

Antifungal activity of the essential oil of *Cinamomum zeynalicum* on *Candida albicans*, *Aspergillus niger* and *Aspergillus flavus*

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ABSTRACT

The composition of the essential oil of *Cinamomum zeynalicum* and its antifungal activity on *Candida albicans*, *Aspergillus niger* and *Aspergillus flavus* fungal strains were studied in Iran. Essential oil from the bark parts of the plant was obtained by hydrodistillation and analysed by GC and GC-MS. The MIC was used to evaluate the antifungal activity against *Candida albicans* ATCC 10231, *Aspergillus niger* ATCC 9642 and *Aspergillus flavus* ATCC 9643. Antifungal activity was evaluated for the essential oil and simultaneously for Amphotricin B. Results showed that *Cinamomum zeynalicum* essential oil exhibited a significant activity against fungi, and its MIC on *Candida albicans*, *Aspergillus niger* and *Aspergillus flavus* were respectively 0.125, 0.125 and 0.125 $\mu\text{g ml}^{-1}$ (ppm). The present study indicates that *Cinamomum zeynalicum* essential oil has considerable antifungal activity, deserving further investigation for clinical applications.

Key words: *Cinamomum zeynalicum*, MIC(minimal inhibitory concentration), Antifungal activity.

INTRODUCTION

Fungal infections have been increasing in recent years due to a growing number of high-risk patients, particularly immunocompromised hosts. *Candida* is the third- or fourth-most-common isolate in nosocomial bloodstream infections in the USA. In addition, The mortality rate due to invasive aspergillosis increased by 357 % between 1980 and 1997 in the USA .In spite of the introduction of new antifungal drugs, they are limited in number. The increase of fungal resistance to classical drugs, the treatment costs, and the fact that most available antifungal drugs have only fungistatic activity, justify the search for new strategies (Eugenia pinto, 2006). Aromatic plants have been widely used in folk medicine. It is known that most of their properties are due to their volatile oils. Essential oils from many

plants are known to possess antifungal activity, but only limited information exists about activity toward human fungal pathogens. They have been empirically used as antimicrobial agents, but the mechanisms of action are still unknown.

Some essential oils show an important antifungal activity against yeasts, dermatophyte fungi and *Aspergillus* strains, which could predict therapeutic benefits, mainly for diseases with mucosal, cutaneous and respiratory tract involvement. (Pina-Vaz *et al.*, 2004; Salueiro *et al.*, 2003, 2004),

The objective of our present research was to evaluate the antifungal activity and investigate the mechanism of action of *Cinamomum zeynalicum* essential oil from Iran.

METHOD

Essential oil analysis

Essential oil from the bark parts of the plant was obtained by hydrodistillation. Cinnamon sticks were finely chopped before being boiled, and analysed by GC and GC-MS. Gas chromatography (GC) and GC-mass spectrometry (MS) analysis of essential oil were performed.

Plant material and chemicals

Bark parts of the plants were collected from Tokestan, 11th Km. Gorgan-Mashhad Road, Gorgan, Iran.

Essential oil analysis

Essential oil was isolated by water distillation for 3 h from air-dried material, using a Clevenger-type apparatus, according to the procedure described in the European Pharmacopoeia (Council of Europe, 1997).

Gas chromatography (GC-FID)

Gas chromatography analysis was performed on a Hewlett-Packard Model 5890 Series II gas chromatograph equipped with flame ionization detector and capillary column HP-101 (Methyl silicone fluid), 25 m × 0.2 mm i.d., coating thickness 0.2 µm. Chromatographic conditions were as follows: helium as carrier gas at 1.0 ml min⁻¹; injector and detector temperatures, 250°C and 300°C. Oven temperature was isothermal at 70°C for 2 min, then increased to 200°C, at a rate of 3°C min⁻¹ and held isothermal for 15 min. Volume injected 1 µl. Split ratio 1 : 50.

Gas chromatography-mass spectrometry

Essential oil was also analysed by Hewlett Packard GC-MS (model 5890 series II) with mass selective detector (model 5971A). Two columns of different polarity were used: an HP-101 column (Methyl silicone fluid, Hewlett Packard; 25 m × 0.2 mm i.d., film thickness 0.2 µm) and an HP-20M column (Carbowax 20M, Hewlett Packard; 50 m × 0.2 mm i.d., film thickness 0.2 µm). Oven temperature was programmed as follows: isothermal at 70°C for 4 min, then increased to 180°C, at a rate of 4°C min⁻¹ and subsequently held isothermal for 15 min (for HP-20M column); isothermal at 70°C for 2 min, then increased to 200°C, at a rate of 3°C min⁻¹ and held isothermal for 15 min (for HP-101 column). Carrier gas was helium, flow rate: 1 ml min⁻¹; injector temperature: 250°C; volume injected: 1 µl; split ratio: 1 : 50. MS conditions: ionization voltage: 70 eV; ion source temperature: 280°C; mass range: 30–300 mass units.

Qualitative and quantitative determination

The individual peaks were identified by comparison of their retention indices to those of authentic samples, as well as by comparing their mass spectra with the Wiley 6-0 library (Wiley, New York, NY, USA) and NIST98 (National Institute of Standards and Technology, Gaithersburg, MD, USA) mass spectral database and literature (Adams 1995).

The percentage composition of the samples was computed from the GC peak areas by using the normalization method (without correction factors). Quantitative results are mean of data derived from duplicate GC-FID analyses.

Table 1. Antimicrobial activity (MIC) and (MFC) of the essential oil of the *Cinamomum zeynalicum* for *Candida albicans*, *Aspergillus niger* and *Aspergillus flavus*

Results were obtained from three independent experiments performed in duplicate			
<i>Candida albicans</i> ,	<i>Aspergillums niger</i>	<i>Aspergillus flavus</i>	<i>Cinamomum zeynalicum</i> effects
0.125	0.125	0.125	MIC in µg ml ⁻¹
0.25	0.25	0.25	MFC in µg ml ⁻¹

Isolation and detection of fungi

The antifungal activity of *Cinamomum zeynalicum* essential oil was evaluated against *Candida albicans* ATCC 10231, *Aspergillus niger* ATCC 9642, and *Aspergillus flavus* ATCC 9643. The fungal isolates were identified by standard microbiology methods and stored in Sabouraud dextrose broth with glycerol at -70°C .

Fungi were plated on Sabouraud 2% (w/v) glucose agar (SGA), and incubated at $25 \pm 2^{\circ}\text{C}$ for the 5–7 days.

For the antifungal activity testing, *Cinamomum zeynalicum* essential oil was dissolved in 96% (v/v) ethanol and then diluted with 30% (v/v) ethanol in distilled water with 0.1% (w/v) Tween 80. Final concentrations of the essential oil was 2% (w/v).

Antifungal activity testing by dilution method

Minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) were determined by using the serial broth dilution method described by Pepeljnjak *et al.* (2003). Serial twofold dilutions in DMSO, ranging from 0.02 to $20 \mu\text{l ml}^{-1}$, were tested for essential oil. In addition, the reference antifungal compounds, fluconazole (Pfizer) for *Candida albicans* or amphotericin B (Sigma) for *Aspergillus*, were used as standard antifungal drugs. Twofold serial dilutions ranging from 0.25 to $128 \mu\text{g ml}^{-1}$ for fluconazole and 0.016 to $16 \mu\text{g ml}^{-1}$ for amphotericin B were used. Quality control determinations of the MICs of fluconazole and amphotericin B were performed by testing *C. parapsilosis* ATCC 90018 and *C. krusei* ATCC 6258. The results obtained were within the recommended limits.

MIC is defined as the lowest concentration of extract or essential oil that allows no more than 20% growth of the fungus, visualized as a reduced number of colonies after removing the loop with approx. $10 \mu\text{l}$ of each dilution, and then inoculated on SGA and incubated at $25 \pm 2^{\circ}\text{C}$ for 7 days. MFC is defined as the lowest concentration of essential oil that completely inhibited the growth of fungi. These experiments performed in duplicate were repeated independently three times and yielded

essentially the same results. A range of values is presented where different results were obtained. Two growth controls, RPMI medium and RPMI with 2.0 % (v/v) DMSO, were included for each strain.

Statistics

The data obtained as MIC and MFC of essential oil, expressed in $\mu\text{g ml}^{-1}$, were statistically analysed by using the Wilcoxon matched pairs test. The level of $P < 0.05$ was considered statistically significant.

RESULTS AND DISCUSSION

The oil was obtained from air-dried plant material in a yield of 1.8 % (w/w).

Evaluation of MIC showed that the oil was active against all the tested strains (Table 1). *Cinamomum zeynalicum* essential oil exhibited significant antifungal activity. MIC value $0.125 \mu\text{l ml}^{-1}$ against *Aspergillus* strains and *Candida*. It is difficult to attribute the activity of a complex mixture to particular constituents. The importance of the phenolic hydroxyl groups for the antimicrobial activity of the monoterpenoids has previously been reported (Adam *et al.*, 1998; Alianni *et al.*, 2001; Dorman & Deans, 2000; Nostro *et al.*, 2004; Sivropoulou *et al.*, 1996).

In conclusion, the findings of the present study indicate that *Cinamomum zeynalicum* essential oil has potential as a topical antifungal agent against fungi that are pathogenic to humans. This essential oil is a broad-spectrum agent that inhibites *Aspergillus* and *Candida* species,

Given the results described above, particularly the possible mechanisms of action, which might induce side-effects in humans, these antifungals require further investigation.

The results presented should stimulate studies on toxicity, improved formulations and the determination of optimal concentrations for clinical applications, as well as comparative studies alongside currently used drugs of the therapeutic efficacy of essential oils to control infections.

REFERENCES

1. Adam, K., Sivropoulou, A., Kokkini, S., Lanaras, T. & Arsenakis, M., Antifungal activities of *Origanum vulgare* subsp. *hirtum*, *Mentha spicata*, *Lavandula angustifolia*, and *Salvia fruticosa* essential oils against human pathogenic fungi. *J Agric Food Chem* **46**: 1739–1745 (1998).
2. Adams, R. P., *Identification of Essential Oil Components by Gas Chromatography/Mass Spectroscopy*. Carol Stream, IL: Allured Publishing Corporation (1995).
3. Aligiannis, N., Kalpoutzakis, E., Mitaku, S. & Chinou, I. B., Composition and antimicrobial activity of the essential oils of two *Origanum* species. *J Agric Food Chem* **38**: 4168-4170 (2001).
4. Council of Europe Methods of Pharmacognosy. In *European Pharmacopoeia*, 3rd edn, pp. 121–122. Strasbourg: European Department for the Quality of Medicines (1997).
5. Dorman, H. J. & Deans, S. G., Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *J Appl Microbiol* **88**: 308-316 (2000).
6. Nostro, A., Blanco, A. R., Cannatelli, M. A., Enea, V., Flamini, G., Morelli, I., Roccaro, A. S. & Alonzo, V., Susceptibility of methicillin-resistant staphylococci to oregano essential oil, carvacrol and thymol. *FEMS Microbiol Lett* **230**: 191–195 (2004).
7. Pepeljnjak, S., Kosalec, I., Kalodera, Z. and Kuštrak, D., Natural antimycotics from Croatian plants. In *Plant-Derived Antimycotics* ed. Rai, M. and Mares, D. pp. 49–81. *Binghamton*: The Haworth Press (2003).
8. Pina-Vaz, C., Rodrigues, A. G., Pinto, E., Costa-de-Oliveira, S., Tavares, C., Salgueiro, L. R., Cavaleiro, C., Gonçalves, M. J. & Martinez-de-Oliveira, J., Antifungal activity of *Thymus* oils and their major compounds. *J Eur Acad Dermatol* **18**: 73-78 (2004).
9. Pinto, E., Pina-Vaz, C., Salgueiro, L. R., Gonçalves, M. J., Costa-de-Oliveira, S., Cavaleiro, C., Palmeira, A., Rodrigues, A. & Martinez-de-Oliveira, J., Antifungal activity of the essential oil of *Thymus pulegioides* on *Candida*, *Aspergillus* and *dermatophyte species*. *J Med Microbiol* **55**: 1367-1373 (2006).
10. Salgueiro, L. R., Cavaleiro, C., Pinto, E. & 7 other authors., Chemical composition and antifungal activity of the essential oil of *Origanum virens* on *Candida* species. *Planta Med* **69**: 871-874 (2003).
11. Salgueiro, L. R., Pinto, E., Gonçalves, M. J. & 7 other authors. Chemical composition and antifungal activity of the essential oil of *Thymbra capitata*. *Planta Med* **70**: 572-575 (2004).
12. Sivropoulou, A., Papanikolaou, E., Nikolaou, C., Kokkini, S., Lanaras, T. & Arsenakis, M. Antimicrobial and cytotoxic activities of *Origanum* essential oils. *J Agric Food Chem* **44**: 1202-1205 (1996).