Subacute Oral Toxicity Study of a New Polyherbal Formulation of Usadha on liver and Kidney Function of Wistar Rats

I Nyoman Arsana^{1*}, Ni Ketut Ayu Juliasih¹, Anak Agung Ayu Sauca Sunia Widyantari¹, I Gede Widhiantara² and Putu Angga Wiradana²

 ¹Department of Biology, Faculty of Information Technology and Science, University of Hindu Indonesia, Jl. Sangalangit, Tembau, Penatih, Denpasar, Bali Province, Indonesia.
²Department of Biology, Faculty of Health and Science, Universitas Dhyana Pura, Jalan Raya Padangluwih, Dalung, North Kuta, Badung Regency, Bali Province, Indonesia.
*Corresponding Author E-mail: arsanacita@gmail.com

https://dx.doi.org/10.13005/bpj/3138

(Received: 12 February 2025; accepted: 24 March 2025)

Usadha is a traditional Balinese medicine system that has long been used in ancient medicine systems. However, so far there have been no reports on its safety. This study aims to examine the effect of polyherbal Usadha on hematology and blood biochemistry of Wistar rats. The study used a Completely Randomized Design with six treatments, namely polyherbal Usadha supplementation with doses of 0 mg/kg bw/day (E0), 100 mg/kg bw/day (E1), 200 mg/kg bw/day (E2), 300 mg/kg bw/day (E3), 400 mg/kg bw/day (E4), and 500 mg/kg bw/day (E5). The treatments were given for four weeks. The variables measured were complete hematology and blood biochemistry. Data were analyzed by One-Way ANOVA and continued with the Least Significant Difference test at a 95% confidence interval. Polyherbal Usadha appears to be safe for consumption without significant risk of side effects on blood parameters and liver function. Additional studies including histological analysis and mechanism of action will provide deeper insight into the therapeutic effects of polyherbal Usadha. In conclusion, polyherbal usada is relatively safe for liver and kidney function in Wistar rats. The results of this study support the safety of using Poliherbal Usadha as a candidate herbal medicine for long-term consumption.

Keywords: Blood biochemistry; Citrus amblycarpa (Hassk.) Ochse; Haematology; Piper nigrum L; Polyherbal Usadha.

Traditional medicine is gaining popularity because it supplies phytopharmaceuticals for the development of pharmacological medicines with desired qualities.¹ Herbal plants have been utilized for thousands of years to treat a variety of ailments across the world. According to the World Health Organization (WHO), 80% of the world's population continues to rely largely on traditional medicines for basic treatment². Herbal medications are more popular than allopathic therapies because they are less expensive, have less side effects, are as effective as conventional pharmaceuticals, are more readily available, and are ecologically benign.²

Indonesia has the world's sixth most diverse flora, with 20,000 species, 40% of which are indigenous to the country.³ The existence of native Indonesian flora is fascinating to research, particularly in the health industry. Furthermore, research on medicinal plants has increased

This is an d Open Access article licensed under a Creative Commons license: Attribution 4.0 International (CC-BY). Published by Oriental Scientific Publishing Company © 2025



dramatically in recent years, suggesting a growing interest in natural-based traditional medicine.4 Indonesia is well-known for its sustainable use of traditional medicine5. Traditional Indonesian herbal medicine has been used for hundreds of years in Indonesian society and is still widely used to maintain health and cure ailments due to its perceived safety over chemical pharmaceuticals.6,7 These herbs have historically played a significant role in Indonesian culture and medicine.8-10 Herbal plants, such as ginger, turmeric, and betel leaves, have been employed in traditional "Jamu" (herbal medicine) for thousands of years, demonstrating their long-term relevance in sustaining health and well-being. In addition to Jamu, the terminology for herbal medicinal beverages in Indonesia differ widely by location or ethnicity.11,12

Bali Province has a long history of treating various diseases with herbal plant components in various forms such as powder, decoction, paste, and oil contained in the Balinese traditional medicine system known as "Usadha". The formulation of these medicines is based on two principles, namely utilizing one plant or using many herbs. Combining many herbs, often known as "Polyherbal", is hypothesized to increase therapeutic efficacy because various bioactive chemicals work on various therapeutic targets to accelerate the healing process.¹³ Polyherbal has been used for thousands of years in many countries to treat various types of disorders such as diabetes, as an antioxidant, hepatoprotective, anti-inflammatory, and anxiety disorders.

Polyherbal derived from a mixture of Murraya koenigii L. Spreng leaves, Allium sativum L., Garcinia quaesita Pierre fruits, and Piper nigrum L. seeds is a common herbal remedy in Sri Lanka for treating diabetes mellitus and dyslipidemia.14 It has been found to be nontoxic and safe for long-term use. The polyherbal, which includes plant extracts including Boerhavia diffusa, Solidago virgaurea, Vitex negundo, and thymoquinone compounds, helps reduce hepatorenal damage caused by CCl₄ through antioxidant and anti-inflammatory processes.15 AYUSH-64 is an Ayurvedic herbal polyphenol formulation consisting of many herbal components such as Alstonia Scholaris, Picrorhiza kurroa, Swertia chirata, and Caesalpinia crista, has been used for various disorders including malaria,

chronic fever, and joint pain.¹⁶ Herbal mixture consisting of Cassia absus, Gymnema sylvestre, Nigella sativa, and Piper nigrum has high potential in treating diabetic complications, including liver and kidney damage, dyslipidemia, and oxidative stress.¹⁷ Arthralgex Ayurvedic Polyherbal consisting of 18 plant species did not cause serious side effects on biochemical, hematological, or organ parameters of rats during and after the treatment period.¹⁸ One of the polyherbal of usada formulas is made from a mixture of lime leaves (Citrus amblycarpa), 11 white peppercorns (Piper nigrum L.), and vinegar. These ingredients are ground until smooth, then mixed and used by drinking. This formula is used to treat paresthesia. Pepper (Piper *nigrum* L), has been known to have many active compounds, two of which have a high abundance of Piperidine, 1-[5-(1,3-benzodioxol-5-yl)-1-oxo-2,4-pentadienyl]-,(Z,Z)-; and Caryophyllene.¹⁹

The toxicity of herbal medicines has raised widespread concerns, and various side effects (including allergic responses, hepatotoxicity, nephrotoxicity, and cardiac toxicity) have been described in recent years.²⁰ Plants with medicinal properties should have minimal toxicity due to their long-term use in humans.²¹ However, many therapeutic plants used in traditional medicine have been associated with harmful consequences. Conducting toxicity testing in appropriate animal models is essential to ensure the safety of treatments and is at the heart of toxicology. According to OECD standards, acute toxicity studies are an advanced approach to administer single and multiple doses of chemicals and report the primary safety profile of a particular compound.²²

There are several articles that discuss in an exploratory manner the types of medicinal plants collected in *Usadha* in the province of Bali ^{23–25}; but no empirical facts have been found regarding the safety or benefits of polyherbal *Usadha*. On the other hand, there is a high demand for polyherbals in health services. Therefore, this study was designed to evaluate the acute toxicity properties of polyherbal *Usadha* formulations on kidney and liver function in Wistar rats. The results of this study will be an excellent effort to encourage manufacturers and researchers to examine the optimal composition of raw materials when designing herbal medicines.^{26–28}

MATERIALS AND METHODS

Choice of plant combinations

Lime leaves (*Citrus amblycarpa*) and white pepper (*Piper nigrum* L.) were obtained from local farmers in Bali Province. Both materials were selected for research purposes based on their ethnopharmacological properties and their claims in the Lontar Usadha. Lime leaves and black pepper seeds were separated and dried in the shade. The dried leaves and seeds were then ground and sieved and filtered with a 600 mesh sieve size. The powdered plant samples were stored in sterile airtight zip lock bags at 4 ! until used.²⁸

Preparation of crude extract

Each of the simplicia was then extracted by maceration method using 96% ethanol solvent for 48 hours, and re-macerated twice, filtered using filter paper. The filtrate was then concentrated in a vacuum rotary evaporator at a temperature of 45! until a thick extract was obtained and then dried by the freeze-dry method. Usada poliherbal is made from a mixture of three ingredients, namely; white pepper seed extract, lime leaf extract, and vinegar in equal proportions.

Animals

A total of 24 male Wistar rats (*Rattus norvegicus*), aged three months, weighing 200 g to 250 g were used in this study. The rats were grouped into six groups of four each and then acclimatized for one week to adjust to the temperature and humidity of the research room. The rats were kept in cages containing four rats each, given food and drink as much as they wanted. The rats were given polyherbal usada at a dose according to the treatment group for four weeks through the stomach.

Sub-acute toxicity test

The experiment was conducted following OECD guidelines number 407 in the study.²⁹ The study used a Completely Randomized Design with six treatments. The treatment was the administration of polyherbal usadha with six dose levels, namely; 0 mg/kgbb/day (E0), 100 mg/kgbb/ day (E1), 200 mg/kgbb/day (E2), 300 mg/kgbb/day (E3), 400 mg/kgbb/day (E4), and 500 mg/kgbb/ day (E5). Each treatment was repeated four times so that there were 24 research units. The treatment was given for four weeks. Twenty-four hours after the last treatment, the mice were euthanized using ketamine, then blood was taken from the orbital sinus cantus using a capillary pipette and stored in a tube.

Blood biochemical analysis

Biochemical studies were conducted to evaluate the safety profile of the polyherbal extract of usadha. Biochemical investigations included aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), Blood urea nitrogen (BUN), SC, and Total protein (TP). Blood biochemical parameters were determined using a medical analyzer with fuitest whose working procedure was in accordance with the company's instructions.¹

Hematological analysis

Blood samples from the treatment and control groups were taken and placed into 3 mL vaculab tubes containing EDTA. The parameters used to indicate hematological conditions were RBC (Red Blood Cell), HGB (Hemoglobin), HCT (Hematocrit), MCV (mean cell volume), MCH (Mean cell hemoglobin), MCHC (mean cell hemoglobin concentration), RDW (Red cell distribution width); WBC (white blood cells), Eo (eosinophils), Baso (basophils) Lymp (Lymphocyte), Neut (Neutrophils), Mono (Monocytes), PTL (Platelets), PDW (Platelet distribution width), MPV (Mean platelet volume), PCT (Plateletcrit). These parameters were calculated using the Sysmex pocH-100i[™] Haematology Analyzer and the CBC line kit.1 Statistical analysis

The measurement data were then tabulated in Microsoft Excel and evaluated statistically using One Way ANOVA with Duncan's Test. Results are presented as mean \pm SD. The statistical significance value between groups was at pd"0.05.²⁹

RESULTS

Blood Biochemical Levels

The results showed that the administration of Polyherbal *Usadha* had a significant effect (pd"0.05) on ALT and ALP, but not significantly (pe"0.05) on AST, BUN, SC, and TP. The average ALT, AST, BUN, TP, and SC tended to decrease after administering Polyherbal *Usadha*, while ALP increased after a 300 mg/kg bw dose. The mean ALT appeared to be significantly lower than the control after administration of 100 mg/kg bw to 500 mg/kg bw doses. Meanwhile, ALP appeared to be significantly higher than the control at a dose of 400 mg/kg bw (Table 1).

The results showed that ALP levels showed a pattern that varied depending on the dose. The administration of polyherbal *usadha* at doses of 100 and 200 mg/kg bw resulted in lower ALP than the control. Still, it then increased significantly compared to the control at the dose of 400 mg/kg bw (Table 1). This condition indicates that administering polyherbal *usadha* at a dose of 400 mg/kg bw causes a certain metabolic response. The results showed that SC, BUN, and TP were not significantly different from the control (Table 1). **Blood Hematology Levels**

Research shows that several hematological parameters after administration of Polyherbal Usadha show a pattern of changes that vary and depend on the dose. The administration of Polyherbal Usadha had a significant effect (pd"0.05) on MCV, PDW, Basophil, and MPV, but not significantly (pe"0.05) on WBC, Lymphocytes, Monocytes, Eosinophils, RBC, HGB HCT MCH, MCHC, RDW, Neutrophils, PLT. The administration of polyherbal usada at doses of 400 and 500 mg/kg bw resulted in a significantly higher MCV value (pd"0.05) compared to the control, while the doses 100; 200, and 300mg/kg bw resulted in a significant lower PDW (pd"0.05) compared to the control. Dose 300; 400, and 500mg/kg bw resulted in significantly lower basophils (pd"0.05). compared to controls, and doses of 100, and 200mg/kg bw also resulted in significantly lower MPV (p<0.05) (Table 2).

DISCUSSION

The administration of polyherbal usadha resulted in an average lower ALT, and AST than the control, and even significantly lower ALT than the control. This condition is due to the influence of the active ingredients contained in polyherbal usadha, namely pepper and lime. The active compounds of pepper (Piper nigrum L) are mostly in the form of alkaloid compounds, including Piperidine, 1-[5-(1,3-benzodioxol-5-yl)-1-oxo-2,4-pentadienyl]-, (Z,Z)-; and Caryophyllene.¹⁹ Meanwhile, lime leaves (Citrus *amblycarpa*) have also been known to have many active compounds, most of which are essential oils, including Citronellol; Caryophyllene; Hexadecanoic acid, ethyl ester; 1-heptatriacotanol; Phytol; Ethyl 9,12,15-octadecatrienoate; Methyl glycolate,3TMS derivative; 3,7-dimethyl oct-6-en-1-yl stearate; Methyl iso-allocholate; Rhodopine; and Tricyclo [20.8.0.0(7,16) triacontane, 1(22), 7(16)-diepoxy-.³⁰ Caryophyllene has significant pharmacological benefits, including antioxidant, anticancer, cardioprotective, antiinflammatory, hepatoprotective, nephroprotective, gastroprotective, antimicrobial, and immunomodulatory properties.³¹ These active ingredients have a good effect on liver and kidney function so that polyherbal usadha does not cause hepatocyte damage.

Hepatotoxicity is a side effect that often occurs in treatment. This happens because the liver functions as a center for the disposition of drug metabolism and foreign substances in the body.

Dose (mg/ kg bw)	AST(U/L)	ALT(U/L)	Parameter ALP(U/L)	BUN (mg/dl)	SC (mg/dl)	TP (mg/dl)
0	160.75±27.55	85.75±9.73	300.50±62.98	36.95±9.05	0.95±0.02	5.25±0.02
100	150.00±23.98	66.25±5.91 *	215.00±29.90	34.30±5.41	0.63±0.11	5.13±0.21
200	97.75±7.28	61.25±5.68 *	266.25±46.38	32.05±5.93	0.88 ± 0.06	4.85±0.18
300	98.40±7.98	61.72±3.84 *	378.88±31.40	27.29±3.04	0.82 ± 0.07	5.03±0.16
400	107.82±12.06	64.08±2.11 *	460.90±36.40 *	32.53±6.11	0.92 ± 0.01	4.90±0.10
500	108.33±9.63	54.95±0.81 *	341.75±22.31	21.79±3.45	0.85 ± 0.06	5.01±0.13

Table 1. Blood Biochemical Levels of Wistar Rats After Administration of Polyherbal Usadha (Mean ±SE mean)

*Significant with control at 0.05 level. ALT (Alanine aminotransferase), AST (Aspartate aminotransferase), ALP (Alkaline phosphatase), SC (Serum creatinine), BUN (Blood Urea Nitrogen), TP (Total protein (TP)

Parameter	Control	100	Dose mg/kg bw 200	300	400	500	Normal values
WBC (10 ³ /uL) Neut (10 ³ /uL)	11.80 ± 1.31	12.60 ± 1.6 2 04 + 0 47	12.13 ± 0.74 3.28 ± 0.76	10.99 ± 0.82 4.08 ± 0.60	10.56 ± 1.00 3 80 + 1.05	10.80 ± 0.85 3 35 + 0 71	4.80 - 17.50
Lymph (10 ³ /uL) >	7.51 ± 1.01	8.04 ± 1.31	7.37 ± 0.58	5.61 ± 0.23	5.42 ± 0.80	6.13 ± 0.20	1.00 - 3.70
Mono $(10^3/uL)$	0.81 ± 0.10	0.93 ± 0.17	0.85 ± 0.06	1.11 ± 0.04	1.08 ± 0.14	0.87 ± 0.04	0.00 - 0.70
Eo $(10^{3}/uL)$	0.34 ± 0.13	0.42 ± 0.08	0.42 ± 0.13	0.22 ± 0.04	0.16 ± 0.07	0.34 ± 0.08	0.00 - 0.40
Baso $(10^3/uL)$	0.55 ± 0.12	0.27 ± 0.16	0.22 ± 0.21	$0.00 \pm 0.00 *$	0.00 ± 0.00 *	$0.11\pm0.06*$	0.00 - 0.10
RBC (10 ⁶ /uL) >	7.65 ± 0.19	7.72 ± 0.17	7.72 ± 0.09	7.35 ± 0.24	6.93 ± 0.27	7.56 ± 0.21	3.90 - 6.10
HGB (g/dl)	13.45 ± 0.23	13.65 ± 0.50	13.45 ± 0.27	13.09 ± 0.42	12.50 ± 0.54	13.70 ± 0.31	11.1 - 18.0
HCT (%)	40.25 ± 0.79	41.38 ± 1.00	41.03 ± 0.32	38.85 ± 0.89	38.12 ± 1.45	41.74 ± 0.74	31.0 - 52.0
MCV (fL) <	52.68 ± 0.60	53.60 ± 0.40	53.13 ± 0.31	52.87 ± 0.60	$54.99 \pm 0.36 *$	$55.24 \pm 0.68 *$	86.0 - 110.0
MCH (pg) <	17.63 ± 0.20	17.65 ± 0.31	17.42 ± 0.30	17.79 ± 0.18	18.02 ± 0.17	18.13 ± 0.21	26.0 - 38.0
MCHC (g/dl)	33.43 ± 0.20	32.95 ± 0.41	32.78 ± 0.50	33.63 ± 0.34	32.77 ± 0.39	32.86 ± 0.25	31.0 - 37.0
RDW_SD (fL)	33.93 ± 2.40	28.95 ± 0.10	29.83 ± 0.49	32.56 ± 1.42	34.31 ± 3.13	32.41 ± 1.46	·
RDW_CV (%)	20.85 ± 0.94	18.25 ± 0.25	18.90 ± 0.09	20.09 ± 0.42	19.86 ± 0.88	19.28 ± 0.28	11.0 - 16.0
$PLT (10^{6}/uL) >$	701.25 ± 170.42	946.25 ± 60.98	956.00 ± 168.27	1066.08 ± 68.13	1201.58 ± 234.00	$1077.33 \pm 118,77$	150 - 450
MPV(fL) <	7.80 ± 0.13	7.35 ± 0.12 *	7.38 ± 0.13 *	7.46 ± 0.08	7.55 ± 0.18	7.90 ± 0.13	9.0 - 13.0
PCT (%)	0.54 ± 0.13	0.70 ± 0.03	0.70 ± 0.11	0.80 ± 0.04	0.90 ± 0.17	0.85 ± 0.10	0.17 - 0.35
PDW (fL)	9.20 ± 0.11	$8.35 \pm 0.25 *$	$8.35 \pm 0.15 *$	$8.61 \pm 0.11 *$	8.91 ± 0.26	9.23 ± 0.15	9.0 - 17.0
P_LCR (%)	11.23 ± 1.49	7.23 ± 0.40	7.48 ± 0.77	7.56 ± 0.74	8.50 ± 1.35	10.26 ± 0.81	ı
* Significant with contrc volume), MCH (Mean ce I vmn (I vmnhovvte) Ne	al at 0.05 level; > excee Il hemoglobin), MCHC Montroubils) Mono	eding normal value, < (mean cell hemoglobi (Monocytes): PTT (PI	less than normal value. n concentration), RDW (atelets) DDW (Platelet of	Note: RBC (Red Bloo (Red cell distribution w fictribution width) MI	d Cell), HGB (Hemogle vidth); WBC (white bloo 2V (Mean nlatelet volum	bin), HCT (Hematocrit d cells), Eo (eosinophils), MCV (mean cell), Baso (basophils)

ARSANA et al., Biomed. & Pharmacol. J, Vol. 18(1), 899-908 (2025)

Drugs given orally to pass through the intestinal cell membrane must be fat-soluble, then taken to the liver to be converted into water-soluble (more polar), then excreted in urine (if the molecule is small), or through bile (if the molecule is large). Liver lesions can occur due to hepatocellular reactions through the production of enzyme-drug complexes. This complex will then cause cell dysfunction, membrane dysfunction, and, cytotoxic response of T cells.³² A similar study using a mixture of Asparagus racemosus, Cajanus cajan, Cassia fistula, and Carissa spinarum, was able to protect the liver from damage induced by carbon tetrachloride (CCl₄).³³ Other studies also found that Piper nigrum stems provided protection against hepatotoxicity and dyslipidemia in tenofovir/ lamivudine/efavirenz-induced wistar rats.34

The roles of ALT and AST as markers of hepatocellular damage.34-36 Increased levels of ALT and AST in the bloodstream are generally considered a sign of liver tissue damage ³⁷; the higher the level in the blood, the higher the liver damage that occurs 38. ALT and AST are cytosolic enzymes involved in gluconeogenesis. In this process, glucose is synthesized from the amino acids alanine and aspartate and then exported to the blood circulation.³⁹ AST is found primarily in the mitochondria and cytosol of hepatocytes. Still, it can also be found in the heart muscle and skeletal muscle, kidneys, brain, pancreas, lungs, leukocytes, and erythrocytes. Nevertheless, ALT is more specific to the liver, being found in much higher concentrations compared to other tissues. Thus, it can be said that the administration of polyherbal usadha does not cause damage to hepatocytes.

An increase in ALP, while liver test results are normal, may indicate increased osteoblastic activity or tissue regeneration, but requires additional testing for certainty. ALP enzymes are found in the liver, bones, and placenta. ALP is an enzyme that plays a role in accelerating the hydrolysis of organic phosphate by releasing inorganic phosphates. ALP enzyme runoff in the blood can be used as a marker of cell damage.^{40,41} Lime and pepper mint can protect the kidneys so that they do not cause problems with kidney function. Caryophyllene with high concentrations in polyherbal ^{19,30}; is known to have significant pharmacological benefits, including antioxidant, anticancer, cardioprotective, anti-inflammatory, hepatoprotective, nephroprotective, gastroprotective, antimicrobial, and immunomodulatory properties.³¹

The presence of SC, BUN, and TP in the blood is an important indicator of kidney dysfunction. Creatinine is the end product of creatine metabolism excreted through the kidneys.⁴² Meanwhile, increased BUN indicates kidney failure in carrying out its filtration function. Meanwhile, total protein plays a role in determining the diseases that cause protein production disorders. Thus, it can be said that the administration of polyherbal *usadha* does not cause changes in the liver and kidney function of Wistar rats, so it is relatively safe when used as a medicinal ingredient up to a certain dose.

The results showed that the RBC value at all doses was higher than the normal value. This suggests the potential effects of polyherbal usadha supplementation on red blood cell production but does not significantly affect hemoglobin concentrations. This suggests that an increase in RBC is not accompanied by changes in red blood cell quality or excessive oxygenation capacity. Other erythrocyte parameters (MCV, MCH) were generally below normal values at low doses, but MCV showed significant differences compared to controls after polyherbal supplementation at doses of 400 and 500mg/kg bw. A slightly lower MCV value after low-dose polyherbal supplementation could indicate that at low doses, this supplement has the potential to lower red blood cell size, but this effect does not lead to anemia or iron deficiency, given that MCH and HGB remain stable.43

Red blood cell production is a complex and multi-layered process that is primarily regulated by erythropoietin (Epo). The main physiological regulator of EPO production is renal hypoxia or hypoxemia which both reduce the oxygen-carrying capacity of the blood. EPO gene expression is controlled by a transcription factor known as hypoxia-inducible factor (HIF) which plays an important role in cellular adaptation to decreased oxygen levels. Once released from the kidneys, the EPO circulates to the hematopoietic marrow which binds to and activates its receptors (EPOR). EPOR in turn phosphorylates and activates the signal transduction of Janus kinase-2 (JAK-2), and Signal Transducer and Activator of Transcription-5 (STAT-5) in erythroid progenitors, as well as phosphatidylinositol-3 (PI-3) kinase. Signals transduced through JAK-2/STAT-5 and PI-3 kinase cause changes in gene expression that promote effective erythropoiesis by increasing proliferation and reducing red blood cell progenitor apoptosis.⁴⁴

The mean WBC count remained within the normal range (4.80–17.50 10³/uL) at all doses, with no significant changes indicating serious adverse effects on the immune system. The number of neutrophils increases with doses up to 300 mg/ kg but is still within normal limits. In contrast, lymphocytes show higher values than normal, but show a decrease at higher doses, especially at 300-500 mg/kg. An increase in neutrophils indicates a mild immune response, while a decrease in lymphocytes at high doses signals an adjustment of the immune system to polyherbal components without any indication of serious immune damage or suppression.⁴⁵

Platelet counts at all doses of supplementation, and especially at high doses reaching 1201.58 10v /uL, well above the normal range (150-450 10v /uL). This response may be caused by polyherbal components that promote platelet formation. MPV and PDW tended to decrease at low doses compared to the control group, suggesting specific effects on platelet size and distribution at low doses of this supplement. At low doses, it suggests that polyherbal may affect the average size of platelets, but this is not accompanied by changes that compromise platelet function. This condition is likely the result of infection, chronic inflammation, or drug reactions, which are the most common causes of thrombocytosis.46,47

Overall, polyherbal *usadha* supplementation exerted a significant influence on several hematological parameters in Wistar rats, specifically on the increase in platelets and RBCs without any visible hematological side effects. This supplementation appears to be able to support the hematopoiesis process, which is potentially beneficial in certain conditions that require an increase in platelet count or erythrocytes. However, given the fairly high increase in platelets, more research is needed to monitor the potential risk of hypercoagulability or increased blood clots. This study provides a solid basis for further studies regarding the benefits and risks of using polyherbal *usadha* in clinical applications.

CONCLUSION

Polyherbal usadha supplementation in Wistar rats affects several hematological and biochemical parameters. The main effects appear to be immune system stimulation and platelet formation. In addition, the positive impact of ALT shows potential benefits in supporting liver function. However, the increase in ALP at high doses should be looked at further, as it could indicate the potential for liver stress at higher doses. Polyherbal usadha appears consumable without the risk of significant side effects on blood parameters and liver function. Additional studies that include histological analysis and mechanisms of action will provide deeper insights into the therapeutic effects of Polyherbal usadha.

ACKNOWLEDGEMENT

The author would like to thank the Hindu University of Indonesia (UNHI) for supporting the implementation of this research. The author would also like to thank the Study Program of Biology, Faculty of Health Sciences, Universitas Dhyana Pura (UNDHIRA-BALI) for helping to implement this research and writing this manuscript.

Funding Source

This research was funded by the Institute for Research and Community Service (LPPM) of the Hindu University of Indonesia (UNHI) through the 2024 Research Funding Grant Program with Contract Number: 001.PA/KPEN-LPPM/UNHI/ VIII/2024

Conflict of Interest

The author(s) do not have any conflict of interest.

Data Availability Statement

This statement does not apply to this article.

Ethics Statement

The use of experimental animals in this study has received ethical approval from the animal ethics committee of the Faculty of Veterinary Medicine, Udayana University (UNUD) with Number: B/110/UN14.2.9/PT.01.04/2024.

Informed Consent Statement

This study did not involve human participants, and therefore, informed consent was not required.

Clinical Trial Registration

This research does not involve any clinical trials.

Permission to reproduce material from other sources

Not Applicable.

Author Contributions

I Nyoman Arsana: Conceptualization, Methodology, Data collection, Data analysis, Project administration, and Funding Acquisition; Ni Ketut Ayu Juliasih and Anak Agung Ayu Sauca Sunia Widyantari: Data collection, Data analysis, and Project administration; I Gede Widhiantara and Putu Angga Wiradana: Writing – Original Draft, Editing, Supervision, and Visualization.

REFERENCES

- 1. Jatoth R, Dhanabal S., Senthil V, et al. Acute oral toxicity study of novel polyherbal formulations by using wistar rats and Swiss albino mice as per OECD 425 TG. *Phytomedicine Plus.*, 2025; 5(1):1-11.
- World Health Organization (WHO). WHO global report on traditional and complementary medicine 2019. Global report. 1-226.
- 3. Dias PGI, Marapana RAUJ, Rathnayaka RMUSK, et al. Identification of the best plant ratios for a polyherbal tea mix to obtain optimum antioxidant, antidiabetic, and â-glucuronidase inhibition activities. *J Ayurveda Integr Med.* 2024;15(5):1-7.
- 4. Budiarti M, Maruzy A, Mujahid R, et al. The use of antimalarial plants as traditional treatment in Papua Island, Indonesia. *Heliyon*. 2020;6(12):1-10.
- Illian DN, Siregar ES, Sumaiyah S, et al. Potential compounds from several Indonesian plants to prevent SARS-CoV-2 infection: A mini-review of SARS-CoV-2 therapeutic targets. *Heliyon*. 2021;7(1):1-11.
- Fitmawati F, Sofiyanti N, Roza RM, et al. Short Communication: Traditional medicinal formulation: Obat pahit from Lingga Malay Ethnic in Riau Archipelago, Indonesia. *Biodiversitas J Biol Divers*. 2017;18(3):1196– 200.
- Isnawati A, Gitawati R, Raini M, Alegantina S, Setiawaty V. Indonesia basic health survey: selfmedication profile for diarrhea with traditional

medicine. Afr Health Sci. 2019;19(3):2365-2371.

- Pengpid S, Peltzer K. Utilization of traditional and complementary medicine in Indonesia: Results of a national survey in 2014–15. Complement Ther Clin Pract. 2018; 33:156–163.
- 9. Jadid N, Kurniawan E, Himayani CES, et al. An ethnobotanical study of medicinal plants used by the Tengger tribe in Ngadisari village, Indonesia. *PLoS One.* 2020;15(7):1-16.
- Supiandi MI, Mahanal S, Zubaidah S, Julung H, Ege B. Ethnobotany of traditional medicinal plants used by Dayak Desa Community in Sintang, West Kalimantan, Indonesia. *Biodiversitas J Biol Divers*. 2019;20(5): 1264-1270.
- Elfrida E, Tarigan NS, Suwardi AB. Ethnobotanical study of medicinal plants used by community in Jambur Labu Village, East Aceh, Indonesia. *Biodiversitas J Biol Divers*. 2021;22(7): 2893-2900.
- 12. Elfahmi, Woerdenbag HJ, Kayser O. Jamu: Indonesian traditional herbal medicine towards rational phytopharmacological use. *J Herb Med*. 2014;4(2):51–73.
- Jun P, Rahmat E, Han C-H, Yang C, Kang Y. Traditional Chinese Medicine and Traditional Indonesian Medicine: A Comparative Review of Herbal Medicines Restricted in Pregnancy. *Chin J Integr Med.* 2021;27(10):794–800.
- Saad B, Zaid H, Shanak S, Kadan S. Hypoglycemic and Anti-obesity Polyherbal Mixtures. In: Antidiabetes and Anti-obesity Medicinal Plants and Phytochemicals. Cham: *Springer International Publishing*; 2017. p. 217–51.
- 15. Liyanagamage DSNK, Jayasinghe S, Attanayake AP, Karunaratne V. Acute and Subchronic Toxicity Profile of a Polyherbal Drug Used in Sri Lankan Traditional Medicine. *Evidence-Based Complement Altern Med.* 2020; 13:1-12.
- Ahmad A, Abuzinadah M, Alkreathy H, et al. A novel polyherbal formulation containing thymoquinone attenuates carbon tetrachlorideinduced hepatorenal injury in a rat model. *Asian Pac J Trop Biomed.* 2020;10(4):147-155.
- Gundeti MS, Bhurke LW, Mundada PS, et al. AYUSH 64, a polyherbal Ayurvedic formulation in Influenza-like illness - Results of a pilot study. *J Ayurveda Integr Med.* 2022;13(1):1-7.
- Aslam B, Hussain A. Phytochemical Characterization and Solvent Fraction Depending in vitro Antioxidant Activities of *Cassia absus*, Gymnema sylvestre, *Nigella sativa* and *Piper nigrum. Rev Chim.* 2021;72(2):38–49.
- Ukkinadka J, Badanthadka M. Safety evaluation of a proprietary ayruveda-based polyherbal preparation (arthralgex) used for arthritis.

Brazilian J Biol. 2024;84.

- 20. Arsana IN, Juliasih NKA, Widyantari AAASS, Suriani NL, Manto A. GC-MS Analysis of the Active Compound in Ethanol Extracts of White Pepper (*Piper nigrum* L.) and Pharmacological Effects. *Cell Mol Biomed Rep.* 2022;2(3):151– 61.
- Chen Y, Guo D, Deng H, et al. Acute and chronic toxicity of a polyherbal preparation – Jueyin granules. *BMC Complement Altern Med*. 2018;18(1):1-13.
- Saleem U, Amin S, Ahmad B, et al. Acute oral toxicity evaluation of aqueous ethanolic extract of *Saccharum munja* Roxb. roots in albino mice as per OECD 425 TG. *Toxicol Reports*. 2017; 4:580-585.
- Majumdar A, Shukla SS, Pandey RK. In-vitro and in-vivo Immunomodulatory Effect of Polyherbal Suspension on Cyclophosphamide Induced Experimental Animal. *Indian J Pharm Educ Res.* 2021;55(1s): s225–232.
- Andila PS, Tirta IG, Warseno T, Sutomo S. Medicinal Plants Diversity Used by Balinese in Buleleng Regency, Bali. J Trop Biodivers Biotechnol. 2023;8(1):1-18.
- 25. Warditiani N, Leliqia N, Savitri P. Plant Data and Treatment on Rare Usada Palmyra. *J Farm Udayana*. 2015;4(1): 29-32.
- Mu'jizah. Usada Manuscripts as Local Wisdom of Balinese Society. *Dialektika*. 2016;3(2):191– 200.
- 27. Mohotti S, Rajendran S, Muhammad T, et al. Screening for bioactive secondary metabolites in Sri Lankan medicinal plants by microfractionation and targeted isolation of antimicrobial flavonoids from Derris scandens. *J Ethnopharmacol.* 2020;10(1): 1-13.
- Ramasamy M, Karthikeyan E, Srinivasan S, Navaneetha Krishnan S. Investigation of the effects of polyherbal formulations against the combination of a high-fat cafeteria diet along with exercise modalities. *Adv Biomark Sci Technol.* 2024; 6:35–45.
- 29. Bemidinezhad A, Zojaji SA, Taraz JS, Mohammadi M, Alavi MS, Ghorbani A. Evaluation of acute, subacute, and subchronic toxicity of a hepatoprotective herbal formulation. *Toxicol Reports*. 2023; 11:452–459.
- 30. Brígido HPC, Varela ELP, Gomes ARQ, et al. Evaluation of acute and subacute toxicity of ethanolic extract and fraction of alkaloids from bark of *Aspidosperma nitidum* in mice. *Sci Rep.* 2021;11(1):1-14.
- Arsana IN, Juliasih NKA, Widyantari AAASS. GC MS Analysis of Bioactive Compounds in Lime Leaf Ethanol Extract (Citrus amblycarpa

(Hassk.) Ochse), and Its Potential as a Traditional Medicine Agents. *J Penelit Pendidik IPA*. 2024;10(4):1994–2006.

- 32. Machado Kd.C, Islam MT, Ali ES, et al. A systematic review on the neuroprotective perspectives of beta-caryophyllene. *Phytother Res.* 2018; 32(12): 2376–2388.
- 33. Rasyid SA, Armayani, Yuniati, Lio TMP. Analysis of serum glutamic pyruvic transaminase and serum glutamic oxaloacetic transaminase levels in tuberculosis patients who are undergoing oat treatment in Kendari City General Hospital, Kota Kendari, Indonesia. *Infect Dis Rep.* 2020;12(S1): 75-77.
- Akhter S, Jahan I, Roy DC, et al. Combined hepatoprotective potentials of medicinal plants on CCl₄-induced hepatotoxic Wistar rats. *Indian J Tradit Knowl*. 2024;23(5):433–440.
- 35. Enyang D, Sonibare MA, Tchamgoue AD, et al. Protective and Ameliorative Effects of Hydroethanolic Extract of *Piper nigrum* (L.) Stem against Antiretroviral Therapy-Induced Hepatotoxicity and Dyslipidemia in Wistar Rats. *J Toxicol*. 2024: 1-16.
- Domanski JP. The AST to ALT ratio: A pattern worth considering. *Curr Hepat Rep.* 2013;12(1):47–52.
- Dollah MA. Toxicity effect of *Phaleria* macrocarpa (Mahkota dewa) on liver function in sprague dawley rats. *Trop J Nat Prod Res.* 2021;5(2):304–308.
- Alshuweishi Y, Alfaifi M, Almoghrabi Y, Alfhili MA. AST and ALT APRI Scores and Dysglycemia in Saudi Arabia: A Retrospective Population Study. *Life*. 2023;13(9):1–14.
- Maryam S, Arsani NLKA, Tangguda S. Grape (Vitis vinifera L.) skin extract reduced levels of SGPT and SGOT and improved the liver tissue structure of Wistar rats (Rattus novergicus) fed a high-cholesterol diet. Bali Med J. 2022;11(3):1404–1409.
- Han HS, Kang G, Kim JS, Choi BH, Koo SH. Regulation of glucose metabolism from a livercentric perspective. *Exp Mol Med.* 2016;48(3):1– 10.
- Ogunmoyole T, Johnson OD, Yusuff AA. Ethanolic Extract of Whole Unripe Plantain *Musa* paradisiaca Ameliorates Carbon Tetrachloride-Induced Hepatotoxicity and Nephrotoxicity in Wistar Rat. *Annual Research & Review in Biology*. Sciencedomain International; 2021. p. 78–87.
- Makris K, Mousa C, Cavalier E. Alkaline Phosphatases: Biochemistry, Functions, and Measurement. *Calcif Tissue Int*. 2023;112(2):233– 242.

- 43. Syukriah S. The Effect of Dexametasone Administration on Alp and Creatinine Levels of Wistar Strain Rats (*Rattus norvegicus*). *In*: Prosiding Seminar Nasional Biotik. 2017. p. 331–335.
- 44. Mithoowani S. Investigation and management of erythrocytosis. Vol. 192, *CMAJ*. 2016; 11: 342-347.
- 45. Tome J, Debeljak N. Molecular insights into the oxygen sensing pathway and erythropoietin

expression regulation in erythropoiesis. *Int J Mol Sci.* 2021;22(13): 1-16.

- Susman D. Lymphocytosis with Smudge Cells Is Not Equivalent to Chronic Lymphocytic Leukemia. Case Rep Oncol. 2021;14(2):950– 956.
- Vo QT, Dennis F. A Review and Assessment of Drug-Induced Thrombocytosis. Vol. 53, *Annals* of *Pharmacotherapy*. 2019. p. 523–536.

908