HCY) levels were significantly elevated in Jordanian patients with primary open-angle glaucoma (POAG) compared to healthy controls. This observation suggests a potential association between hyperhomocysteinemia and POAG, indicating that elevated HCY may contribute to the disease's pathogenesis. Consistent with this hypothesis, Leibovitch *et al.*²⁸ reported a higher prevalence of hyperhomocysteinemia and increased mean plasma HCY levels in glaucoma patients, accompanied by vascular abnormalities that may play a role in disease progression. Similarly, Bleich *et al.*²⁹ observed a twofold increase in HCY concentrations in the aqueous humor of glaucoma patients, suggesting a possible intraocular contribution of HCY to the disease mechanism. Moreover, Roedl *et al.*³⁰ found elevated HCY levels in tear fluid, providing additional evidence of its pathogenic involvement. Collectively, these observations strengthen the hypothesis that HCY may act as a biochemical mediator in glaucomatous optic nerve damage. Nonetheless, previous studies have yielded divergent outcomes. Wang *et al.*³¹ observed no significant difference in plasma HCY levels between patients with POAG and healthy controls. Likewise, Tongabay *et al.*³² found elevated HCY concentrations only in individuals

with pseudoexfoliation glaucoma (PEXG), but not in those with POAG. Consistently, Turaçlı *et al.*³³ also reported no significant variation in HCY levels between PEXG patients and control subjects, despite a high overall prevalence of hyperhomocysteinemia in both groups.

These discrepancies may be attributed to methodological variations, differences in study design, limited sample sizes, or the influence of ethnic and dietary factors. Specifically, variations in dietary intake of folate and B vitamins key regulators of HCY metabolism as well as differences in the analytical methods used to measure plasma HCY levels, could account for population-specific differences observed across studies.

Our study revealed no significant differences in the MTHFR C677T polymorphism between POAG patients and controls, in terms of both polymorphisms and allele frequencies. The distributions of CT and TT polymorphisms, as well as the C and T alleles, were nearly identical across the two groups. Consistently, odds ratio analysis showed no significant association between this polymorphism and the risk of developing POAG.

These findings align with several previous studies. George *et al.*^{3t} reported no significant association between MTHFR C677T polymorphisms and either POAG or pseudoexfoliation glaucoma. Similarly, Mabuchi *et al.*^{3u} observed no differences in polymorphism frequencies between patients and controls. A study conducted in a Pakistani population^{3v} also found no significant link between MTHFR C677T polymorphisms and POAG; however, they did report a significant association between the TT genotype and primary closed-angle glaucoma (PCAG), suggesting that the influence of this polymorphism may vary depending on the glaucoma subtype.

Our results indicate that MTHFR C677T polymorphisms modulate HCY levels; however, they do not appear to be a significant risk factor for POAG in our population. It is likely that the pathogenic role of hyperhomocysteinemia in POAG arises from a complex interplay of genetic predisposition, environmental factors, and nutritional status, rather than a single genetic mutation.

In our Jordanian cohort, the T allele frequency of the MTHFR C677T polymorphism

was 18% among POAG patients. By comparison, previous studies have reported frequencies of 24% in Brazilians, 9% in West Africans, 55% in Mainland Chinese, and 33% in Turks^{3w} τp , highlighting significant global variation. These differences may affect statistical power and influence the strength of genetic associations reported across populations.

Moreover, other polymorphisms within the MTHFR gene or in genes involved in the HCY metabolic pathway such as MTR, MTRR, or CBS may exert a greater influence in our population, potentially through gene-gene or gene-environment interactions.

CONCLUSION

This study demonstrates significantly elevated HCY levels and higher rates of hyperhomocysteinemia in Jordanian POAG patients compared to controls, suggesting a contributory role in disease pathogenesis. MTHFR C677T polymorphisms distribution and C allele frequency were higher among individuals with the homozygous CC polymorphisms compared to those with CT or TT polymorphisms and T allele. Furthermore, the results indicate that elevated HCY levels rather than MTHFR polymorphisms are associated with the severity of POAG. This suggests that MTHFR C677T polymorphisms are unlikely to be a significant independent risk factor for POAG. Clinically, this suggests that elevated HCY levels regardless of genetic background warrant further investigation be carried out on a larger number of populations as a potentially modifiable biochemical risk factor in glaucoma management.

ACKNOWLEDGEMENT

The authors would like to thank all staff members in the Special Surgeries Department / Ophthalmology Division, Faculty of Medicine, Mutah University and all participants in the study.

Funding Sources

The author(s) received no financial support for the research, authorship, and/or publication of this article.

Conflict of Interest

The author(s) do not have any conflict of interest.

Data Availability Statement

This statement does not apply to this article.

Ethics Statement

This research did not involve human participants, animal subjects, or any material that requires ethical approval.

Informed Consent Statement

This study did not involve human participants, and therefore, informed consent was not required.

Clinical Trial Registration

This research does not involve any clinical trials

Permission to reproduce material from other sources

Not Applicable

Author contributions

Mahmoud Kaswal: Conceptualization, visualization; Mohammad Abu-Jeyyab: Data collection, analysis; Khalid Alzubi: Supervision, methodology, editing; Fawaz Alsarireh: Supervision, methodology, editing; Hala Fouad: Project administration, supervision; Samir Mahgoub: Project administration, methodology, writing – original draft, writing – review & editing.

REFERENCES

1. The Japanese Archive of Multicentral Database in Glaucoma (JAMDIG) construction group. A novel method to predict visual field progression more accurately, using intraocular pressure measurements in glaucoma patients. *Sci. Rep.* 2016, 6, 31728-31735. <https://doi.org/10.1038/srep31728>
2. Quigley HA, Broman AT. The number of people with glaucoma worldwide in 2010 and 2020. *Br. J. Ophthalmol.* 2006, 90, 262-267. DOI: 10.1136/bjo.2005.081224
3. Friedman DS, Foster PJ, Aung T, He M. Angle closure and angle-closure glaucoma: what we are doing now and what we will be doing in the future. *Clin. Exp. Ophthalmol.* 2012, 40, 381-387. doi: 10.1111/j.1442-9071.2012.02774.x
4. Tham YC, Li X, Wong TY, Quigley HA, Aung T, Cheng CY. Global prevalence of glaucoma and projections of glaucoma burden through 2040: a systematic review and meta-analysis. *Ophthalmology* 2014, 121, 2081-2090. doi: 10.1016/j.ophtha.2014.05.013.
5. The Japan Glaucoma Society guidelines for glaucoma (3rd edition). *Nippon GankaGakkaiZasshi* 2012, 116 (1), 3–46. PMID: 22352070
6. Shaoying T, Nafees B, Linda H, et al. Comparison of self-measured diurnal intraocular pressure profiles using rebound tonometry between primary angle closure glaucoma and primary open angle glaucoma patients. *PLoS ONE* 2017, 12(3), e0173905. doi: 10.1371/journal.pone.0173905
7. Janssen SF, Gorgels TG, Ramdas WD, et al. The vast complexity of primary open angle glaucoma: disease genes, risks, molecular mechanisms and pathobiology. *ProgRetin Eye Res.* 2013, 37, 31–67. doi: 10.1016/j.preteyeres.2013.09.001.
8. Al-Shahrani H, Al-Dabbagh N, Al-Dohayan N, et al. Association of the methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism with primary glaucoma in Saudi population. *BMC Ophthalmol.* 2016, 16(1), 156. doi: 10.1186/s12886-016-0337-7
9. Mansi V, Anchal S, Lalit K, et al. Genetic association and stress mediated down-regulation in trabecular meshwork implicates MPP7 as a novel candidate gene in primary open angle glaucoma. *BMC Medical Genomics* 2016, 9, 15. doi: 10.1186/s12920-016-0177-6.
10. Alicja N, Karolina P, Katarzyna S, Jerzy S, Jacek PS, Ireneusz M. The relationship of TP53 and GRIN2B gene polymorphisms with risk of occurrence and progression of primary open-angle glaucoma in a Polish Population. *Pol J Pathol.* 2014, 65 (4), 313-321. DOI:10.5114/pjp.2014.48193
11. Buys ES, Ko YC, Alt C, et al. Soluble guanylate cyclase $\alpha 1$ -deficient mice: a novel murine model for primary open angle glaucoma. *PLoS One* 2013, 8(3), e60156. doi: 10.1371/journal.pone.0060156
12. Wilson CP, McNulty H, Scott JM, Strain JJ, Ward M. Postgraduate symposium: the MTHFR C677T polymorphism, B-vitamins and blood pressure. *Proc Nutr Soc.* 2010, 69, 156–165. DOI: 10.1017/S0029665109991728
13. Huang T, Wahlqvist ML, Li D. Effect of n-3 polyunsaturated fatty acid on gene expression of the critical enzymes involved in homocysteine metabolism. *Nutr J.* 2012, 11, 6. doi: 10.1186/1475-2891-11-6
14. Taioli E, Garza MA, Ahn YO, et al. Meta- and pooled analyses of the methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism and colorectal cancer: a HuGE-GSEC review. *Am J Epidemiol.* 2009, 170, 1207–1221. doi: 10.1093/

- aje/kwp275.
15. Zonouzi AP, Chaparzadeh N, Estiar AM, et al. Methylenetetrahydrofolate reductase C677T and A1298C mutations in women with recurrent spontaneous abortions in the Northwest of Iran. *ISRN ObstetGynecol.* 2012, 2012, 945486. doi: 10.5402/2012/945486.
 16. Xu H, Liu C, Wang Q. Plaque image characteristics, hyperhomocysteinemia, and gene polymorphism of homocysteine metabolism-related enzyme (MTHFR C677T) in acute coronary syndrome. *Cell Biochem Biophys.* 2013, 66, 403–407. doi: 10.1007/s12013-012-9483-6
 17. Bleich S, Junemann A, von Ahsen N, et al. Homocysteine and risk of open-angle glaucoma. *J Neural Transm.* 2002, 109, 1499-504. doi: 10.1007/s007020200097
 18. McCully KS. Chemical pathology of homocysteine. I. Atherosclerosis. *Ann Clin Lab Sci.* 1993, 23, 477-93. PMID: 8291902
 19. Tyagi SC. Homocysteine redox receptor and regulation of extracellular matrix components in vascular cells. *Am J Physiol.* 1998, 274, C396-405. doi: 10.1152/ajpcell.1998.274.2.C396
 20. Moore P, El-sherbeny A, Roon P, Schoenlein PV, Ganapathy V, Smith SB. Apoptotic cell death in the mouse retinal ganglion cell layer is induced in vivo by the excitatory amino acid homocysteine. *Exp Eye Res.* 2001, 73, 45-57. doi: 10.1006/exer.2001.1009
 21. Mujumdar VS, Tummalapalli CM, Aru GM, Tyagi SC. Mechanism of constrictive vascular remodeling by homocysteine: role of PPAR. *Am J Physiol Cell Physiol.* 2002, 282, C1009-15. doi: 10.1152/ajpcell.00353.2001
 22. Brustolin S, Giugliani R, Felix TM. Genetics of homocysteine metabolism and associated disorders. *Braz J Med Biol Res.* 2010, 43, 1–7. doi: 10.1590/s0100-879x2009007500021.
 23. Engvall E, Jonsson K, Perlmann P. Enzyme-Linked Immunosorbent Assay. II. Quantitative assay of protein antigen, immunoglobulin G, by means of enzyme-labeled antigen and antibody-coated tubes. *Biochim. Biophys. Acta* 1971, 251, 427-434. doi: 10.1016/0005-2795(71)90132-2
 24. Borson-Chazot F, Harthe C, Teboul F, et al. Occurrence of Hyperhomocysteinemia 1 Year after Gastrectomy for Severe Obesity. *J Clin Endocrinol Metab.* 1999, 84(2), 541-5. doi: 10.1210/jcem.84.2.5476
 25. Lahiri DK, Nurnberger JI Jr. A rapid non-enzymatic method for the preparation of HMW DNA from blood for RFLP studies. *Nucleic Acids Res.* 1991, 19, 5444. doi: 10.1093/nar/19.19.5444
 26. Papoutsakis C, Yiannakouris N, Manios Y, et al. Plasma Homocysteine Concentrations in Greek Children Are Influenced by an Interaction between the Methylenetetrahydrofolate Reductase C677T Genotype and Folate Status. *J Nutr.* 2005, 135(3), 383-8. doi: 10.1093/jn/135.3.383
 27. Junemann AG, von Ahsen N, Reulbach U, et al. C677T variant in the methylenetetrahydrofolate reductase gene is a genetic risk factor for primary open-angle glaucoma. *Am J Ophthalmol.* 2005, 139, 721-3. doi: 10.1016/j.ajo.2004.09.081
 28. Leibovitch I, Kurtz S, Shemesh G, et al. Hyperhomocysteinemia in pseudoexfoliation glaucoma. *Journal of Glaucoma* 2003, 12(1), 36-39. doi: 10.1097/00061198-200302000-00007
 29. Bleich S, Roedl J, Ahsen VN, et al. Elevated Homocysteine levels in aqueous humor of patients with pseudoexfoliation glaucoma. *American Journal of Ophthalmology* 2004, 138(1), 162–164. doi: 10.1016/j.ajo.2004.02.027
 30. Roedl JB, Bleich S, Reulbach U, et al. Homocysteine in tear fluid of patients with pseudoexfoliation glaucoma. *J. Glaucoma* 2007, 16, 234-9. doi: 10.1097/IJG.0b013e31802d6942
 31. Wang G, Medeiros FA, Barshop BA, Weinreb RN. Total plasma homocysteine and primary open-angle glaucoma. *American Journal of Ophthalmology* 2004, 137(3), 401-406. doi: 10.1016/j.ajo.2003.09.041
 32. Tongabay C, Semsetin S, Erdinc A. Serum homocysteine, vitamin B12 and folic acid levels in different types of glaucoma. *BMC Ophthalmology* 2006, 6, 6. doi: 10.1186/1471-2415-6-6
 33. Turaçlı ME, Tekeli O, Özdemir F, Akar N. Methylenetetrahydrofolate reductase 677 C-T and homocysteine levels in Turkish patients with pseudoexfoliation. *Clinical and Experimental Ophthalmology* 2005, 33, 505–508. doi: 10.1111/j.1442-9071.2005.01070.x
 34. George M, Martin W, Christoph F, et al. Methylenetetrahydrofolate reductase (MTHFR) 677C>T polymorphism and open angle glaucoma. *Molecular Vision* 2006, 12, 356-9. PMID: 16636653
 35. Mabuchi F, Tang S, Kashiwagi K, Yamagata Z, Iijima H, Tsukahara S. Methylenetetrahydrofolate reductase gene polymorphisms c.677c/t and c.1298a/c are not associated with open angle glaucoma. *Mol Vis.* 2006, 12, 735-9. PMID: 16862068
 36. Shazia M, Raheel Q, Farah A, Muhammad IK, Wajid AK, Asifa A. MTHFR gene c677t and a1298c polymorphisms and homocysteine levels in primary open angle and primary closed angle glaucoma. *Molecular vision* 2009, 15, 2268-2278. PMID: 19936026

37. Franco RF, Araújo AG, Guerreiro JF, Elion J, Zago MA. Analysis of the 677 C>T mutation of the methylenetetrahydrofolate reductase gene in different ethnic groups. *ThrombHaemost.* 1998, 79, 119-121. DOI: 10.1055/s-0037-1614230
38. Sazci A, Ergul E, Kaya G, Kara I. Genotype and allele frequencies of the polymorphic methylenetetrahydrofolate reductase gene in Turkey. *CellBiochemFunct.* 2005, 23, 51-54. doi: 10.1002/cbf.1132
39. Angeline T, Jeyaraj N, Granito S, Tsongalis GJ. Prevalence of MTHFR gene polymorphisms (C677T and A1298C) among Tamilians. *ExpMolPathol.* 2004, 77, 85-88. doi: 10.1016/j.yexmp.2004.04.006
40. Mohsin Y, Naushad M, Siddiqi P, Bushra C, Iqbal A, Mohammad PI. Polymorphisms in MTHFR, MS and CBS Genes and Homocysteine Levels in a Pakistani Population. *PLoS ONE* 2012, 7(3), e33222. doi: 10.1371/journal.pone.0033222