

## Probiotics for Control of Nosocomial Infections

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### ABSTRACT

Probiotics are usually defined as live microbial food ingredients beneficial to health which comprise of normal commensally bacteria as a part of the healthy human gut microflora. Different species of microorganisms such as lactic acid bacteria (*Bifidobacterium* and *Lactobacillus* spp.) or yeasts have been proposed for human use. Nosocomial infections occur worldwide and affect both developed and poor countries. The most common organisms causing nosocomial infection in neonates include *Staphylococcus*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Klebsiella*, and *Candida*. There is preliminary evidence that probiotic type microorganisms may control the growth of nosocomial pathogens. The aim of this review is to consider the current evidence on the effects of probiotics for control of nosocomial infections.

**Key words:** Probiotics, nosocomial infections, safety, Infection control.

### INTRODUCTION

Probiotics can be defined as nonpathogenic microorganisms that, when ingested, exert a positive influence on the health of the host. They consist of either yeast or bacteria, especially lactic acid bacteria. The most commonly used organisms in probiotic products belongs to *Lactobacillus* and *Bifidobacterium* spp., Other organisms have also been used including *Bacillus* spp., and yeast such as *Saccharomyces boulardii* (Darsanaki and Aliabadi, 2013). In Table 1 are listed some of the known probiotics available (Mombelli and Gismondo, 2000).

Probiotics can produce inhibitory compounds such as lactic acid, bacteriocin, hydrogen peroxide, acetaldehyde and diacetyl. These compounds are able to inhibit the growth of pathogenic microorganisms (Zapata, 2013). Probiotics could be used for several conditions such as Urinary Tract Infections, Irritable Bowel Syndrome, Immune Disorders, Lactose Intolerance, Hyper Cholesterolaemia, Inflammatory

Bowel Disease, Cancer, Allergy and Diarrhea (Zerehpooch and darsanaki, 2013). Nosocomial infections occur worldwide and affect both poor and developed countries. Infections acquired in health care settings are among the major causes of death and increased morbidity among hospitalized patients. They are a significant burden both for the patient and for public health (WHO, 2002). Over the past 50 years, the epidemiology of pathogens responsible for nosocomial infections in neonates has changed dramatically. During the 1950s, *Staphylococcus aureus* was the most common nosocomial pathogen in hospitalized infants. During the 1960s, gram-negative bacilli, including *Pseudomonas aeruginosa*, *Klebsiella* sp, and *Escherichia coli*, became the most common pathogens. By the 1970s, coagulase negative staphylococci (CONS) (*S. epidermidis*) and *S. aureus*, including methicillin resistant *S. aureus* (MRSA), became the predominant causes of hospital-acquired infections in the neonatal intensive care unit. Today, gram-positive cocci continue to cause the largest proportion of infections, and many, including CONS, MRSA, and

vancomycin-resistant enterococci (VRE), are multidrug-resistant. Gram-negative bacilli are responsible for 20% to 30% of cases of late-onset sepsis and 30% of nosocomial pneumonias (Table 2) (Polin and Saiman, 2003).

There is preliminary evidence that probiotic type microorganisms may control the growth of nosocomial pathogens (Falagas and Makris, 2009). The aim of this review is to consider the current evidence on the effects of probiotics for control of nosocomial infections.

#### Control of Nosocomial Infections by Probiotics

Due to concerns regarding development of antibiotic resistance in hospitals, rising healthcare costs and lack of new antimicrobial classes being developed, probiotics have been considered a good prophylactic or therapeutic alternative in numerous

conditions. Probiotics do not have the risk of antimicrobial resistance and offer practical benefits like low-cost preparation, long shelf life, and ease of administration (Oudhuis *et al.*, 2011). We identified 29 studies involving in-vitro experiments on the potential role of probiotics in the inhibition of nosocomial pathogens (bacterial or fungal) (Table 3).

#### CONCLUSION

However, there are still many questions to be answered before probiotics can be used routinely as therapy. At present, with increasing of the antibiotic resistance and side effects of chemical drugs, it seems, we need to use alternative remedies. Probiotics can have therapeutic application in future.

**Table 1: Common probiotics for human use**

<b>Lactobacillus species</b>	<b>Bifidobacterium species</b>	<b>Other bacteria</b>	<b>Non-lactic acid producing bacteria</b>
<i>L. acidophilus</i>	<i>B. lactis</i>	<i>L. lactis</i>	<i>S. cerevisiae</i>
<i>L. casei</i>	<i>B. animalis</i>	<i>E. faecium</i>	<i>B. subtilis</i>
<i>L. crispatus</i>	<i>B. bifidum</i>	<i>E. faecalis</i>	<i>S. boulardii</i>
<i>L. gasseri</i>	<i>B. infantis</i>	<i>P. acidilactici</i>	<i>B. cereus</i>
<i>L. johnsonii</i>	<i>B. adolescentis</i>	-	-
<i>L. reuteri</i>	-	-	-
<i>L. rhamnosus</i>	-	-	-

**Table 2: Common Nosocomial Infections**

<b>Common Pathogens</b>	<b>Less Common Pathogens</b>	<b>Site of Infection</b>
<i>Pseudomonas aeruginosa</i> <i>Staphylococcus aureus</i> <i>Candida</i> sp. Coagulase negative staphylococci	<i>Enterococci</i> <i>Klebsiella</i> sp. <i>Serratia marcescens</i> <i>Enterobacter</i> sp. <i>Malassezia</i> sp.	Bloodstream/sepsis
Gram-negative bacilli <i>Enterococci</i> <i>S. aureus</i> Coagulase negative staphylococci	<i>Candida</i> sp. <i>Enterococci</i> <i>S. marcescens</i> <i>Aspergillus</i> sp.	Urinary tract Skin/soft tissue/ surgical site
<i>S. aureus</i> <i>P. aeruginosa</i> Coagulase negative staphylococci Respiratory syncytial virus	<i>Enterococci</i> <i>Klebsiella</i> sp. <i>S. marcescens</i>	Pneumonia
<i>S. aureus</i> Coagulase negative staphylococci Coagulase negative staphylococci <i>S. aureus</i>	<i>Candida</i> sp. <i>S. marcescens</i> <i>Enterobacter</i> sp. <i>Candida</i> sp.	Endocarditis Central nervous system

Table 3: In-vitro studies regarding the effect of probiotics in the inhibition of nosocomial pathogens

Probiotic used	Isolated from	Nosocomial pathogens	Region	Reference
<i>L. plantarum</i> , <i>L. delbrueckii</i> , <i>L. acidophilus</i> , <i>L. brevis</i> , <i>L. casei</i> <i>Lactobacillus</i> spp <i>Bifidobacterium</i> spp <i>L. casei L. bulgaricus</i>	Sausages	<i>S. aureus</i>	Iran	Nowroozi et al., 2004
<i>L. plantarum</i> , <i>L. casei</i> subsp. <i>Paracasei</i> , <i>L. casei</i> <i>Lactobacillus</i> spp.	Probiotic Milk	<i>S. aureus</i> ATCC 25922S <i>S. pneumoniae</i> ATCC 41619 <i>P. aeruginosa</i> , <i>S. aureus</i> <i>Klebsiella</i> , <i>Enterobacter</i> <i>P. aeruginosa</i> , <i>S. aureus</i>	USA	Rosario et al., 2005
<i>L. acidophilus</i> , <i>B. animalis</i> <i>L. paracasei</i> subsp. <i>paracasei</i> , <i>Propionibacterium</i> <i>freudenreichii</i> sub sp. <i>shermanii</i>	Various Foods		Turkey	Erdourul and Erbulur, 2006
<i>L. acidophilus</i> , <i>B. bifidum</i>	Standard	<i>S. aureus</i>	Romania	lordache et al., 2008
<i>L. lactis</i> , <i>L. delbrueckii</i>	Yogurt	<i>S. aureus</i> MRSA <i>P. aeruginosa</i> , <i>E. coli</i> <i>S. aureus</i> , <i>E. coli</i>	Bulgaria	Petrova et al., 2009
<i>Lactobacillus</i> , <i>Streptococcus</i> and <i>Bifidobacterium</i> spp <i>L. bulgaricus</i>	Cheese		Australia	Tharmaraj and shah, 2009
<i>Lactobacillus</i> spp.	Probiotic Tablet	<i>S. aureus</i> PTCC 1431 <i>P. aeruginosa</i> ATCC 27853 <i>E. coli</i> PTCC 1399 <i>S. aureus</i> , <i>E. coli</i> , <i>P. aeruginosa</i> .	Iran	Darsanaki et al., 2011
<i>Lactobacillus</i> ,	Traditional Dairy Product	<i>S. aureus</i> , <i>E. coli</i>	Sudan	Salih et al., 2011
<i>Lactobacillus</i> spp.	Yoghurt	<i>E. coli</i> ATCC 10536 <i>S. aureus</i> ATCC 6538 <i>E. coli</i> PTCC1399	Iran	Hami et al., 2011
<i>Lactobacillus</i> ,	Traditional Dairy Products		Pakistan	Maria et al, 2011
	Cabbage, Milk,	<i>E. coli</i> , <i>S. aureus</i>	Iran	Jafari et al, 2011
			United	Tejero et al., 2012

				Kingdom	
<i>Bifidobacterium</i> , <i>Lactococcus</i> and <i>Streptococcus</i> <i>L. Plantarum</i> . <i>L. casei</i> . <i>L. brevis</i> <i>Lactobacillus</i> and <i>Streptococcus</i> spp. <i>L. acidophilus</i> <i>L. casei</i> . <i>L. brevis</i> <i>Lactobacillus</i> and <i>Bifidobacterium</i> spp. <i>Lactobacillus</i> , <i>Bifidobacterium</i> and <i>Propionibacterium</i> sp.	Cheese and Human origin Fresh Vegetables Fermented Toddy Traditional Yoghurt Cheeses Yoghurt, Milk Kishk Sourdough, Vegetables, Yoghurt, Cheese and Human origin Traditional Yoghurt	<i>S. aureus</i> PTCC 1431 <i>E. coli</i> PTCC 1399 <i>Enterobacter</i> <i>E. coli</i> <i>Klebsiella</i> sp. <i>S. aureus</i> <i>E. coli</i> PTCC1399 <i>S. aureus</i> PTCC 1431 <i>S. aureus</i> <i>E. coli</i> <i>E. coli</i> ATCC 25922 <i>E. coli</i> ATCC 8739 <i>S. aureus</i> ATCC 25093 <i>P. aeruginosa</i> ATCC 9027 <i>S. aureus</i> ATCC 6538 <i>Aspergillus</i> <i>P. aeruginosa</i> , <i>S. aureus</i> (MRSA)	Iran India Iran Egypt Bulgaria Iraq	Darsanaki <i>et al.</i> , 2012 Krishnamoorthy and Arjun, 2012 Issazadeh1 <i>et al.</i> , 2013 Ali <i>et al.</i> , 2013 Denkova <i>et al.</i> , 2013 Ahmed, 2013	
<i>L. acidophilus</i> <i>L. bulgaricus</i> , <i>L. casei</i> , <i>L. plantarum</i> <i>Lactobacillus</i> spp.	Yoghurt Milk	<i>E. coli</i> <i>S. aureus</i>	Saudi Arabia India	Abdallah <i>et al.</i> , 2013 Srinu <i>et al.</i> , 2013	
<i>L. fermentum</i> <i>L. rhamnosus</i> <i>L. plantarum</i> ' <i>L. casei</i> . <i>L. delbrueckii</i> subsp. <i>bulgaricus</i> <i>L. bulgaricus</i> . <i>thermophilus</i> . <i>L. lactis</i> <i>L. plantarum</i> <i>L. sakei</i> subsp <i>sakei</i> PTCC 1712 <i>L. casei</i> subsp <i>casei</i> PTCC 1608 <i>L. plantarum</i> subsp <i>plantarum</i> PTCC1745'	Standard Dairy products Grass silage Standard	<i>P. aeruginosa</i> , <i>S. aureus</i> <i>Enterococcus</i> . <i>S. epidermidis</i> <i>E. coli</i> <i>S. aureus</i> Clinical S. aureus PTCC 1431 <i>E. coli</i> Clinical <i>E. coli</i> PTCC 1399	Algeria India Iran	Mezaini and Bouras, 2013 Prema, 2013 Chakoosari <i>et al.</i> , 2014	

<i>L. lactis</i> subsp <i>lactis</i> PTCC 1336	Commercial Yoghurt	<i>E. coli</i> PTCC1399 <i>S. aureus</i> PTCC 1431	Iran	Tajehmiri et al., 2014
<i>L. acidophilus</i> <i>L. Plantarum</i> <i>L. casei</i> <i>L. delbrueckii</i> sub <i>sp. bulgaricus</i>	Traditional Yogurt	<i>E. coli</i> PTCC1399 <i>S. aureus</i> PTCC 1431	Iran	Nasiri Moslem et al., 2014
<i>L. plantarum</i> <i>L. rhamnosus</i> <i>L. casei</i> <i>L. acidophilus</i> <i>L. brevis</i> <i>Lactobacillus</i> spp.	Standard	<i>S. aureus</i> MTCC1144 <i>E. coli</i> 0157:H7	India	Vij et al., 2014
<i>Bifidobacterium</i> . <i>Lactobacillus</i> Spp	Commercial probiotic strains	<i>P. aeruginosa</i> PTCC 1707 <i>S. aureus</i> PTCC 1431 <i>Enterococci</i>	Iran	Darabi et al., 2014
<i>Lactobacillus</i> spp.	Fermented Rice	<i>Candida</i> spp. <i>E. coli</i> ATC 259222	Srilanka	Jeygowri et al., 2014
<i>Lactobacillus</i> spp.	Human origin	<i>S. epidermidis</i> ATCC 12228 <i>E. coli</i> ATCC 29181 <i>S. aureus</i>	Malaysia	Shokryazdan et al., 2014

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