Polyamidoamine-Remdesivir Conjugate: Physical Stability and Cellular Uptake Enhancement

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Nucleoside analogue antiviral remdesivir works by inhibiting the RNA-dependent RNA polymerase enzyme and terminating the viral replication. Currently, remdesivir is under a clinical trial for its activity against SARS-CoV-2. In the blood, remdesivir will undergo an enzymatic reaction to become monophosphate analogue form which is difficult to penetrate into the cell membrane. PAMAM (polyamidoamine) dendrimer is a good carrier to encapsulate remdesivir as a water-insoluble drug (0,339 mg/mL). Entrapment of remdesivir in the PAMAM cavity avoided remdesivir molecules to not undergo the enzymatic reactions. This study aimed to synthesize, characterize and evaluate cellular uptake of PAMAM-Remdesivir conjugate. PAMAM-Remdesivir was prepared with various stirring times (3, 6, 12, 24, and 48 hours). The conjugates were characterized to observe the size and particle distribution using Particle Size Analyzer, encapsulating efficiency using UV-Vis Spectroscopy, interaction between PAMAM and remdesivir particle using Fourier Transform Infrared Spectroscopy and cellular uptake of PAMAM-RDV using Fluorescence Microscope. The optimized stirring time of PAMAM-Remdesivir conjugate was 24 hours wich resulted the particles charge of + 23,07 mV of zeta potential, 1008 nm of particle size, 0,730 of PDI, and 69% entrapment efficiency. In addition, the FTIR analysis showed that remdesivir molecules successfully conjugated to PAMAM. Thus, through strring optimization time, the remdesivir molecules were successfully entrapped to PAMAM cavity. The cellular uptake in Vero Cell of PAMAM-RDV conjugated fluorescein isothiocyanate was observed with fluorescence microscope and had a stronger intensity than remdesivir only solution.

Keywords: Conjugate; Fluorescein; Isothiocyanate; Polyamidoamine; Remdesivir.

Nucleoside analogue antiviral remdesivir is effectively in inhibiting Ebola and Marburg virus infections ¹. This antiviral works by inhibiting the RNA-dependent RNA polymerase enzyme, thus terminate the viral replication ^{1,2}. Currently, remdesivir is under a clinical trials of its activity against SARS-CoV-2. Previously, the antiviral activity of remdesivir against SARS-CoV-2 has been evaluated in vitro and in vivo. As result, remdesivir has better activity against SARS-CoV-2 than other antiviral ³. It indicates that remdesivir has potential as an antiviral for SARS-CoV-2.

The nucleoside triphosphate analogue from remdesivir can form a complex with RNA-dependent RNA-polymerase (RdRp) virus to inhibit the synthesis of viral RNA in cells. However, the result of in vivo pharmacokinetic test of remdesivir shows that the antiviral will

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undergo an enzymatic reaction become metabolite alanine, and rapidly become a monophosphate analogue after administrated intravenously ⁵ ⁶. The monophosphate analogue of remdesivir is negatively charged and polar which is difficult to penetrate into the cell membrane, while antiviral activity is highly dependent on the accumulation of drug molecules in the cells to cause an effect ⁴. ⁵. Therefore, remdesivir requires a drug delivery system that protects it from the enzymatic reactions berfot penetrating into the cells.

PAMAM is a with polyamide branch dendrimer with tertiary amine as the nucleus. It also has a cavity to encapsulate hydrophobic drug molecules. Therefore, PAMAM is a good carrier to encapsulate remdesivir as a water-insoluble drug (0,339 mg/mL). Commercially, PAMAM is available from the 1st to the 10th generations. The 4th generation is less toxic than the 5th or the 6th, and has a greater capacity of drug encapsulation than the previous generation ^{6,7}. Therefore, in this study PAMAM G4 was used. In physiological pH, encapsulated remdesivir in the PAMAM cavity will be protected from the environment, thusthe remdesivir molecules do not undergo the enzymatic reactions. ^{8,9}.

Currently, the synthesize of encapsulated remdesivir in PAMAM has not been reported yet. Thus, this study aimed to synthesize the PAMAM-Remdesivir (PAMAM-RDV) conjugate with has better stability than remdesivir without PAMAM. The conjugate will be characterized by examine the size and particle distribution using Particle Size Analyzer (PSA), encapsulating efficiency (EE) using UV-Vis spectroscopy, and interaction between PAMAM and remdesivir particle using Fourier Transform Infrared (FTIR) spectroscopy, and its cellular uptake at Vero cell by Flourescence Microscope.

MATERIAL AND METHODS

Reagents and Instruments

Remdesivir (Glentham Life Sciences), PAMAM G4 10% in methanol (Merck), dimethyl sulfoxide (Merck), water for injection (PT. Ikapharmindo Putramas), potassium bromide (Merck), Dulbecco's Modified Eagle's Medium (DMEM) (Merck), formaldehide (Merck). Spectrophotometer UV-Vis V-530 (Jasco, Japan), NANO-ZS series Malvern Zetasizer (Malvern Instruments, Malvern, UK), FTIR spectrophotometer 8400S (Shimadzu, Japan), fluorescence microscope opthiphot 2 (Nikon, Japan)

Methods

Synthesis of PAMAM-RDV conjugate

The stock solution of remdesivir was prepared by dissolving the remdesivir powder into 1 mL of DMSO. The mixture was stirred slowly with a spin bar on a magnetic stirrer until it dissolved. Then, 9 mL of water for injection was added to the mixture to obtain a concentration of 332 μ M stock solution of remdesivir. The encapsulation of remdesivir into the PAMAM cavity was made using the molarity ratio (10:20). The conjugate was made by stirring remdesivir in 10% DMSO for 10 minutes at room temperature. The solution was added by 10% PAMAM and stirred for 3, 6, 12, 24, and 48 hours. The conjugate was purified using a centrifugation ultrafiltration tube (MWCO of 10 kDa) at 5000 rpm for 90 minutes, the pure conjugate on the top of the tube was stored at 4°C and kept away from light.

Characterization of PAMAM-RDV conjugate Encapsulation efficiency

Preparation of a standard solution for the calibration curve was carried out by making remdesivir stock solution (332 μ M) by dissolving 5 mg of remdesivir into 10 mL of 10% DMSO. The solution was diluted to nine concentrations; 7,5 μ M; 10 μ M; 12,5 μ M; 15 μ M; 17,5 μ M; 20 μ M; 22,5 μ M; 25 μ M; 27,5 μ M; 30 μ M. Each solution was analyzed using a UV-Vis spectrophotometer at a wavelength of 246 nm. Therefore, the calibration curve with the relationship between absorbance (x) and concentration (y) was made and a linear regression equation is obtained.

The encapsulation efficiency of PAMAM-RDV conjugate was aimed to determine the % value of remdesivir encapsulation efficiency in the PAMAM. The remdesivir particle that did not encapsulate to the PAMAM cavity from the filtrate of the centrifugation ultrafiltration tube (MWCO: 10 kDa) were measured using UV-Vis spectrophotometer at the 246 nm wavelength. The % value of encapsulation efficiency is calculated by the equation: % Encapsulation efficiency = (RDV 1-RDV2)/ (RDV 1) x 100%

Where:

RDV1: The concentration of remdesivir in PAMAM-RDV conjugate

RDV2: The concentration of remdesivir in the filtrate of the centrifugation ultrafiltration tube

Particle size and zeta potential measurement

Particle size, distribution, and polydispersity index value were determined by Particle Size Analyzer NANO-ZS series Malvern Zetasizer (Malvern Instruments, Malvern, UK), using the photon correlation spectroscopy method. The conjugate solution was put into the analysis cuvette at a temperature of 25°C. The zeta potential of PAMAM-RDV conjugate was analyzed to determine the particle charge of the conjugate, the analysis was carried out using Particle Size Analyzer (Delsa Nano C type) with electrophoretic light scattering method. The sample was placed in a cuvette and analyzed at room temperature (25°C).

FTIR Spectrum of PAMAM-Remdesivir

The FTIR spectrum was analyzed by FTIR spectrophotometer 8400 S (Shimadzu, Japan). Analysis of conjugate PAMAM-RDV was carried out to identify the interaction of molecules, chemical structures, molecular bonds, and functional groups of the PAMAM-RDV conjugate. The samples were ground in the mortar with KBr



Fig. 1. The purification process of the conjugate. PAMAM RDV which retained on the top of membrane as retenate (a) and the free remdesivir which passed the membrane (b)

(250 mg) and inserted into the FTIR analysis ring to observed the spectra. The samples were tested at 4000 to 400 cm-1 wavenumber. The FTIR spectrum of the PAMAM-remdesivir conjugate was compared with the FTIR spectrum of pure remdesivir and PAMAM

Cellular uptake evaluation

Fluorescein isothiocyanate (FITC)remdesivir and FITC-PAMAM-RDV were prepared (1:1) by dissolved 1 mL of FITC (10 g/ mL) in 1 mL of PAMAM-RDV and remdesivironly solutions. These mixtures were incubated overnight in the dark at room temperature ^{11,12}. Vero cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM) at pH 7.4 at 37°C in 5% CO2. The cells were put into an 8-well chamber (10,000 cells per well) in 500 L of growth medium ^{11, 12}. The Vero cell medium was removed and washed with sterile PBS three times, then added the 4% formaldehyde, and incubated for 1 hour at room temperature. The formaldehyde was removed and the slide chamber was washed by sterile PBS three times. Samples (FITC-remdesivir, FITC-PAMAM-RDV, and FITC) were added to cells and incubated at 37°C with 5% CO2 for 15 minutes. The slide chamber was then washed with sterile PBS three times, dried and observed under a fluorescence microscope using a B-2A light filter to observed the cellular uptake ¹⁵.

RESULTS AND DISCUSSION

Synthesis of PAMAM-RDV conjugate

In order to maximally dispersed remdesivir, the remdesivir were diluted in 10% DMSO at low stirrer speed (250 rpm) for 10 minutes. The stock solution of remdesivir that has been prepared cannot be stored for more than 24 hours either at room temperature or in the refrigerator (4°C) because it will cause the remdesivir powder that has been dispersed became settle at the bottom of the



Fig. 2. The conjugate PAMAM-RDV (a) and Remdesivir-only (b) after 24 hours

solution. Thus, the prepared stock solution should be used for conjugate preparation on the same day.

The ratio concentration of remdesivir and PAMAM to synthesis conjugate PAMAM-RDV was 1:2. The conjugate was synthesized by dissolving 301 μ L remdesivir from the stock solution in 10 mL of WFI and followed by addition of 35 μ L PAMAM. Since PAMAM was not stable and easily degraded by light ¹⁰, the mixture of PAMAM and remdesivir was put in erlemeyer flask coated with aluminium foil. We optimeized the stirring time (at 150 rpm) for 3, 6, 12, 24, and 48 hours at room temperature. Optimization of stirring time is important to obtain the conjugate with the



Calibration Curve of Remdesivir

Fig. 3. The remdesivir's calibration curve



Fig. 4. The remdesivir's calibration spectrum in spectrophotometer UV-Vis at 246 nm

highest encapsulation efficiency of remdesivir. The result of encapsulation efficiency is show in Table 1.

In order to separate the non encapsulated remdesivir in PAMAM, the purification process of conjugate was performed. The purification was carried out using a centrifugation-ultrafiltration tube at molecular weight cut off (MWCO) 10 kDa). Since the molecular weight of remdesivir is less than PAMAM, thus the free remdesivir will pass the filtration tube. Purification carried out for seven cycles each at 5000 rpm 90 minutes until the volume of the filtrate no longer increased, as shown fig 1, resepectively. The PAMAM-RDV conjugate was detained at the top of the filter while the free molecule remdesivir at the bottom of the tube. The obtained PAMAM-RDV conjugate was in clear solution form (Fig 2).

Characterization of PAMAM-RDV conjugate Physical stability

In order to evaluate the PAMAM-RDV conjugate physical stability, the solution was kept at room temperature for 24 hours. The conjugate was observed in clear-stated condition however, at remdesivir-only solution, agglomeration was occurred after 24 h as shown at figure 2 (b) This phenomena explained the conjugate PAMAM-RDV maintained the dispersion of remdesivir in the solution as shown at figure 2 (a)

Entrapment efficiency

The calibration curve of remdesivir was made from nine series of concentrations; 7,5 μ M, 10 μ M, 12,5 μ M, 15 μ M, 17,5 μ M, 20 μ M, 22,5 μ M, 25 μ M, 27,5 μ M, and 30 μ M which were analyzed using a UV-Vis spectrophotometer at 246 nm¹². The obtained calibration curve's equation was: y= 0.0308x – 0.0222 (Fig 3.) where y is the concentration and x is the absorbance, and the R2 value was 0.993 (Fig 3.). The R2 value which is getting closer to 1 indicates that a high correlation between y and x. In addition, the R2 is also often used to describe data variation, where the desired data is linear ¹³.

CV= (Standar deviation)/Mean X 100%

After calculation, the variation coefficient is 1,5%, indicated the data variation is less and calibration curve is acceptable. The remdesivir calibration curve and calibration spectrum can be seen in Fig 3 and 4, respectively.

To determine the percentage of encapsu;ated remdesivir molecules in the PAMAM cavity, entrapment efficiency analysis was performed. The indirect method was carried out by measuring the concentration of free remdesivir from the filtrat of purification process as mentioned above. In the synthesize of the PAMAM-RDV

Table 1. The entrapment efficiency
of PAMAM-RDV conjugate

Stirring time (Hr)	EE (%)
3	37,969
6	51,101
12	56,500
24	69,082
48	48,435

Table 3. Zeta potential of PAMAM-RDV conjugates

Stirring time (Hr)	ζ Potensial
3	+1,143 mV
6	+3,410 mV
12	+3,063 mV
24	+23,07 mV
48	+2,083 mV

 Table 2. Particle size (Z-avg) and polydispersity index (PDI) of PAMAM-RDV conjugates

Stirring time (Hr)	Z-Avg	PDI	Dv90
3	1704,000	0,918	264,333
6	886,866	0,713	374,667
12	633,533	0,592	428,667
24	1008,100	0,730	399,333
48	7172,667	0,559	791,567

Table 4. Zeta potential of PAMAM-
RDV conjugates

Sample	ζ Potensial	
PAMAM Remdesivir	+21,7 mV -26,2 mV	

conjugate, the conjugate was stirred for 3, 6, 12, 24, and 48 hours using a shaker at room temperature at 150 rpm. The optimization of stirring time awas performed to get the highest entrapment efficiency of remdesivir. The results can be seen in Table 1.

As shown at table 1, 24 hours stirring time showed the highest entrapment efficiency of remdesivir, followed by the 6 hour, and then at the 48, 12, and 3 hours. This indicates that the maximum time required for remdesivir to enter and be entrapped into the PAMAM cavity was 24 hours at 150 rpm. The high entrapment efficiency of remdesivir at 24 h stirring time indicated that the amount of remdesivir encapsulated in the PAMAM cavity was greater than the unencapsulated. PAMAM dendrimers cavities able to encapsulate the hydrophobic drugs by covalent or non-covalent bond ⁷ However, it was observed that stirring for 48 hours caused the decreasing of entrapment efficiency. This phenomenone occured due to the possibility that PAMAM became unstable at room temperature for too long, thereby reducing its ability to entrap the remdesivir.

Particle size and particle distribution measurement

In this study, particle size (Z-avg) and polydispersity index (PDI) was measured using a Malvern type Particle Size Analyzer (PSA) with the photon correlation spectroscopy (PSC) method. PSC can measure the scattering of light by the Brownian motion of the dispersed particles ¹². Similar to the entrapment efficiency test, the





Zeta Potential Distribution



Fig. 6. The graphic of PAMAM's zeta potential

samples were PAMAM-RDV conjugates at 3, 6, 12, 24, and 48 hours of stirring. The results can be seen in Table 2, respectively.

Table 2 shows the largest Z-avg value was obtained at 48 hours and the smallest one was 12 hours of stirring. The smallest Dv90 was the 3 hours of stirring time (264,333 nm) which indicated the 90% of particles was below 264,333 nm. However, the PDI of 3 hours stirring time was 0,918. Polydispersity index or PDI which is also known as heterogeneity index was carried out to see the breadth or narrowness of the particle size distribution. PDI defines as a number on a scale from 0 to 1. A polydispersity index value less than 0.05 indicates a highly monodispersed particle or a narrow particle size distribution, while a broad particle size distribution is greater than 0,7^{14,15}, which indicated the 3 hours of stirring time obtained the largest variety of distribution particle.

Table 2 shows that the PAMAM-RDV conjugate has the lowest PDI which was stirred for 48 hours (0.559). However, the 48 hours of stirring time obtained the largest Dv90 and Z-avg, 0,559 and 7172,667 nm respectively. The 12 hours of stirring time obtained the good PDI (0,592) and the smallest Z-avg (633,533 nm). Nevertheless, the entrapment efficiency of 12 hours stirring

time was 56,5%, smaller than 24 hours of stirring time. The PDI of 24 hours stirring time is 0.730 which indicated the large variety of particle size distribution. However, this conjugate was still optimal because it had the largest number of remdesivir molecules among the others. Thus, conjugates with a stirring time of 24 hours are still acceptable even though the PDI was above 0, 7. **Zeta potential measurement**

The interaction energy between particles in a liquid can be determined by observing the zeta potential value and related to the stability of the particles ¹⁶. In this study, the zeta potential value of the PAMAM-RDV conjugate was measured using the Malvern type Particle Size Analyzer (PSA) with the electrophoretic light scattering (ELS) method. The zeta potential of PAMAM-RDV with various strring time shown at table 3 and zeta potential of PAMAM and remdesivir show at table 4 and Fig 4 and 5.

The zeta potential values of the PAMAM-RDV conjugates in Table 3 show that all the conjugates which were stirred for various hours had a positive charge. This due to the positive charge of amine group of PAMAM dendrimer ¹⁷. The conjugate stirred for 24 hours had the highest zeta potential value, which was +23.07 mV, while



(a) PAMAM-RDV conjugate; _____ (b) Remdesivir _____; and (c) _____ PAMAM



Fig. 8. The cellular uptake of control (a), remdesivir (b) and PAMAM-RDV (c)

the conjugate stirred for 3 hours had the lowest zeta potential value, which was ± 1.143 . Particles with a zeta potential value greater than ± 30 mV and less than ± 30 mV are believed to be strong cationic and anionic. Table 4 shows that remdesivir had a negative charge ($\pm 26,2$ mV) while PAMAM had a positive charge ($\pm 21,7$), therefore, it could be concluded that the remdesivir molecules were successfully entrapped in PAMAM because the surface charge of conjugates particles is positive. The presence of positive charge can enhance their binding to the cell surface with the negative charge by electrostatic interactions and enhance the conjugate cellular uptake 20 .

Zeta potential can be used as a parameter to predict the stability of a liquid because it is related to the interactions between particles in a colloidal system. High zeta potential values, both negative and positive, are generally stable because the charges between the particles tend to repel each other. On the other hand, a low zeta potential value, both positive and negative, will cause an attraction between the particles and cause particle aggregation in the liquid ¹⁶. In this study, the conjugate with the highest zeta potential was obtain from for 24 hours stirring time because it had a particle charge that was closest to +30 mV among other conjugates. Therefore, the best PAMAM-RDV conjugate was stirred for 24 hours. Most cellular membranes have a negative charge, thus particles that have positive zeta potential tend to permeate into the membrane. This will make the PAMAM-RDV conjugate enter the cells easier and increase their cellular uptake ¹⁸.

FTIR spectrum of PAMAM-RDV

FTIR spectrum analysis was carried out at wavenumbers 4000 to 400 cm⁻¹ in oder to compare the spectrum of the PAMAM-RDV conjugate with remdesivir and PAMAM. FTIR spectroscopy was used in this analysis because some molecules have strong absorbance in the infrared. The infrared spectrum provides information about molecular structure (peak position and peak intensity). Remdesivir, PAMAM, and PAMAM-RDV was mixed with KBr until it reaches the homogeneous state so the typical spectrum of the sample could be observed. KBr (potassium bromide) pellets are used because they are inert and transparent at wavenumbers 400 cm-1 and above ¹⁹

PAMAM dendrimer has a group of primary amine and secondary amine which are indicated by the presence of bands at wavenumbers

2080

3236 and 3448 cm⁻¹ as shown in Fig. 6 (c), while the presence of bands at wavenumbers 1014 and 1317 cm⁻¹ indicates that the presence of amide and alkane groups, while the wavenumbers at 1668 cm⁻¹ indicated the presence of a carbonyl group. This wavelength indicated the presence of the PAMAM dendrimer. While in the remdesivir (Fig. 6(b)), has a group of alkane which are indicated by the presence of bands at wavenumbers 1600-1680 cm⁻¹, while the presence of bands at wavenumbers 1690-1640 cm⁻¹ indicated the presence of imine group, while the wavenumbers at 1000-1350 cm⁻¹ indicated the presence of amine group, and the wavenumbers 1650-2000 cm⁻¹ indicates the presence of an aromatic group.

The infrared spectrum of the PAMAM-RDV conjugate (Fig. 6(c) has a band at wavenumbers in 1085 cm-1 which indicated the presence of the C-N from the amine group. Amines contained in the conjugate can be derived from remdesivir or PAMAM which has a band in the wavenumber in 1014-1317 cm⁻¹which also comes from the C-N group. In addition, there was broadband of PAMAM-RDV conjugate at a wave number in 3412 cm⁻¹from the N-H group present in both primary and secondary amines. The infrared spectrum of PAMAM and remdesivir also has a band in the area 3138 – 3448 cm⁻¹ which is also derived from the N-H group found in primary or secondary amines.

The infrared spectra of the PAMAM-RDV and remdesivir showed bands at wavenumbers of 1662 and 1647 cm⁻¹, shows that the two samples may have C=N groups of imine compounds. In addition, bands of the C-O or C-O-C groups of the ether compound were also seen at wavenumbers from 1010 to 1292 cm-1 in both the remdesivir sample and the PAMAM-RDV conjugate. Therefore, it could be concluded remdesivir has been encapsulated in PAMAM and formed PAMAM-RDV conjugates because there is a shift in wavenumbers from 3138 to 3431 cm⁻¹ and 3236 to 3448 cm⁻¹ from primary and secondary amine groups in remdesivir and PAMAM to 3412 cm⁻¹ derived from the PAMAM-RDV conjugate. In addition, a wavenumber of 3400 cm⁻¹ from PAMAM-RDV, shows that there is an O-H bond that allows hydrogen bonds to occur between remdesivir and PAMAM molecules in the conjugate.

Cellular uptake evaluation

To evaluate the ability of PAMAM-RDV conjugate to penetrate into the cells, Vero Cells which were normal cells derived from the kidney epithelial cells of African green monkeys was used. A flourescence microscope was used to observe the uptake of PAMAM-RDV in the Vero Cells, with 40 times magnification. The green colour came from the Fluorescein isothiocyanate (FITC), a stain which was conjugated to the samples.

The Intensity of Vero cell which were treated by FITC only, which was used as control, were showed at Figure 7 a (1) & (b), respectively. At Figures 7 b (1) and b (2), The intensity of FITC which were conjugated by Remdesivir was observed stronger compare to the control. However, the strongest intensity was obtained from FITC which were conjugate by the PAMAM-RDV (Fig. 7 c (1) & (2), respectively).

Generally, the membran cell as a barrier to separate the inside of the cell from the outside and block the particles to enter the cell. The cell membrane has a phospholipid bilayer and other proteins with a negative charge. This causes the membrane to selective permeability with particles. So, the particles that will enter the cell must pass through the cell membrane. The PAMAM-RDV conjugate has a positive charge from the PAMAM. Thus, the conjugate has the ability to bind the negatively charged plasma membrane, then cause pore formation and damage the lipids of the membrane ^{22, 23}.

Fig. 7 a show a weaker intensity of green colour compared to figures B and C in the absence of strong intensity in the centre of the cell. The control was FITC (in 1 mg/mL Dimethyl sulfoxide) which was incubated in cells for 15 minutes. The observed green color at control was due to the presence of Dimethyl sulfoxide which was used as a FITC solvent since it has the ability to penetrate into the cells, as reported previously²⁴. However, the intensity was still weaker than remdesivir and PAMAM-RDV. Thus, it can be concluded from cellular uptake testing that PAMAM-RDV has a stronger intensity than remdesivir.

CONCLUSIONS

In order to enhance remdesivir physical stability, we successfully synthesized PAMAM-

Remdesivir conjugate with the optimum strring time was 24 hours at 150 rpm, followed by ultrafiltration-purification process for 90 min at 5000 rpm. The characterization results showed the PAMAM-Remdesivir conjugate had positive charge of 23,07 mV, 1008 nm of particle size, 0,730 of PDI, and clear-stated solution even after 24 hours incubation. Those results suggested that PAMAM-Remdesivir has better physical stabillity compare to remdesivir only. Furthermore, PAMAM-Remdesivir particle had 69% entrapment efficiency and hydrogen bonds was formed between remdesivir and PAMAM molecules. Thus, indicated the successful remdesivir entrapment into PAMAM. In addition, the PAMAM-Remdesivir was observed had better cellular uptake in Vero Cell than remdesivir only as shown by its higher fluoresensi intensity.

Authors' contributions

All authors have contributed according to their duties and responsibilities.

Conflict of interests

No conflict of interests has been declared by all authors.

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