

Mucoxin (Acetogenin) Induce Expression of Pro-Apoptosis Proteins, Bax and p53, in T47D Breast Cancer Cells

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ABSTRACT

Though mucoxin has manufactured and widely marketed online, but indepth study of the role of these annonaceous substances on cellular level is still limited. This study aims to find out more about mucoxin anti-cancer effects, especially those associated with the expression of pro-apoptotic proteins, Bax and p53 in T47D breast cancer cell line. The T47D cell lines was divided into four groups based on the hour of mucoxin exposure until assays done, namely 0, 24, 48, and 72 hours. Concentration levels of mucoxin applied in the experiment were 0ng/ml, 0.1ng/ml, 0.5ng/ml, 1ng/ml, 5ng/ml and 10ng/ml. The bax gene expression assayed using quantitative PCR (qPCR) methods, whereas protein expression of bax and p53 determined by immunocytochemistry techniques. The results showed that in comparison with the control, both bax and p53 proteins increase by mucoxin treatment. It is suggested that mucoxin worthy classified as a promising anti-cancer drug.

Keywords: Mucoxin, acetogenin, p53, bax, pro-apoptosis protein, T47D cells, protein expression.

INTRODUCTION

Acetogenin is a class of compounds derived from Annonaceae plant family which recently intensively proposed as anti-cancer substances¹⁻³. Among various annonaceous acetogenin derivatives, mucoxin is the latest and claimed the most powerful to eradicate cancer cells because the only type acetogenin containing a hydroxylated trisubstituted tetrahydrofuran (THF) ring. Mucoxin is a highly potent and specific antitumor agent against number of human cancer cell lines such as A549 lung cancer⁴, MCF-7 (breast carcinoma) cell lines (ED₅₀ = 3.7 x10⁽⁻³⁾µg/mL compared to adriamycin, ED₅₀ = 1.0 x10⁽⁻²⁾µg/mL⁵, and epidermoid carcinoma, K562 and HL-60⁶.

Though mucoxin claimed as a promising anti-cancer, has manufactured and marketed on line all over the world, indepth study of the role of the bioactive materials on cellular level is still limited. In an effort to deepen understanding on the biological activity of mucoxin, Muhartono and colleagues from Indonesia has conducted a series of studies using a T47D breast cancer cell line. Among the results are as follows. Mucoxin effective in suppressing proliferation, increases apoptosis, suppress the expression of cyclin-D1 gene, as well as increasing the expression of the p53 gene in T47D breast cancer cells^{7,8}.

This work is another part of a research project conducted by Murtono and colleagues as an attempt to find out more about the effect of mucoxin

treatment on the expression of proliferation and apoptosis related proteins, p53 and bax, in T47D cells.

METHODS

The mucoxin and cell lines

Bioactive substances tested in this study was mucoxin (acetogenin) ID AG E 32919 and CASNo. 183195995 obtained from Angene International Limited. The product package contains 5mg of pure mucoxin in powder form. Whereas human breast cancer cell lines used in this study was T47D (ATCC®HTB133™) obtained from American Type Culture Collection (Manassas, VA 20108 USA) with a lot number 61062006.

Experimental design

For each protein assessment (p53 and Bax), a randomized block design with six concentrations of treatment and three replications was applied in this study. The T47D cell lines was divided into four groups based on the hour of mucoxin exposure until assays done, namely 0, 24, 48, and 72 hours. Concentration levels of mucoxin applied in the experiment are as follows: 0ng/ml, 0.1ng/ml, 0.5ng/ml, 1ng/ml, 5ng/ml and 10ng/ml.

Cell culture

The cells were grown in Roswell Park Memorial Institute medium (RPMI 1640) culture media supplemented with 10% Foetal Bovine Serum (FBS) Gibco™ (from Thermo Fisher Scientific Cat. No. 26140 079) and 0.2 units/ml bovine insulin (from Sigma Aldrich Cat. No. I5500 and CAS RN11070 73 8) at 37°C in 5% CO₂. Thawing process performed in water bath at 37°C for 2-4 minutes. Then, as much as 5x10⁴ cells/cm² was taken into T flask and incubated at 37°C in 5% CO₂. When cell density reached 80% confluent, trypsinization done using 0.25% Trypsin + 0.53 mM EDTA solution and then subcultured into new culture vessels, also at 37°C in CO₂ 5%. After two times passaging the T47D cells ready to be treated.

Mucoxin treatments

Culture media with a volume of 0.5 mL containing approximately 5,000 cells were added to a 24-well culture plate containing gelatin-coated coverslips and incubated for 24 hours. Once the

cells density reach 80% confluent the culture media from each well were removed and then serum-free medium and mucoxin of different concentrations in accordance with the treatment levels were added. The cells were then incubated in accordance with the length of hours that have been assigned to each group.

Gene expression assay

Because this study is a continuation of research that has been published, in which the gene expression of *p53* and *cyclin-D1* has been reported, then this section only explains the assessment procedure of the *bax* gene expression alone. Primers used for determining expression *bax* gene, refers to Porichi *et al.*⁹, was *forward primer* 5' GGACGAACTGGACAGTAACATGG 3', and *reverse primer* 5' GCAAAGTAGAAAAGGGCGACAAC 3'. As a control, β -actin gene with the forward primer 5'-TCTGGCACCACACCTTCTACAATG-3' and reverse primer 5'-AGCACAGCCTGGATAGCAACG-3' was used.

RNA was extracted from the T47D cell samples using easy-spin™ (DNA free) total RNA extraction kit from Intron Biotechnology. The concentration of total isolated RNA then assessed by NanoDrop 2000 Spectrophotometer from Thermo Scientific (Thermo Fisher Scientific Inc., MA, USA). Expression of *bax* genes in the T47D cells were determined by quantitative PCR (qPCR) methods using RealMOD™ Green Real-time PCR Kit from Intron Biotechnology with the Cat. No. REF25109. The qPCR data, finally, analyzed using Light Cycler® software from ROCHE. Beta actin (β -actin) gene was used as the internal control (housekeeping gene) while control samples was used as the calibrator genes.

Immunocytochemistry Assays

At the end of the incubation period, the cells were washed using PBS, then fixed in 4% paraformaldehyde. After being rinsed once with dH₂O, the fixed cells washed twice with PBS and then incubated for 5 minutes. Into each well was added 3% H₂O₂ in PBS at pH 7.4. After being washed three times with PBS and incubated for 5 minutes, blocking-buffer containing 0.25% Triton X 100 and 5% FBS was added and incubated at room temperature for one hour. After three time wash with

PBS and 5 minutes incubation, into each well was added primary antibody with the dilution of 1:100 in PBS with 1% FBS. The primary antibody used for assaying *p53* and *Bax* protein consecutively are p53 protein (wt-p53) polyclonal antibody (*Rabbit IgG anti p53*) from Bioss USA with the Cat. Num. bs 0033R and anti-Bax polyclonal antibody (*Mouse IgG anti Bax*) from Bioss USA with the Cat. Num. bs 0127M.

After overnight incubation at 4°C, the cells then washed three times with PBS and incubated for 5 minutes. Into each well was added the secondary antibody, *UltraTek HRP Anti-Polyvalent (DAB)* from SkyTex Laboratories with the Cat. Num. AMF080 IFU incubated at room temperature for 2 hours. After

horseradish peroxidase (HRP) was added the cells were incubated for 40 minutes. After being washed with PBS three times, into the preparates was added 3,3,2 -diaminobenzidine (DAB), incubated for 40 minutes, and washed with dH₂O.

Finally, cells were counterstained with Meyer's hematoxyline (Dako Cytomation, Denmark) and then washed with dH₂O. Coverslips removed from the wells and then mounted on the deck glass coated with 5% gelatin. The results were visualized using light microscope Nikon E 100 and photographed using Nikon Coolpix 4500. To quantify protein expression of p53 and bax, software Image-Pro Plus (IPP) was used. The quantity of protein

Tabel 1: Expression of *Bax* gene (number of copies) in T47D cancer cells treated with mucocin with different concentrations at different exposure times

Exposure Time	Concentration of Mucocin	Expression (mean±SD)	Anova (P value)
0 h	0 ng/ml	15.857±1.163 ^a	0.007
	0.1 ng/ml	10.923±0.351 ^a	
	0.5 ng/ml	26.967±2.957 ^{ab}	
	1 ng/ml	32.980±1.716 ^{bc}	
	5 ng/ml	32.533±1.137 ^{bc}	
	10 ng/ml	35.233±1.518 ^c	
24 h	0 ng/ml	27.410±0.810 ^{ab}	0.011
	0.1 ng/ml	18.500±3.143 ^a	
	0.5 ng/ml	35.100±2.078 ^b	
	1 ng/ml	38.047±1.570 ^c	
	5 ng/ml	33.334±0.808 ^b	
	10 ng/ml	36.813±2.161 ^{bc}	
48 h	0 ng/ml	38.383±0.616 ^a	0.0001
	0.1 ng/ml	39.427±1.116 ^{ab}	
	0.5 ng/ml	33.550±1.496 ^a	
	1 ng/ml	52.477±1.390 ^b	
	5 ng/ml	66.667±1.896 ^c	
	10 ng/ml	47.500±1.212 ^b	
72 h	0 ng/ml	39.580±1.076 ^a	0.012
	0.1 ng/ml	38.747±2.604 ^a	
	0.5 ng/ml	29.840±1.465 ^a	
	1 ng/ml	31.453±1.439 ^a	
	5 ng/ml	31.077±0.901 ^a	
	10 ng/ml	53.007±2.650 ^b	

Mean±SD values in the same hour group followed by the same superscript are not different at $\alpha=0.05$ by LSD test.

expression in each treatment was calculated by summing the number of brownish product arising from the reaction between HRP and DAB in five fields of view and then divided by the number of replicates ($n = 3$).

Statistical Analysis

Comparison of mean values between treatment are presented as mean \pm SD and analyzed using ANOVA followed LSD test. A p -value of less than 0.05 was considered to be statistically significant for ANOVA and $\alpha < 0.05$ was considered to be statistically significant for LSD test.

RESULTS

Effects of Mucoxin on Gene Expression of Bax

Effects of mucoxin treatment on bax gene expression in T47D cells exposed to mucoxin with

different concentration at hour 0, 24, 48, and 72 are presented in Table 1. Compared to control (0 ng/ml), though not in a consistence manner, mucoxin treatment (0.1 ng/ml – 10 ng/ml) effect on expression of bax gene in all hour groups, even in cell group that were exposed to mucoxin with the duration of less than one hour (0 h).

Effects of Mucoxin on Expression of Bax Protein

Visualization of bax protein expression in T47D breast cancer cells exposed to mucoxin with different concentration at hour 0, 24, 48, and 72 was depicted in Fig.1. The results of quantification of the bax protein expression using the IPP software are presented in Table 2. In contrast to its effects on p53, effects of mucoxin treatment on bax protein expression just evident at 48 hours ($p = 0.019$) and 72 hours ($p = 0.0001$). In addition, referring to the results of LSD test ($\alpha = 0.05$), the highest

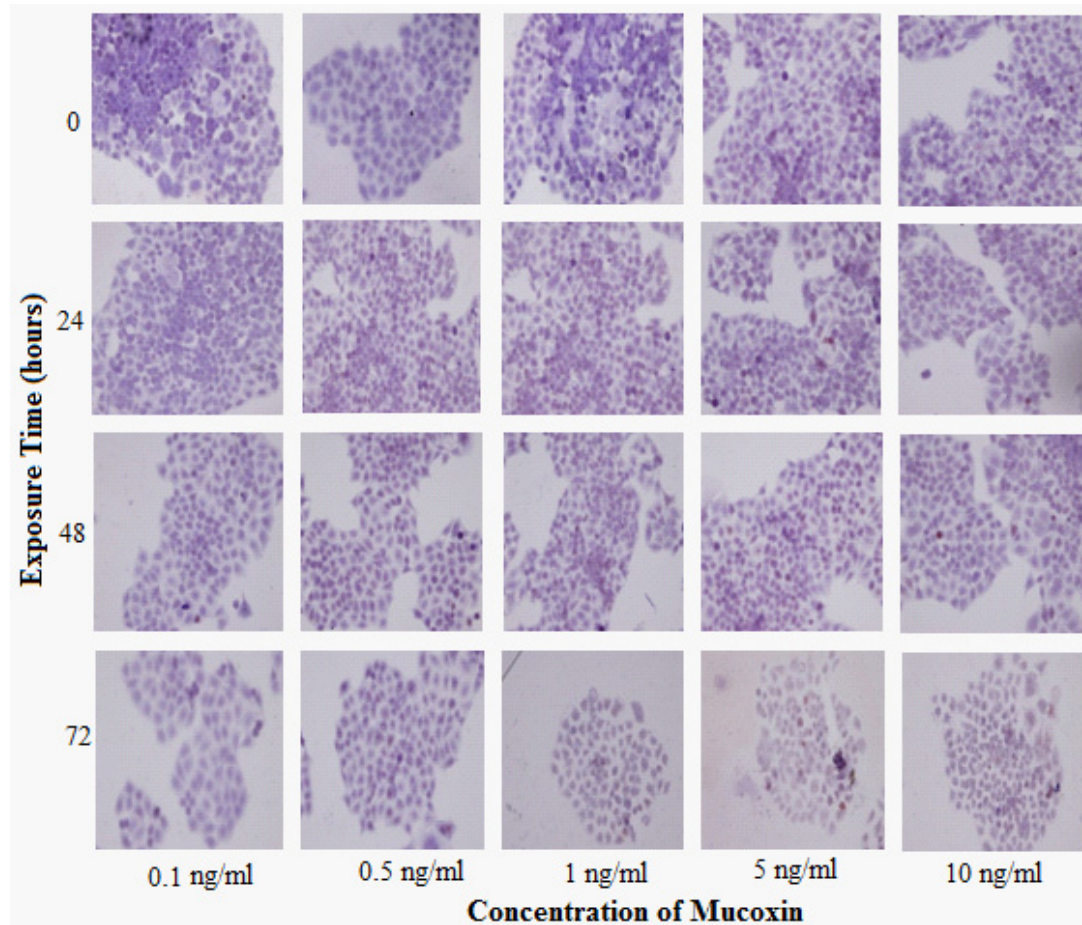


Fig.1: Immunocytochemical expression of *bax* protein in T47D cells treated with mucoxin with different concentrations at different exposure times

concentration of mucoxin (10 ng/ml) shows a highest expression of bax protein.

Effects of Mucoxin on Protein Expression of p53

Immunocytochemical expression of p53 protein in T47D breast cancer cells exposed to mucoxin with different concentration at hour 0, 24, 48, and 72 were visualized in Fig.2. The quantification of the p53 protein expression using IPP software resulted in the data presented in Table 3. Referring to the *p*-value of Anova in the table, it appears that mucoxin already showing effect within 24 hours (*p* = 0.001), and continues to increase at 48 hours (*p* = 0.045) and 72 hours (*p* = 0.009). In all groups of hours, as compared with the control group (0 ng/ml),

mucoxin of all concentrations significantly increased the expression of p53 protein ($\alpha = 0.05$). At the end of hour 72, the effect of mucoxin from the lowest (0.1 ng/ml) to the highest (10 ng/ml) is significantly different, the higher the concentration of mucoxin given the higher expression of p53 protein ($\alpha = 0.05$).

DISCUSSION

As has been previously reported that in the T47D breast cancer cells mucoxin shown to increase p53 gene expression and decreasing gene expression of cyclin-D1⁷, in addition mucoxin also proven to decrease proliferation and increase apoptosis of the cell lines⁸. In current study it was

Tabel 2: Expression level of Bax protein in T47D breast cancer cell treated with mucoxin with different concentrations at different exposure times

Exposure Time	Concentration of Mucoxin	Expression (mean±SD)	Anova (P value)
0 h	0 ng/ml	1,333±0,577 ^a	0.,453
	0.1 ng/ml	2,000±1,000 ^a	
	0.5 ng/ml	1,333±1,528 ^a	
	1 ng/ml	2,667±0,577 ^a	
	5 ng/ml	2,333±1,528 ^a	
	10 ng/ml	2,667±0,577 ^a	
24 h	0 ng/ml	2,333±2,517 ^a	0.164
	0.1 ng/ml	3,667±0,577 ^a	
	0.5 ng/ml	5,000±1,000 ^a	
	1 ng/ml	4,333±1,155 ^a	
	5 ng/ml	4,667±1,155 ^a	
	10 ng/ml	5,667±1,528 ^a	
48 h	0 ng/ml	2,000±2,000 ^a	0.019
	0.1 ng/ml	5,667±0,577 ^b	
	0.5 ng/ml	6,000±1,000 ^b	
	1 ng/ml	7,333±0,577 ^{bc}	
	5 ng/ml	7,333±1,528 ^{bc}	
	10 ng/ml	8.333±0.577 ^c	
72 h	0 ng/ml	2.000±1.000 ^a	0.0001
	0.1 ng/ml	15.000±4.583 ^b	
	0.5 ng/ml	14.333±3.055 ^b	
	1 ng/ml	29.000±6.557 ^c	
	5 ng/ml	30.0001±4.359 ^c	
	10 ng/ml	28.333±2.517 ^c	

Mean±SD values in the same hour group followed by the same superscript are not different at $\alpha=0.05$ by LSD test.

found that mucoxin treatment induce expression of bax gene (Table 1), promote expression of bax protein (Fig.1 and Table 2) as well as p53 protein (Fig.2 and Table 3).

Increased expression of Bax by mucoxin can be explained by several mechanisms. Mucoxin allegedly can activate, stabilize, and accumulate Bax in the mitochondria to function as an proapoptosis agent. Annonacin, is one type of acetogenin, as reported by Yuan *et al.*¹⁰, can cause cell cycle stops at the G1 phase and causes cytotoxic through Bax and caspase pathway. Bax is a proapoptosis protein that promotes cell death upon the presence of stress stimuli, that in this study the stress likely caused by mucoxin.¹¹

In cancer cells, apoptosis induction was correlated with inhibition of bax degradation¹². The

increase in the degradation of Bax is one way for cancer cells to survive¹³. Whereas in cancer cells treated with mucoxin, bax will be oligomerized easily in mitochondrial membrane that make the protein more stable. All bax molecules become oligomerized after membrane insertion and are present at pore edges. Such oligomerization may lead to membrane deformation that promote pore enlargement¹⁴.

Due to pores enlargement, mitochondrial membrane become more permeable and release mitochondrial *cytochrome c*, Smac and AIF into the cytosol and activation of caspase-mediated apoptosis¹⁵. Next, *cytochrome c* binds to the apoptosis-activating factor 1 (Apaf-1) and *procaspase 9* forms apoptosom which will activate *caspase* pathway leading to apoptosis¹⁶. Given mucoxin belonged to the annonaceous acetogenin family, then there is no harm if mucoxin analogized with other acetogenin

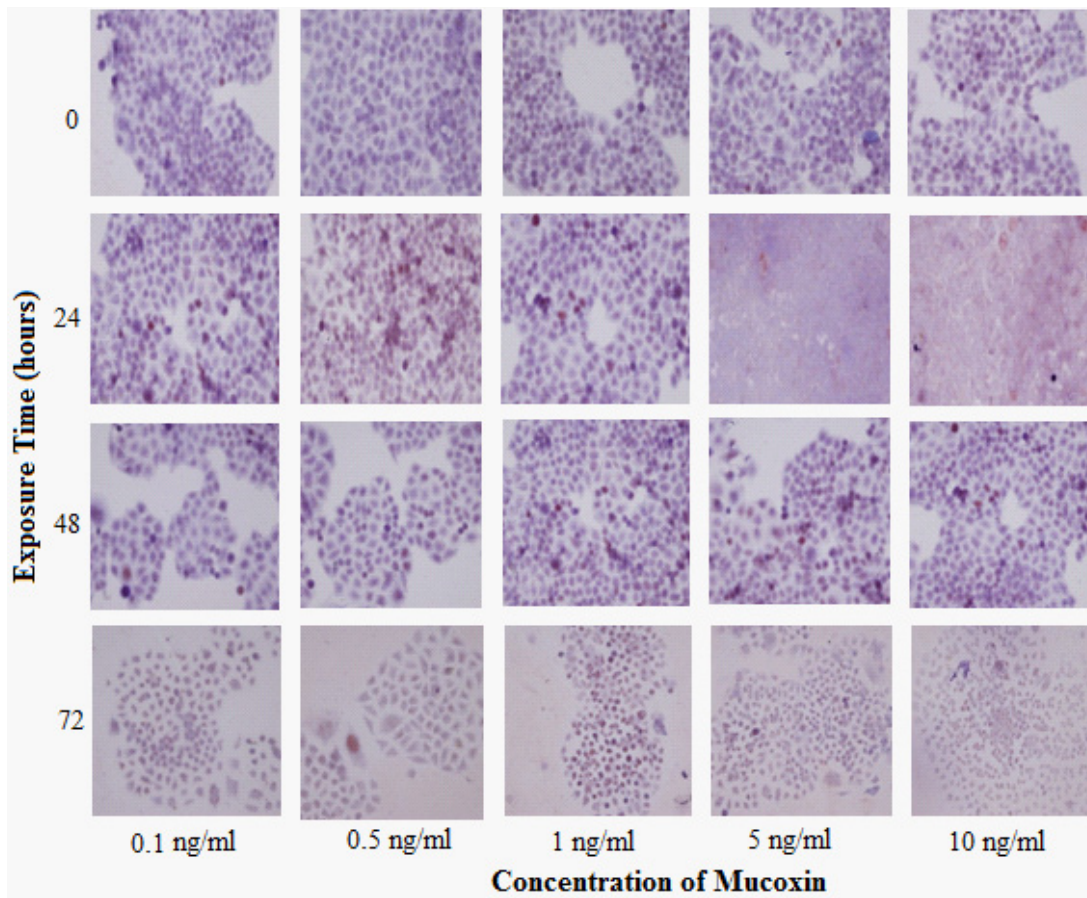


Fig. 2: Immunocytochemical expression of p53 protein in T47D cells treated with mucoxin with different concentrations at different exposure times

such as bullatacin. Bullatacin, as indicated by Chen *et al.*¹⁷, is an acetogenin substances that was able to increase of reactive oxygen species (ROS). Accumulation of ROS in cancer cells will activate Bax and caspase pathway which will further initiate apoptosis¹⁸. In short, the pro-apoptotic protein Bax commits a cell to death by permeabilizing the mitochondrial outer membrane (MOM)¹⁹.

As has been shown in Fig.2 and Table 3, all concentration of mucoxin treatment significantly increase expression of p53 gene. However there is little study that could explain satisfactorily about mucoxin mechanisms in inhibiting the proliferation and increasing apoptosis whether through increased

stabilization and activation of p53. Yuan *et al.*²⁰ from Taiwan, by testing cytotoxicity effects of annonacin on prostate cancer cells concluded that annonacin can arrest cell cycle at phase S1 depended on *caspase 3* and bax. In Indonesia, Rachmawati *et al.*²¹ has also managed to identify that activation and stabilization of p53 play a key role in apoptosis of cervical cancer cells. The activated p53 will act as a regulatory protein that triggers a variety of biological responses mainly in the process of cell proliferation and apoptosis²².

When p53 is active, there will be phosphorylation at one or more serine residues on the N-Terminus or C-Terminus which would

Tabel 3: Expression level of p53 protein in T47D breast cancer cell treated with mucoxin with different concentrations at different exposure times

Exposure Time	Concentration of Mucoxin	Expression (mean±SD)	Anova (P value)
0 h	0 ng/ml	4.333±1.528 ^a	0.565
	0.1 ng/ml	4.333±0.577 ^a	
	0.5 ng/ml	3.667±0.577 ^a	
	1 ng/ml	5.000±1.000 ^a	
	5 ng/ml	4.333±0.577 ^a	
	10 ng/ml	4.333±0.577 ^a	
24 h	0 ng/ml	2.333±2.017 ^a	0.001
	0.1 ng/ml	8.333±1.528 ^b	
	0.5 ng/ml	13.333±3.055 ^{bc}	
	1 ng/ml	11.667±3.786 ^{bc}	
	5 ng/ml	13.667±3.055 ^{bc}	
	10 ng/ml	14.000±2.000 ^c	
48 h	0 ng/ml	3.333±3.055 ^a	0.045
	0.1 ng/ml	13.333±4.041 ^b	
	0.5 ng/ml	15.333±2.082 ^{bc}	
	1 ng/ml	14.333±3.215 ^{bc}	
	5 ng/ml	17.333±1.528 ^{bc}	
	10 ng/ml	18.333±0.577 ^c	
72 h	0 ng/ml	5.000±3.606 ^a	0.009
	0.1 ng/ml	18.333±2.082 ^b	
	0.5 ng/ml	34.333±3.055 ^c	
	1 ng/ml	79.000±26.514 ^d	
	5 ng/ml	86.667±27.429 ^d	
	10 ng/ml	145.000±72.959 ^e	

Mean±SD values in the same hour group followed by the same superscript are not different at $\alpha=0.05$ by LSD test

then be bound to enhancer elements that are the downstream targets of p53²³. The downstream targets of p53 will then respond to the binding by self-activation or not²⁴. Until now it has known hundreds of p53 target genes with various functions and cellular mechanisms. In general, the p53 target genes are grouped in three main pathways as the downstream activation of p53 including cell cycle arrest when DNA damage occurs, repair of damaged DNA, and control of apoptosis when the damage can not be repaired anymore²⁵.

In relation to Bax protein, Maximov and Maximov²⁶ suggested that the p53 tumor-suppressor protein can intervene at every major step in apoptotic pathways. As indicated by Schuler *et al.*²⁷ that p53 activates the apoptotic machinery through induction

of the release of cytochrome c from the mitochondrial intermembrane space. The release of cytochrome c is facilitated, of course, by pore enlargement of the mitochondrial membrane as the Bax activity.

Based on the fact that mucoxin significantly increases gene expression and protein expression of Bax as well as p53 protein, wherein both proteins are the main factors in the apoptosis of cancer cells, it can be concluded that mucoxin deserves to be called an anti-cancer substance.

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