

## Aggregatibacter Actinomycetemcomitans: its Role in Periodontitis

RAADHA RAGAVENDRAN<sup>1</sup>, V. RAMYA, PREETHE<sup>2</sup> and PADDMANABHAN<sup>2</sup>

<sup>1</sup>Department of Periodontics, Tagore Dental College & Hospital, Rathinamangalam, Chennai, India.

<sup>2</sup>Department of Periodontics, Sree Balaji Dental College & Hospital,  
Bharath University, Chennai-600100, India.

DOI: <http://dx.doi.org/10.13005/bpj/685>

(Received: July 25, 2015; accepted: September 10, 2015)

### ABSTRACT

Strong evidence is available on the role of *Aggregatibacter actinomycetemcomitans* (A.a) as the causative agent of localised juvenile periodontitis (LJP), a disease characterised by rapid destruction of the tooth-supporting tissues. This organism possesses a large number of virulence factors with a wide range of activities which enable it to colonise the oral cavity, invade periodontal tissues, evade host defences, initiate connective tissue destruction and interfere with tissue repair. Adhesion to epithelial and tooth surfaces is dependent on the presence of surface proteins and structures such as microvesicles and fimbriae. Invasion has been demonstrated in vivo and in vitro. The organism has a number of means of evading host defences, which include: (i) production of leukotoxin; (ii) producing immunosuppressive factors; (iv) secreting proteases capable of cleaving IgG; and (v) producing Fc-binding.

**Key words:** Periodontitis, Leukotoxin, Lipopolysaccharide.

### INTRODUCTION

It has been well documented that biofilm bacteria predominate, numerically and metabolically, in virtually all nutrient-sufficient ecosystems, including the oral cavity<sup>1,2</sup>. Bacterial cells in biofilms are surrounded by a self-synthesized, three-dimensional matrix, which holds the cells together and firmly attaches the bacterial cells to the underlying surface<sup>3</sup>. The extracellular polymeric substance has been attributed to a protective role as well as it is a source of dissolved nutrients, secreted enzymes, extracellular DNA and exopolysaccharide. The exopolysaccharide of *Aggregatibacter actinomycetemcomitans* (PGA) is a homopolymer of N-acetyl-D-glucosamine residues in  $\alpha(1,6)$  linkage and has been well characterized in several bacteria including *Staphylococcus aureus*, *S. epidermidis* and *E. coli*. This exopolysaccharide has been named differently in various bacteria but its synthesis is encoded by a set of four genes, *icaADBC* in *Staphylococcal* species and *pgaABCD* in *E. coli* and *A. actinomycetemcomitans*. The exopolysaccharide from *A. actinomycetemcomitans*,

PGA, is a surface-associated polymer that can protect *A. actinomycetemcomitans* at the cellular level from phagocytic killing<sup>4</sup>. A similar protective function was ascribed to the exopolysaccharide PIA of *S. epidermidis*<sup>5</sup>. PGA/PIA mediates resistance to killing by antibiotics<sup>6</sup>, detergents<sup>7</sup> and antimicrobial peptides<sup>5</sup>. PGA may act through a general mechanism wherein it binds to or electrostatically repulses immune modulators and antimicrobial agents, thereby preventing their access to the bacterial cell<sup>5</sup>. The importance of PGA in a protective role for *A. actinomycetemcomitans* and other bacteria has been well established<sup>4</sup>. In addition, a recent study of Ps1, the exopolysaccharide of *P. aeruginosa*, has demonstrated that it also determines the fate of elite cells in the initial microcolony development<sup>8</sup>. While these studies highlight the significance of exopolysaccharide, they also bring the genes encoding the exopolysaccharide to the forefront in the disease process. For example, in several infection models, the exopolysaccharide PIA has been demonstrated to be relevant for the virulence of *S. epidermidis*<sup>9,10,11</sup>.

### Morphology

Aggregatibacter actinomycetemcomitans (actis, a ray; myces, a fungus; comitans, accompanying; actinomycetemcomitans, accompanying an actinomycete) is a gram-negative coccobacillus measuring about  $0.4 \pm 0.1 \times 0.1 \pm 0.4$  micrometers in size. Aggregatibacter actinomycetemcomitans possess fimbriae, vesicles and extracellular amorphous materials. MGB (trypticase soy broth) with malachite green and bacitracin was the earliest media used to culture (A.a). It was then followed by medium with trypticase soy agar, serum with bacitracin and vancomycin (TSBV). Exclusive growth of A.a was found in a particular culture medium which contained TSBV, spiramycin, fucidic acid and carbencillin. RPMI – 1640 and Dulbecco's modified Eagle medium are now used with a generation time of 246 and 346 min<sup>12</sup>.

### Virulence factors

The putative virulence factors of A. actinomycetemcomitans can be subdivided into those that: (i) modulate inflammation, (ii) induce tissue destruction and (iii) inhibit tissue repair. The most actively studied gene product of the organism is a leukotoxin and a member of the RTX (repeats in toxin) family whose cellular receptor is the integrin, LFA-1, thus accounting for its selective effect on leucocytes (although only those from primates)<sup>13-15</sup>. Almost all the RTX leukotoxins are secreted except LtxA toxin of A. actinomycetemcomitans which is thought to be entirely cell associated; either bound to cell surface-associated nucleic acids<sup>16</sup> or within membranous vesicles which bud from bacterium's surface<sup>17,18</sup>. This affirms the possibility that the bacterium itself is toxic to the target cells. The apoptosis of the target cells in response to A.actinomycetemcomitansleuko toxin is by a mechanism involving mitochondrial perturbation<sup>19</sup>. Injection of A. actinomycetemcomitans into mice has been claimed to induce immunosuppression and sonicates of this organism suppressed the IgG response to sheep red blood cells in mice<sup>20,21</sup>. It has also been proposed that A.actinomycetemcomitans can produce super antigens, which have the ability to bring about T cell apoptosis by binding to T cell receptors<sup>22,23</sup>. A. actinomycetemcomitans has been reported to produce a number of, as yet unidentified, proteins

with cell cycle-inhibitory activity causing arrest in the G2 phase of the cell cycle. These proteins range in molecular mass from the 8-kDa protein termed gapstatin to 60 kDa and all the way up to 80 kDa<sup>24</sup>. One cell cycle-modulatory protein with immunosuppressive function that has recently been identified as being produced by A. actinomycetemcomitans is cytolethal distending toxin (CDT) Fc binding protein termed as Omp34 identical with OmpA of E. coli, a protein implicated in the virulence of this organism is another immunomodulatory virulence factor of A. actinomycetemcomitans. A. actinomycetemcomitans produces a 65-kDa macromolecule able to bind to the IL-10 receptor and henceforth can modulate monocyte/macrophage function as IL- 10 is considered to be a major macrophage de-activating cytokine. A. actinomycetemcomitans has also been reported to produce a low molecular mass inhibitor of neutrophil chemotaxis to FMLP. LPS is reported to stimulate bone resorption in vitro and in vivo. But it is considered to be a less significant cytokine inducer than the secreted protein. A cell stress protein, chaperonin 60 is considered to be a potent bone degrading molecule by stimulating bone resorption by acting as an osteoclast 'growth factor'

### Virulence mechanism

#### Adhesion

Bacterial adhesion, which facilitates colonization is the key virulent mechanism<sup>25</sup> Bacterial components involved in Adhesion are called adhesins. They are proteinaceous structures found on cell surfaces. They bind with specific receptors in the saliva, tooth, extra cellular matrix and epithelial cells. Surface entities like vesicles mediate aggregation. A.actinomycetemcomitans adheres to the gingival crevice epithelium. Strains with fimbriae adhere three to four folds better. A.actinomycetemcomitans binds to collagen I,II,III and V but not IV. It also binds to fibronectin but not fibrinogen. The tight auto-adhesion of A. actinomycetemcomitans has been described is due to the expression of long, bundled fibrils composed of a 6.5-kDa subunit protein, Flp-1 (fimbrial low-mol. wt protein) which has been reported to be glycosylated. Bacteriocins are proteins produced by bacteria that are lethal for other strains and species of bacteria. These agents confer colonization by lessening ecological pressures. This is an advantage for the bacterium.

### Invasion

It has been affirmed that many bacteria have the ability to invade host cells and *A. actinomycetemcomitans* is one among them. Studies of invasion of *A. actinomycetemcomitans* reveal that 25% of *A. actinomycetemcomitans* isolates are invasive. *A. Actinomycetemcomitans* penetrate and survive within eukaryotic cells. They penetrate gingival epithelium. They occur in specific intracellular locations like the epithelial wall, enlarged intracellular pocket spaces and the epithelial side of basal lamina in connective tissue and alveolar bone. It has been observed that microfilaments and microtubules for intracellular movement. The process of intracellular movement and the cell spreading could be inhibited by agents that interfered with microtubule dynamics, suggesting that this bacterium when internalized interacts closely with the microtubules of the host cell. It has been suggested that the transferrin and integrin receptors are involved in the adhesion of the bacteria to host cells.

of potential virulence factors and mechanisms. Though it was initially named as *Actinobacillus actinomycetemcomitans*, it was found that the bacterium is more similar to *haemophilus* than *actinobacillus* and hence it was reclassified under *aggregatibacter* as *Aggregatibacter actinomycetemcomitans*. Scientific data clearly underlines its etiological role in localized aggressive periodontitis. This review also tries to throw light on the virulence abilities of this pathogen like immune evasion mechanisms like production of leukotoxin, cell cycle modulatory protein and immunomodulatory protein like Fc binding proteins. It also brings about tissue destruction by other novel mechanisms like binding to host matrices and invading host cells. Still, a lot is still to be understood and established. With the advent of newer technological methodologies and genome information, we would be able to understand not only how *A. actinomycetemcomitans* produces such profound but local pathology like periodontal infections but also its role in systemic pathology

### CONCLUSION

*A. actinomycetemcomitans* is a highly non motile gram negative coccobacillus with a vast array

### REFERENCES

1. Costerton JW, Lewandowski Z, Caldwell DE, Korber DR, Lappin-Scott HM, Microbial biofilms. *Annu Rev Microbiol* : 711–745 (1995). PMID: 8561477
2. Kolenbrander PE, Oral microbial communities: biofilms, interactions, and genetic systems. *Annu Rev Microbiol* **54**: 413–437 (2000). PMID: 11018133
3. Costerton JW, Stewart PS, Greenberg EP, Bacterial biofilms: a common cause of persistent infections. *Science* **284**: 1318–1322 (1999). PMID: 10334980
4. Venketaraman V, Lin AK, Le A, Kachlany SC, Connell ND, *et al.* Both leukotoxin and poly-N-acetylglucosamine surface polysaccharide protect *Aggregatibacter actinomycetemcomitans* cells from macrophage killing. *MicrobPathog* **45**: 173–180 (2008). doi: 10.1016/j.micpath.2008.05.007 PMID: 18573331
5. Vuong C, Voyich JM, Fischer ER, Braughton KR, Whitney AR, *et al.* Polysaccharide intercellular adhesin (PIA) protects *Staphylococcus epidermidis* against major components of the human innate immune system. *Cell Microbiol* 269–275 (2004). PMID: 14764110
6. Izano EA, Sadovskaya I, Vinogradov E, Mulks MH, Velliyagounder K, *et al.* Poly-N-acetylglucosamine mediates biofilm formation and antibiotic resistance in *Actinobacillus pleuropneumoniae*. *MicrobPathog* **43**: 1–9 (2007). PMID: 17412552
7. Izano EA, Wang H, Ragunath C, Ramasubbu N, Kaplan JB, Detachment and killing of *Aggregatibacter actinomycetemcomitans* biofilms by dispersin B and SDS. *J Dent Res* **86**: 618–622 (2007). PMID:
8. Zhao K, Tseng BS, Beckerman B, Jin F, Gibiansky ML, *et al.* Psl trails guide exploration and microcolony formation in

- Pseudomonas aeruginosa* biofilms. *Nature* **497**: 388–391 (2013). doi: 10.1038/nature12155 PMID: 23657259
9. Rupp ME, Fey PD, In vivo models to evaluate adhesion and biofilm formation by *Staphylococcus epidermidis*. *Methods Enzymol* **336**: 206–215 (2001). PMID: 11398400
  10. Rupp ME, Ulphani JS, Fey PD, Bartscht K, Mack D, Characterization of the importance of polysaccharide intercellular adhesin/hemagglutinin of *Staphylococcus epidermidis* in the pathogenesis of biomaterial-based infection in a mouse foreign body infection model. *Infect Immun* **67**: 2627–2632 (1999).
  11. Rupp ME, Ulphani JS, Fey PD, Mack D, Characterization of *Staphylococcus epidermidis* polysaccharide intercellular adhesin/hemagglutinin in the pathogenesis of intravascular catheter-associated infection in a rat model. *Infect Immun* **67**: 2656–2659 (1999).
  12. Rurenga P, Raangs E, Singadji Z, Wekema-Mulder G, Veloo AC, Van Winkelhoff AJ. Evaluation of three selective media for isolation of *Aggregatibacter actinomycetemcomitans*. *J Periodontal Res.* **48**(5):549-52 (2013)
  13. Lally ET, Hill RB, Kieba IR, Korostoff J. The interaction between RTX toxins and target cells. *Trends Microbiol.* **7**: 356–61 (1999).
  14. Narayanan SV, Nagaraja TG, Chengappa MM, Stewart GC. Leukotoxins of gram-negative bacteria. *Vet Microbiol.* **84**:337-39 (2002).
  15. Lally ET, Kieba IR, Sato A, *et al.* RTX toxins recognize integrin on the surface of human target cells. *J Biol Chem.* **272**:30463–69 (1997).
  16. Ohta H, Hara H, Fukui K, Kurihara H, Murayama Y, Kato K. Association of *Actinobacillus actinomycetemcomitans* leukotoxin with nucleic acids on the bacterial cell surface. *Infect Immun.* **61**: 4878–84 (1993).
  17. Berthold P, Forti D, Kieba IR, Rosenbloom J, Taichman NS, Lally ET. Electron immunocytochemical localization of *Actinobacillus actinomycetemcomitans* leukotoxin. *Oral Microbiol Immunol.* **7**: 24–27 (1992).
  18. Kato S, Kowashi Y, Demuth DR. Outer membrane-like vesicles secreted by *Actinobacillus actinomycetemcomitans* are enriched in leukotoxin. *Microb Pathog.* **32**: 1–13 (2002).
  19. Korostoff J, Yamaguchi N, Miller M, Kieba I, Lally ET. Perturbation of mitochondrial structure and function plays a central role in *Actinobacillus actinomycetemcomitans* leukotoxin-induced apoptosis. *Microb Pathog.* ; **29**: 267–78 (2000).
  20. Chen PB, Davern LB, Neiders ME, Reynolds HS, Zambon JJ. Analysis of in vitro lymphoproliferative responses and antibody formation following the subcutaneous injection of *Actinobacillus actinomycetemcomitans* and *Wolinella recta* in a murine model. *Oral Microbiol Immunol,* **6**: 12–16 (1991).
  21. Kuritai Ochiai T, Ochiai K, Ikeda T. Immunosuppressive effects induced by *Actinobacillus actinomycetemcomitans*: Effect on immunoglobulin production and lymphokine synthesis. *Oral Microbiol Immunol.* ; **7**: 338–43 (1992).
  22. Nalbant A, Zadeh HH. Evidence for apoptosis of the majority of T cells activated in vitro with *Actinobacillus actinomycetemcomitans*. *Oral Microbiol Immunol.* **15**: 290–98 (2000).
  23. Zadeh HH, Nalbant A, Park K. Large-scale early in vitro response to *Actinobacillus actinomycetemcomitans* suggests superantigenic activation of T-cells. *J Dent Res.* **80**: 356–62 (2001).
  24. White PA, Wilson M, Nair SP, Kirby AC, Reddi K, Henderson B. Characterization of an antiproliferative surface-associated protein from *Actinobacillus actinomycetemcomitans* which can be neutralized by sera from a proportion of patients with localized juvenile periodontitis. *Infect Immun.* **63**:2612–18 (1995).
  25. Wilson M, McNab R, Henderson B. Bacterial disease mechanisms: an introduction to cellular microbiology. *Cambridge University Press.* (2002).