

***In silico* Analysis of Common Autism Spectrum Disorder Genetic Risk Variations**

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Autism spectrum disorder (ASD) is a chronic neurological and developmental disability characterised by inability to develop social relationships, trouble expressing feelings, and repeated behaviours - clinically defined as stereotyped behaviour - that affect how people interact, learn, and behave. Because of the vast range of types and severity of symptoms, it is classified as a "spectrum" disorder. Over the last two decades, the prevalence of ASD has progressively increased, and one out of every 160 children worldwide is estimated to have an ASD. Over 75 percent of ASD patients show psychiatric disorders like depression, stress, bipolar disorder, Tourette syndrome, attention deficit hyperactivity disorder (ADHD). In the present study, in silico analysis was done to identify different rare mutations in genes implicated in ASD. Single nucleotide polymorphisms in ADNP, ARID1B, ASH1L, CHD2, CHD8, DYRK1A, POGZ, SHANK3, and SYNGAP1 genes were identified to be associated with ASD aetiology. A single mutation in these genes can result in defective chromatin remodeling, altering the function of several genes and potentially causing intellectual impairment and autism spectrum disorder (ASD). Understanding and analyzing these SNPs linked to ASD as risk factors can aid in the early detection and diagnosis of the disorder.

Keywords: Autism spectrum disorder; Chromatin remodeling; Histone methylation; Mutation; SNPs.

Autism spectrum disorder is a complex developmental disorder that involves persistent challenges with social interaction, restricted interests, repetitive behaviors, impaired communication ability and deficits in social-emotional reciprocity¹. As per Global Burden of Disease Study (GBD) autism affects 52.9 million children under the age of five². According to WHO 2021 report, the majority of young children suffering from autism live in underdeveloped nations or low and middle-income countries. In South Asia and Sub-Saharan Africa, more than one million children suffer from ASD, and the Middle East, Central Asia, and North Africa have the highest rates of childhood autism³.

Children diagnosed with ASD at the age of two to four years are reported to have an accelerated total brain volume growth in the frontal and temporal region and as they reach the age of ten to fifteen years, the brain's volumetric capacity declines⁴. ASD is referred to as a "spectrum" disorder since there is a wide diversification of its types and severity of symptoms⁵. One of the first indicators of ASD is a delay in language development and children with ASD commonly show impaired language development leading to a lack of social communication¹. Autism is caused by a combination of inherited, nongenetic, and environmental factors, and the majority of them appear to influence key aspects of early brain

development⁶. Some appear to affect the nerve cells interactions with one another in the brain, while others appear to influence how various areas of the brain communicate with one another⁷.

Variations in gene and genomes studies have identified an increasing number of SNPs associated with ASD over the past decade^{8,9}. It has been estimated that SNPs explain between 17 and 60% of ASD, hence their contribution must not be overlooked^{10,11}. The functions of the genetic variants responsible for the association with ASD are, however, poorly understood. A single nucleotide polymorphism (SNP) can lead to translation of the incorrect amino acid, resulting in defective or non-functional proteins. Such mutations in the lysine-specific methyltransferase 2H enzyme have been reported to alter histone methylation and affects brain development, increasing the risk of ASD and other neural disorders^{12,13}. Abnormal regulation of gene expression, disruption of proper neural development, and chromatin deregulation, results in aberrant chromatin remodeling. This interferes with the usual expression of genes essential in brain development, as well as abnormalities in synaptic adaptability and ultimately leads to ASD¹⁴.

In the present study, *in silico* analysis was carried out to identify different genes rare mutations associated with ASD as risk factors. These SNPs possess the potential to serve as important biomarkers for early detection of ASD, monitoring the course of ASD, and to develop possible drug therapies for ASD [Figure 1].

METHODOLOGY

Online bioinformatics tools were used to extract the data. Genetics Home Reference (GHR) database which is a MedlinePlus trusted health information, provided by National Library for Medicine was used to collect data of the genes whose mutations led to autism spectrum disorder which is found under the tab Genetics followed by Genetic Conditions¹⁵. Pathogenic and non-pathogenic mutant alleles associated with ASD were studied using the database retrieved from variation viewer tab under the related information column of National Center for Biotechnology Information (NCBI) site for the said gene which we found out from Genetics Home Reference database¹⁶. Location of the gene, families to

which they belong, and function of the proteins encoded by them were analyzed by MedlinePlus database and documented. NCBI-gene site was used to identify transcript mutation and Rapid Stain Identification Series (rsIDs). The NCBI-Single Nucleotide Polymorphisms (SNP) database was used to examine and identify mutations in codon sequences that resulted in amino acid sequence changes. Variation and phenotype, clinical significance, and effect of the mutated genes on the amino acid sequence was recorded.

RESULT AND DISCUSSIONS

The current investigation discovered that ASD is associated with mutations in the activity dependent neuroprotector homeobox (*ADNP*), AT-rich interaction domain 1B (*ARID1B*), *ASH1* like histone lysine methyltransferase (*ASH1L*), chromodomain helicase DNA binding protein 2 (*CHD2*), chromodomain helicase DNA binding protein 8 (*CHD8*), dual specificity tyrosine phosphorylation regulated kinase 1A (*DYRK1A*), pogo transposable element derived with ZNF domain (*POGZ*), SH3 and multiple ankyrin repeat domains 3 (*SHANK3*), and synaptic Ras GTPase activating protein 1 (*SYNGAP1*) genes (Table 1).

Activity dependent neuroprotector homeobox (*ADNP*)

ADNP gene produces a neuro-protective peptide that operates during early embryogenesis, particularly during neurulation, and promotes glia-derived, survival-promoting chemicals that protect impaired nerve cells from cell death^{17, 18}. *ADNP* gene is located on Chromosome 20 band 13's long arm (q) and the encoded protein controls the expression of genes like *BRG1* and *CHD4*, which are involved in normal brain development through chromatin remodeling¹⁹. The substantial neuroprotective activity of *ADNP* protein can be attributed entirely to the NAP domain, an octapeptide Asn-Ala-Pro-Val-Ser-Ile-Pro-Ala or NAPVSPIQ²⁰. *ADNP* protein binds to DNA and interacts with SWI/SNF complexes by connecting the C-terminus to three of its key components throughout the remodeling process, which affect the shape of chromatin directly^{21,22}. The majority of *ADNP* variants leads to the formation of short *ADNP* protein that can bind to DNA but not

interact with SWI/SNF complexes. Variations at the terminus of a protein are typically unlikely to influence protein function²³. Decreased ADNP expression in haploinsufficient populations has been shown to effectively deregulate the feedback loop by blocking wild-type protein from interacting to the promoter binding domain or occupying alternate target sequences²³. These changes likely explain the intellectual disability in case of ASD. Around 24 mutations (small intragenic deletions, insertions, missense, and splice site variations) were found in *ADNP* gene. Normal coding amino acids like tyrosine and leucine are mutated to terminating codons by frameshift mutations in *ADNP* gene, which may result in the creation of shortened proteins and disordered chromatin remodeling. Activation of many genes, as well as the development and function of several human tissues and organs, including the brain are all affected by chromatin remodeling disruptions. Furthermore, some of these mutations may prevent the production of neurotransmitters like epinephrine and norepinephrine, which are important for neural communication and mood regulation by encoding different amino acids. For example, a mutation of asparagine which is required for neurotransmitter production to lysine.

AT-rich interaction domain 1B (*ARID1B*)

ARID1B plays a pivotal role in controlling gene activity by forming a DNA-binding protein of the Brahma-associated factor chromatin remodeling complexes of numerous distinct SWI/SNF protein complexes which modulate gene activity by chromatin remodeling^{24, 25}. *ARID1B* contains over 2,000 amino acid residues yet it has only two characterized protein domains, an AT-rich interaction domain (ARID) and domain of undefined function 3518 (DUF3518)²². *ARID1B*'s DNA-binding ability is impaired, and the BAF complex's function is jeopardized as a result of missense mutations in the ARID domain²⁶. In BAF complexes, the DUF3518 domain has been shown to interact with the BRG1 and BRM helicase subunits²⁴. Because the *ARID1B* component may bind to DNA, it is thought to contribute in the targeting of SWI/SNF complexes to the chromatin ought to be remodeled²⁴. Missense mutations in any of these domains can cause BAF complexes to become non-functional. Nonsense-mediated mRNA decay (NMD) is primarily triggered

by nonsense and frameshift mutations, which cause premature translation termination²⁷. As a result, these mutations result in the NMD of the *ARID1B* mRNA rather than the expression of mutant *ARID1B* protein. Truncating mutations that circumvent NMD typically result in a distinct and more extreme phenotype than the dominant negative impacts of the mutant protein²⁵. Human learning and memory may be affected by frameshift mutations in the *ARID1B* gene, which replace glutamine with arginine, glycine, or serine. Similarly, mutations causing normal coding amino acids like threonine and alanine to a different branched amino proline is likely to disrupt the structure of the *ARID1B* protein. Stop-gain mutation of glycine affects immunomodulatory in the peripheral nervous system. *ARID1B* gene variants linked to ASD may lead to a reduction *ARID1B* protein level or a disturbance in the protein's chromatin remodeling activity. As a result, the *ARID1B* gene plays an essential part in brain development. Further research needs to be conducted to determine whether individuals with *ARID1B* haploinsufficiency have altered brain development, which contributes to the intellectual disability and speech difficulty defined by ASD^{28,29}. **ASH1 like histone lysine methyltransferase (*ASH1L*)**

The lysine-specific methyltransferase 2H enzyme is produced by *ASH1L* gene and can be found in a range of organs and tissues throughout the body³¹. Lysine-specific methyltransferase 2H functions as a histone methyltransferase enzyme that modify histone proteins. Histone methyltransferases control the function of few genes by methylation of histone²⁷. Additionally, certain genes involved in brain development are activated by lysine-specific methyltransferase 2H. Histone methylation is disrupted by the lack of a functioning lysine-specific methyltransferase 2H enzyme³². Loss of *ASH1L* gene has been shown to impair embryonic and postnatal brain development, as well as the formation of neuronal networks in the developing hypothalamus, which is required for optimal feeding behaviors and initial postnatal growth^{32, 33}. At least seven *ASH1L* gene mutations were found to be associated with ASD. Some *ASH1L* gene mutations linked to ASD induce a single amino acid alteration in the lysine-specific methyltransferase 2H enzyme, while others disrupt

genetic material from the *ASHIL* gene sequence or produce an early stop signal, rendering the enzyme inactive and affecting neural connection. For example; when alanine to serine and arginine to glutamine missense mutations occur in *ASHIL* gene, alanine production is impaired, which is a source of energy in the CNS, and the body's metabolism is also affected as arginine production is hindered.

Chromodomain helicase DNA binding protein 2 (*CHD2*)

CHD2 gene codes for chromodomain DNA helicase protein 2 (*CHD2*) which controls gene activity through chromatin remodeling and is present in all cells of the body³⁴. This protein plays a significant role in the function and growth of neurons present in the brain. It is proposed that the protein translated from the mutant *CHD2* gene causes enhanced neuronal excitability due to the action of GABAergic neuron excitability or other electrophysiological channels and cause convulsions, and have little or no effect on RNA expression levels³⁵. *CHD2* related

neurodevelopmental disorders are autosomal dominant disorders caused by a *de novo* pathogenic variant. Twenty-seven *CHD2* gene variants linked to ASD were investigated. These mutations may result in moderate alterations in the expression of multiple genes, due to their induced functional disruption of the *CHD2* protein, all of which affect development of brain and increase the possibility of ASD. For example, valine is used for energy production while serine helps in biosynthesis of purines and pyrimidines. The frameshift mutation in *CHD2* gene of valine and serine results in a stop codon that completely disrupts chromatin remodeling and can possibly lead to ASD.

Dual specificity tyrosine phosphorylation regulated kinase 1A (*DYRK1A*)

DYRK1A encodes a kinase enzyme involved in the phosphorylation of proteins which helps in regulating the activity of proliferation and differentiation of cells and play important role in the development of the nervous system³¹. Dendritic spines assist convey nerve impulses and facilitate communication between neurons, and the

Table 1. Table showing genes responsible for ASD, their location and function

Genes	Location	Normal Function In Cell
ADNP	Long arm (q) of Chromosome 20 band 13 (Location: 20q13.13)	Encodes for the protein involved in proper brain development by controlling the expression of other genes through chromatin remodeling.
ARID1B	Long arm (q) of Chromosome 6 band 25 (Location: 6q25.3)	Provides instructions to encode a protein composed of several SWI/SNF protein complexes, forming a subunit which regulates gene activity by chromatin remodeling and functions as a tumour suppressor.
ASH1L	Long arm (q) of Chromosome 1 band 22 (Location: 1q22)	Provides instructions for making a histone methyltransferase enzyme known as lysine-specific methyltransferase 2H. This enzyme stimulates the expression of genes involved in brain development.
CHD2	Long arm (q) of Chromosome 15 band 26 (Location: 15q26.1)	This gene encodes a chromodomain DNA helicase protein 2 that regulates gene activity via chromatin remodeling. It aids in the modulation of neuron growth and function.
DYRK1A	Long arm (q) of Chromosome 21 (Location: 21q22.2)	DYRK1A is involved in phosphorylation and in the formation and maturation of dendritic spines from dendrites.
POGZ	Long arm (q) of chromosome 1 (Location: 1q21.3)	POGZ protein regulates gene expression by altering how tightly sections of DNA are packed by binding to chromatin for chromatin remodeling.
SHANK3	Long arm (q) of Chromosome 22 (Location: 22q13.3)	SHANK3 protein ensures that the signals sent by one neuron are received by another. It serves as a framework for the connections between neurons and is also involved in dendritic spine development and maturation.
SYNGAP1	Short arm (p) of chromosome 6 (Location: 6p21.32)	SynGAP controls neuronal adaptations, supports correct brain circuitry.

Table 2. Table showing pathogenic alleles of the genes* responsible for ASD

Gene	rsID	Amino Acid Seq Change	Consequence of mutation	Variation Type	
<i>ADNP</i>	rs58777525	Y [Tyr] ⇒ Ter[*] [AMB]	Frameshift variant	Indel	
	rs58777523	N [Asn] ⇒ K [Lys]	Frameshift variant	Indel	
	rs58777522	L [Leu] ⇒ I [Ile]	Frameshift variant	Indel	
	rs1057518978	Y [Tyr] ⇒ V [Val]	Frameshift variant	Indel	
	rs886042026	K [Lys] ⇒ Q [Gln]	Frameshift variant	Indel	
	rs779340209	S [Ser] ⇒ Ter [*] [OPA]	Stop-gained	Single Nucleotide Variation	
	rs1131691770	L [Leu] ⇒ Ter [*] [AMB]	Stop Gained	Indel	
	rs886041116	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
	rs58777526	Y [Tyr] ⇒ Y [Tyr]	Stop Gained	Single Nucleotide Variation	
	rs1135401808	Y [Tyr] ⇒ Ter [*] [OCH]	Stop gained	Indel	
	rs886041449	L [Leu] ⇒ P [Pro]	Frameshift variant	Deletion	
	rs58777524	S [Ser] ⇒ Ter [*] [AMB]	Stop gained	Single Nucleotide Variation	
	<i>ARID1B</i>	rs587777114	S [Ser] ⇒ P [Pro]	Intron Variant	Single Nucleotide Variation
		rs867707366	S [Ser] ⇒ L [Leu]	Stop gained	Single Nucleotide Variation
		rs387907143	K [Lys] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation
		rs750447037	Q [Gln] ⇒ K [Lys]	Stop Gained	Single Nucleotide Variation
		rs797045282	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
		rs886041819	W [Trp] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
		rs1028186690	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
		rs797045283	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
rs1057517704		Q [Gln] ⇒ E [Glu]	Stop Gained	Single Nucleotide Variation	
rs879253747		Q [Gln] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation	
rs902091666		K [Lys] ⇒ E [Glu]	Stop Gained	Single Nucleotide Variation	
rs758570139		Y [Tyr] ⇒ Y [Tyr]	Stop Gained	Single Nucleotide Variation	
rs387907144		R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
<i>ASH1L</i>		rs863225241	R [Arg] ⇒ Q [Gln]	Missense variant	Single Nucleotide Variation
		rs1293246328	A [Ala] ⇒ S [Ser]	Missense variant	Single Nucleotide Variation
		rs864309548	Q [Gln] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation
		rs398122998	W [Trp] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
		rs1057523601	Q [Gln] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation
	rs797044912	R [Arg] ⇒ R [Arg]	Stop Gained	Single Nucleotide Variation	
	rs761127171	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
	rs767106034	R [Arg] ⇒ R [Arg]	Stop Gained	Single Nucleotide Variation	
	rs777803897	R [Arg] ⇒ R [Arg]	Stop Gained	Single Nucleotide Variation	
	rs773860345	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
	rs775406327	S [Ser] ⇒ L [Leu]	Stop Gained	Single Nucleotide Variation	
	rs2272457	Y [Tyr] ⇒ Y [Tyr]	Stop Gained	Single Nucleotide Variation	
	rs1057519228	T [Thr] ⇒ T [Thr]	Synonymous Variant	Single Nucleotide Variation	
	rs397514739	T [Thr] ⇒ L [Leu]	Splice Donor Variant	Indel	
	<i>DYRK1A</i>	rs886041658	Y [Tyr] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation
rs797044522		D [Asp] ⇒ R [Arg]	Frameshift Variant	Indel	
rs886039652		V [Val] ⇒ F [Phe]	Frameshift Variant	Indel	
rs797045042		A [Ala] ⇒ G [Gly]	Frameshift Variant	Indel	
rs724159956		R [Arg] ⇒ T [Thr]	Frameshift Variant	Indel	
rs1064794006		K [Lys] ⇒ N [Asn]	Frameshift Variant	Indel	
rs797045539		P [Pro] ⇒ T [Thr]	Frameshift Variant	Insertion	
rs724159953		R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
rs797044520		R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
rs797045041		R [Arg] ⇒ Q [Gln]	Missense variant	Single Nucleotide Variation	
rs1064796713		N [Asn] ⇒ I [Ile]	Frameshift Variant	Indel	
<i>POGZ</i>		rs1235715459	P [Pro] ⇒ S [Ser]	Frameshift Variant	Indel
	rs869312834	Q [Gln] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation	
	rs869320764	E [Glu] ⇒ T [Thr]	Frameshift Variant	Deletion	
	rs864321667	L [Leu] ⇒ C [Cys]	Frameshift Variant	Indel	
	rs796052217	Q [Gln] ⇒ R [Arg]	Frameshift Variant	Deletion	
	rs1085307702	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
	rs864321673	S [Ser] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation	
	rs1057518170	M [Met] ⇒ V [Val]	Frameshift Variant	Deletion	

	rs869320763	E [Glu] ⇒ Ter [*] [OPA]	Frameshift Variant	Indel
	rs142133690	Y [Tyr] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation
	rs864321675	L [Leu] ⇒ F [Phe]	Frameshift variant	INDEL
SHANK3	rs886041467	P [Pro] ⇒ L [Leu]	Frameshift Variant	Indel
	rs1294272918	Q [Gln] ⇒ E [Glu]	Stop Gained	Single Nucleotide Variation
	rs886041430	L [Leu] ⇒ R [Arg]	Frameshift Variant	Indel
	rs762292772	A [Ala] ⇒ P [Pro]	Frameshift Variant	Indel
	rs761720914	E [Glu] ⇒ K [Lys]	Stop Gained	Single Nucleotide Variation
	rs886041238	R [Arg] ⇒ L [Leu]	Frameshift Variant	Indel
	rs1238131472	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
	rs1057519395	S [Ser] ⇒ R [Arg]	Frameshift Variant	Indel
	rs1064793514	L [Leu] ⇒ R [Arg]	Frameshift Variant	Deletion
	rs1131691727	P [Pro] ⇒ H [His]	Frameshift Variant	Indel
	rs1064795759	Q [Gln] ⇒ Ter [*] [AMB]	Stop Gained	Single Nucleotide Variation
	rs121918316	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
SYNGAP1	rs587780470	L [Leu] ⇒ C [Cys]	Frameshift Variant	Indel
	rs1131691635	L [Leu] ⇒ S [Ser]	Frameshift Variant	Indel
	rs1060503386	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
	rs1057519400	S [Ser] ⇒ Q [Gln]	Frameshift Variant	Indel
	rs1060503383	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
	Rs797046029	L [Leu] ⇒ P [Pro]	Frameshift Variant	Indel
	rs797046028	V [Val] ⇒ Ter [*] [AMB]	Stop gained	Indel
	rs1057519405	P [Pro] ⇒ R [Arg]	Frameshift Variant	Indel
	rs397514670	P [Pro] ⇒ L [Leu]	Missense variant	Single Nucleotide Variation
	rs1131692154	R [Arg] ⇒ Ter [*] [OPA]	Stop Gained	Single Nucleotide Variation
	rs121918315	K [Lys] ⇒ Ter [*] [OCH]	Stop Gained	Single Nucleotide Variation

*(only some data is provided)

DYRK1A enzyme is involved in their production and maturation from dendrites in neurons³⁶. A nonsense mutation results in the synthesis of a kinase domain protein with a C-terminally shortened C-terminus (DYRK1A-E396ter), which is then processed by the proteasome into an inactive form, demonstrating overall loss-of-function of DYRK1A³⁷. Anxiety, microcephaly (an abnormally small head), intellectual disabilities, and speech issues are other specific traits seen in ASD patients with *DYRK1A* gene mutations³⁸. Chronic seizures (epilepsy), distinctive facial features, poor muscular tone (hypotonia), foot deformities, and walking impairments may be experienced by these individuals³⁸. At least twenty-five *DYRK1A* gene mutations associated with ASD were identified. DYRK1A enzyme dysfunction or absence causes aberrant gene expression regulation and affects normal brain development. For example, frameshift mutation in *DYRK1A* gene changes normal coding tyrosine to a termination codon which can lead to production of a truncated protein which in turn can have many determinantal effects on the brain.

Pogo transposable element derived with ZNF domain (POGZ)

POGZ proteins are zinc finger proteins which are able to bind to the chromatin for chromatin remodeling owing to its structure which consist of a unique pattern of amino acids and one or more zinc ions and they are capable of binding. Zinc finger domain's folded form stabilizes the protein, and allows it to interact with many other molecules³⁹. POGZ protein modulates gene expression, which is critical for brain development, by interacting with the SP1 transcription factor, heterochromatin protein 1 (HP1), and chromodomain helicase DNA-binding protein 4 (CHD4) and acting as a chromatin regulator^{40, 41}. The large majority of previously reported de novo POGZ gene mutations in patients with neurodevelopmental disorders (NDDs) are nonsense and frameshift mutations dispersed between the C2H2 Zn finger and centromere protein-B-like DNA-binding (CENP-DB) domains, as well as within the CENP-DB domain itself. The large majority of reported de novo POGZ genetic variants in patients with neurodevelopmental disorders (NDDs) are

nonsense and frameshift mutations dispersed between the C2H2 Zn finger and centromere protein-B-like DNA-binding (CENP-DB) domains, including within the CENP-DB domain itself⁴². Twenty-two pathogenic mutations in the POGZ gene were investigated, which may result in a POGZ protein with reduced chromatin binding ability, causing aberrant chromatin remodeling and eventually compromising the normal expression of genes essential in brain development⁴². Mutations in UTR-5' and frameshift mutations in the POGZ gene can affect expression level and mRNA translation kinetics by producing DNA binding elements or cis-regulatory motifs.

SH3 and multiple ankyrin repeat domains 3 (SHANK3)

SHANK3 protein is vital in the working of synapses because it works as a platform that maintains the connections between neurons, ensuring that signals sent by one neuron are recognized by another⁴³. The N-terminal region of SHANK3 comprising of the Shank/ProSAP N-terminal (SPN) domain and a set of ankyrin (Ank) repeats has been reported with several missense mutations⁴⁴. These mutations impact

the Abi1 binding site in SHANK3, which is the core location of the SHANK protein, and missense mutations cause alterations outside of established interacting motifs in the C-terminal region of SHANK proteins, making it impossible to evaluate the precise role of these mutations⁴⁴. In the N-terminal motif of Shank/ProSAP (SPN), which is essential in dendritic spine growth and maturation, seven ankyrin repeats (Ank) have been identified as a hotspot for missense mutations⁴⁵. It has already been reported that disruption in communication between neurons contributes to the development of ASD⁴⁶. A minimum of 24 SHANK3 gene mutations associated with ASD were identified out of which most of them impairing SHANK3 protein function or synthesis. For example, mutation of proline to alanine impedes the assembly of several proteins and the rate of peptide bond production by the ribosome, cds-synon mutation of arginine and tyrosine, frameshift mutation of glutamic acid into a stop codon that handicaps memory and focus power, and missense mutation of isoleucine into phenylalanine may directly affects gene function.

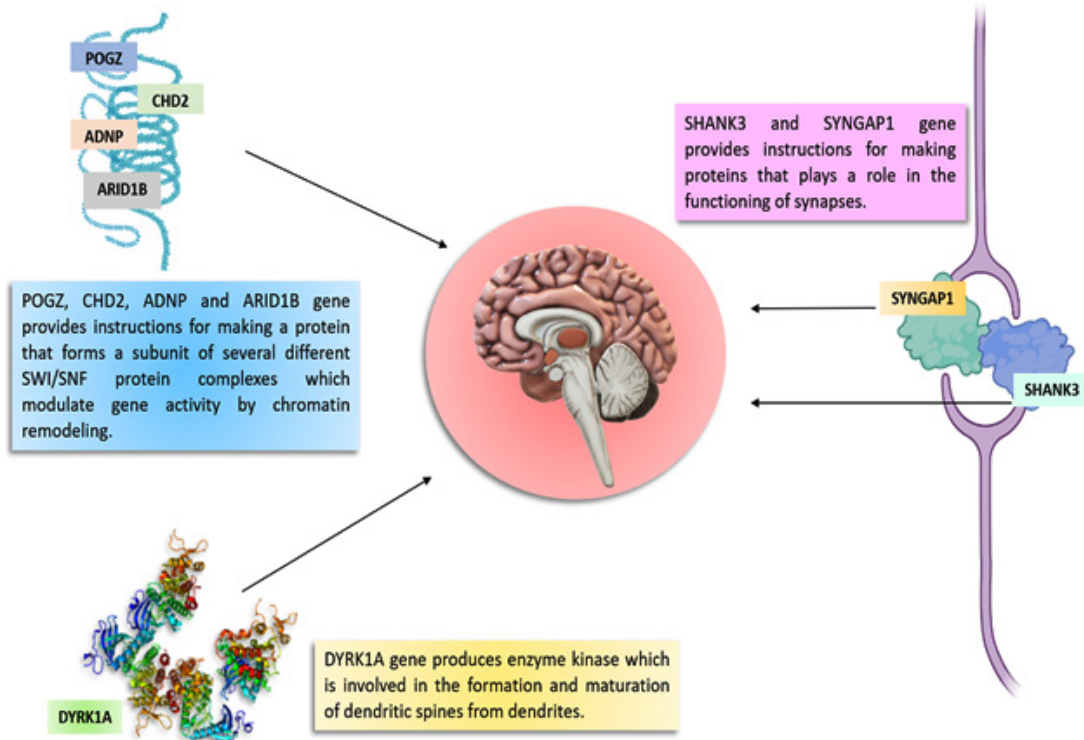


Fig. 1. Mutated genes that cause impairment of brain function leading to ASD

Synaptic Ras GTPase activating protein 1 (*SYNGAP1*)

SynGAP protein is encoded by *SYNGAP1* gene, located in synapses in the brain, and regulates important biochemical signaling pathways that enable learning and memory⁴⁷. SynGAP regulates synapse adaptations and promotes proper brain architecture by linking nerve cells, and it is especially vital during a crucial stage of early developing brain that influences long-term psychological capability⁴⁸. Pre-mRNA splicing in the *SYNGAP1* gene is dependent on the concordance of “cis” sequences that define exon-intron boundaries, which is required for proper protein translation, and regulatory sequences recognized by splicing machinery. Improper exon and intron recognition reported by point mutations at these consensus sequences may lead to the development of an abnormal transcript of the affected gene⁴⁵. At least thirty *SYNGAP1* gene mutations were identified, these mutations hampered or completely blocked the function or production of SynGAP protein associated with ASD. These mutations may be the underlying cause of behavioral impairments associated with ASD. For example, frameshift mutation of amino acids (tryptophan, leucine, valine and arginine) to stop codons and also mutations affecting normal coding the branched-chain amino acids like leucine, isoleucine, and valine with multiple functions in the brain can affect the synaptic plasticity in the brain.

CONCLUSION

The relevance of SNPs in ASD-associated genes as a risk factor was investigated in this study using *in silico* analysis. Mutations in the *ADNP*, *ARID1B*, *ASH1L*, *CHD2*, and *POGZ* genes have been identified to disrupt the development and functioning of the brain by altering gene regulation and causing abnormal chromatin modeling. Many of the SNP mutations studied result in a premature stop signal, resulting in a nonfunctional enzyme that is exceptionally short. Furthermore, these mutations in other genes associated with ASD, *SHANK3* and *SYNGAP1* disrupt the function of synapses and, as a result, cell-to-cell communication. These mutations may cause changes in synaptic adaption, which may exacerbate the behavioral problems associated

with ASD. These SNPs can act as biomarkers and provide insight into the etiology of ASD. Furthermore, the SNPs identified above may aid in the development of ASD medical interventions. More examination and validation of the identified SNPs will be required to assess their clinical importance and applicability in translation research as novel targets.

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Conflict of Interest

The authors have no conflict of interest to declare.

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REFERENCES

1. Hodges, H., Fealko, C., & Soares, N. (2020). Autism spectrum disorder: definition, epidemiology, causes, and clinical evaluation. *Transl. Pediatr.*, 2020; **9**(Suppl 1), S55-S65. <https://www.mhlw.go.jp/content/10500000/000704736.pdf>.
2. World Health Organization (WHO), Autism Spectrum Disorder <https://www.who.int/news-room/fact-sheets/detail/autism-spectrum-disorders>.
3. Ha, S., Sohn, I. J., Kim, N., Sim, H. J., & Cheon, K. A. Characteristics of Brains in Autism Spectrum Disorder: Structure, Function and Connectivity across the Lifespan. *Exp. Neurobiol.*, 2015; **24**(4):273–284. <https://doi.org/10.5607/en.2015.24.4.273>.
4. National Institute of Health, U.S. Department of National Institute of Neurological Disorders and Stroke <https://www.nimh.nih.gov/health/topics/autism-spectrum-disorders-asd>.
5. National Research Council (US) and Institute of Medicine (US) Committee on Integrating the Science of Early Childhood Development, Shonkoff, J. P., & Phillips, D. A. (Eds.). From Neurons to Neighborhoods: The Science of Early Childhood Development, 2000. National Academies Press (US).
6. Ratajczak H. V. Theoretical aspects of autism: causes—a review. *J. Immunotoxicol.* 2011; **8**(1), 68–79. <https://doi.org/10.3109/1547691X.2010.545086>.
7. Ha, S., Sohn, I. J., Kim, N., Sim, H. J., & Cheon, K. A. Characteristics of Brains in Autism Spectrum Disorder: Structure, Function and Connectivity across the Lifespan. *Exp. Neurobiol.*, 2015; **24**(4), 273–284. <https://doi.org/10.5607/en.2015.24.4.273>.

- org/10.5607/en.2015.24.4.273.
8. Grove J, Ripke S, Als TD, Mattheisen M, Walters RK, Won H, et al. Identification of common genetic risk variants for autism spectrum disorder. *Nat Genet.* 2019;**51**:431444.
 9. Robinson EB, iPSYCH-SSI-Broad Autism Group, St Pourcain B, Anttila V, Kosmicki JA, Bulik-Sullivan B, et al. Genetic risk for autism spectrum disorders and neuropsychiatric variation in the general population. *Nature Genetics.* 2016;**48**:552–555.
 10. Klei L, Sanders SJ, Murtha MT, Hus V, Lowe JK, Willsey AJ, et al. Common genetic variants, acting additively, are a major source of risk for autism. *Mol Autism.* 2012;**3**:9.
 11. Gaugler T, Klei L, Sanders SJ, Bodea CA, Goldberg AP, Lee AB, et al. Most genetic risk for autism resides with common variation. *Nat Genet.* 2014;**46**:881–885.
 12. Pandey, P.; Sharma, P. Analysis of Early Onset of Alzheimer's Disease Genes: Disease Causing and Risk Factors. *Eur. J. Biol. Res.*, 2021;**11**, 251-259. <http://dx.doi.org/10.5281/zenodo.4641962>
 13. Koemans TS, Kleefstra T, Chubak MC, et al. Functional convergence of histone methyltransferases EHMT1 and KMT2C involved in intellectual disability and autism spectrum disorder. *PLoS Genet.*, 2017;**13**(10):e1006864. <https://doi.org/10.1371/journal.pgen.1006864>
 14. Hoffmann, A., & Spengler, D. Chromatin Remodeler CHD8 in Autism and Brain Development. *J. Clin. Med.*, 2021; **10**(2), 366. <https://doi.org/10.3390/jcm10020366>
 15. <https://medlineplus.gov/genetics/condition/autism-spectrum-disorder/>
 16. <https://www.ncbi.nlm.nih.gov/variation/view/>
 17. Bassan, M., Zamostiano, R., Davidson, A., Pinhasov, A., Giladi, E., Perl, O., Bassan, H., Blat, C., Gibney, G., Glazner, G., Brenneman, D. E., & Gozes, I. Complete sequence of a novel protein containing a femtomolar-activity-dependent neuroprotective peptide. *J. Neurochem.*, 1999; **72**(3), 1283–1293. <https://doi.org/10.1046/j.1471-4159.1999.0721283.x>
 18. Said S. I. Molecules that protect: the defense of neurons and other cells. *J. Clin. Investig.*, 1996; **97**(10), 2163–2164. <https://doi.org/10.1172/JCI118655>.
 19. Arnett, A. B., Rhoads, C. L., Hoekzema, K., Turner, T. N., Gerds, J., Wallace, A. S., Bedrosian-Sermone, S., Eichler, E. E., & Bernier, R. A. The autism spectrum phenotype in ADNP syndrome. *Autism Res.*, 2018; **11**(9), 1300–1310. <https://doi.org/10.1002/aur.1980>
 20. Magen, I., & Gozes, I. Davunetide: Peptide therapeutic in neurological disorders. *Curr. Med. Chem.*, 2014; **21**(23), 2591–2598. <https://doi.org/10.2174/0929867321666140217124945>
 21. Vandeweyer G, Helsmoortel C, Van Dijck A, et al. The transcriptional regulator ADNP links the BAF (SWI/SNF) complexes with autism. *Am J Med Genet C Semin Med Genet.*, 2014;**166C**(3):315-326. <https://doi.org/10.1002/ajmg.c.31413>
 22. Helsmoortel C, Vulto-van Silfhout AT, Coe BP, et al. A SWI/SNF-related autism syndrome caused by de novo mutations in ADNP. *Nat Genet.*, 2014;**46**(4):380-384. doi:10.1038/ng.2899
 23. Kapitansky O, Gozes I. ADNP differentially interact with genes/proteins in correlation with aging: a novel marker for muscle aging. *Geroscience.*, 2019;**41**(3):321-340. <https://doi.org/10.1007/s11357-019-00079-x>
 24. Sim JC, White SM, Lockhart PJ. ARID1B-mediated disorders: Mutations and possible mechanisms. *Intractable Rare Dis Res.*, 2015;**4**(1):17-23. <https://doi.org/10.5582/irdr.2014.01021>
 25. Wang X, Nagl NG, Wilsker D, et al. Two related ARID family proteins are alternative subunits of human SWI/SNF complexes. *Biochem J.*, 2004;**383**:319-325. <https://doi.org/10.1042/BJ20040524>
 26. Inoue H, Furukawa T, Giannakopoulos S, Zhou S, King DS, Tanese N. Largest subunits of the human SWI/SNF chromatin-remodeling complex promote transcriptional activation by steroid hormone receptors. *J Biol Chem.*, 2002;**277**(44):41674-41685. <https://doi.org/10.1074/jbc.M205961200>
 27. Hurlstone AF, Olave IA, Barker N, van Noort M, Clevers H. Cloning and characterization of hELD/OSA1, a novel BRG1 interacting protein. *Biochem J.*, 2002;**364**(Pt 1):255-264. <https://doi.org/10.1042/bj3640255>
 28. Schweingruber C, Rufener SC, Zünd D, Yamashita A, Mühlemann O. Nonsense-mediated mRNA decay - mechanisms of substrate mRNA recognition and degradation in mammalian cells. *Biochim Biophys Acta.*, 2013;1829(6-7):612-623.. <https://doi.org/10.1016/j.bbarm.2013.02.005>
 29. Halgren C, Kjaergaard S, Bak M, et al. Corpus callosum abnormalities, intellectual disability, speech impairment, and autism in patients with haploinsufficiency of ARID1B. *Clin Genet.*, 2012;**82**(3):248-255. <https://doi.org/10.1111/j.1399-0004.2011.01755.x>
 30. ASH1L gene, Genetic Home Reference.
 31. Gao Y, Duque-Wilckens N, Aljazi MB, et al. Loss of histone methyltransferase ASH1L in

- the developing mouse brain causes autistic-like behaviors. *Commun Biol.*, 2021;**4**(1):756. <https://doi.org/10.1038/s42003-021-02282-z>
32. Cheon S, Culver AM, Bagnell AM, et al. Counteracting epigenetic mechanisms regulate the structural development of neuronal circuitry in human neurons. *Mol Psychiatry.*, 2022;10.1038/s41380-022-01474-1. <https://doi.org/10.1038/s41380-022-01474-1>
 33. Nieto-Estevez V, Hsieh J. CHD2: One Gene, Many Roles. *Neuron.* 2018;**100**(5):1014-1016. <https://doi.org/10.1016/j.neuron.2018.11.036>
 34. Carvill GL, Mefford HC. CHD2-Related Neurodevelopmental Disorders. In: Adam MP, Ardinger HH, Pagon RA, et al., eds. GeneReviews®. Seattle (WA): University of Washington, Seattle; December 10, 2015.
 35. Nourbakhsh K, Yadav S. Kinase Signaling in Dendritic Development and Disease. *Front Cell Neurosci.*, 2021;**15**:624648. <https://doi.org/10.3389/fncel.2021.624648>
 36. Martinez de Lagran M, Benavides-Piccione R, Ballesteros-Yañez I, et al. Dyrk1A influences neuronal morphogenesis through regulation of cytoskeletal dynamics in mammalian cortical neurons. *Cereb Cortex.*, 2012;**22**(12):2867-2877. <https://doi.org/10.1093/cercor/bhr362>
 37. Lee KS, Choi M, Kwon DW, et al. A novel de novo heterozygous DYRK1A mutation causes complete loss of DYRK1A function and developmental delay. *Sci Rep.*, 2020;**10**(1):9849. <https://doi.org/10.1038/s41598-020-66750-y>
 38. Levy JA, LaFlamme CW, Tsaprilis G, Crynen G, Page DT. Dyrk1a Mutations Cause Undergrowth of Cortical Pyramidal Neurons via Dysregulated Growth Factor Signaling. *Biol Psychiatry.*, 2021;**90**(5):295-306. <https://doi.org/10.1016/j.biopsych.2021.01.012>
 39. Markenscoff-Papadimitriou E, Binyameen F, Whalen S, et al. Autism risk gene POGZ promotes chromatin accessibility and expression of clustered synaptic genes. *Cell Rep.*, 2021;**37**(10):110089. <https://doi.org/10.1016/j.celrep.2021.110089>.
 40. Tan B, Zou Y, Zhang Y, et al. A novel de novo POGZ mutation in a patient with intellectual disability. *J Hum Genet.*, 2016;**61**(4):357-359. <https://doi.org/10.1038/jhg.2015.156>
 41. Suliman-Lavie, R., Title, B., Cohen, Y., Hamada, N., Tal, M., Tal, N., Monderer-Rothkoff, G., Suliman-Lavie R, Title B, Cohen Y, et al. Author Correction: Pogz deficiency leads to transcription dysregulation and impaired cerebellar activity underlying autism-like behavior in mice. *Nat Commun.*, 2021;**12**(1):3874. <https://doi.org/10.1038/s41467-021-24166-w>
 42. Suliman-Lavie R, Title B, Cohen Y, et al. Author Correction: Pogz deficiency leads to transcription dysregulation and impaired cerebellar activity underlying autism-like behavior in mice. *Nat Commun.*, 2021;**12**(1):3874. <https://doi.org/10.1038/s41467-020-14697-z>
 43. Halbedl S, Schoen M, Feiler MS, Boeckers TM, Schmeisser MJ. Shank3 is localized in axons and presynaptic specializations of developing hippocampal neurons and involved in the modulation of NMDA receptor levels at axon terminals. *J Neurochem.*, 2016;**137**(1):26-32. <https://doi.org/10.1111/jnc.13523>.
 44. Woike D, Wang E, Tibbe D, et al. Mutations affecting the N-terminal domains of SHANK3 point to different pathomechanisms in neurodevelopmental disorders. *Sci Rep.*, 2022;**12**(1):902. <https://doi.org/10.1038/s41598-021-04723-5>.
 45. Hassani Nia F, Kreienkamp HJ. Functional Relevance of Missense Mutations Affecting the N-Terminal Part of Shank3 Found in Autistic Patients. *Front Mol Neurosci.*, 2018;**11**:268. <https://doi.org/10.3389/fnmol.2018.00268>.
 46. Vyas Y, Cheyne JE, Lee K, Jung Y, Cheung PY, Montgomery JM. Shankopathies in the Developing Brain in Autism Spectrum Disorders. *Front Neurosci.*, 2021;**15**:775431. <https://doi.org/10.3389/fnins.2021.775431>.
 47. Gamache TR, Araki Y, Haganir RL. Twenty Years of SynGAP Research: From Synapses to Cognition. *J Neurosci.*, 2020;**40**(8):1596-1605. <https://doi.org/10.1523/JNEUROSCI.0420-19.2020>.
 48. Brimble E, Lee-Messer C, Nagy PL, Propst J, Ruzhnikov MRZ. Clinical Transcriptome Sequencing Confirms Activation of a Cryptic Splice Site in Suspected SYNGAP1-Related Disorder. *Mol Syndromol.*, 2019;**9**(6):295-299. <https://doi.org/10.1159/000492706>