

# Comparative Assessment of Sensitivity and Specificity of Rose Bengal Test and Modified In-house ELISA by using IS711 TaqMan Real Time PCR Assay as a Gold Standard for the Diagnosis of Bovine Brucellosis

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Serological approaches such as Rose Bengal Test (RBT) and enzyme linked immunosorbent assay (ELISA) are common tests, however they are generally not sensitive or specific enough for diagnosis of *Brucella* because of cross-reactivity with different bacterial antigens. The work objected to evaluate the sensitivity and specificity of rose Bengal and modified in-house ELISA using IS711 real time PCR as a gold test to detect *Brucella* in calves sera. Two hundred and thirty (n=230) blood samples were collected from (2-3) years asymptomatic male calves in two Egyptian abattoirs. Rose Bengal test (RBT) and modified in-house ELISA were applied to determine the seroprevalence of brucellosis in abattoirs animals while quantitative Taqman real-time PCRs (RT-PCR) were implemented for the identification of *Brucella* genus. The overall prevalence of brucellosis was (53.9 %), (75.2 %) and (79.1 %) as determined by ELISA, RBT and RT- PCR assays respectively. Regarding statistical analysis of the reported data and considering real time PCR the gold standard, the RBT recorded a sensitivity of (79.12%) and a specificity of (39.58 %) with an accuracy of (70.87%). While (83.24%) was reported as positive predictive value and (33.33 %) as a negative predictive value. The sensitivity and specificity of ELISA were (55.49%) and (52.08 %) respectively while the accuracy was (54.78%). Positive predictive value and negative predictive value for ELISA were determined as (81.45%) and (23.58 %) respectively. RBT can be routinely used for diagnosis of *Brucella* as cost effective , more sensitive and accurate test than ELISA However, real time PCR is highly recommend as gold test for identification and differentiation of bovine brucellosis.

**Keywords:** Brucellosis, Rose Bengal Test, ELISA, Sensitivity, Specificity.

Brucellosis is a global zoonosis induced by bacteria of the genus *Brucella*, and considered as a crucial public health issue across the world. The great variety of clinical evidences of the human brucellosis make the diagnosis relatively difficult<sup>1, 2, 3</sup> This absence of certain symptoms makes it challenging to differentiate brucellosis from numerous febrile diseases that commonly

appear in the same regions so that laboratory testing is very important for diagnosis<sup>4</sup>. Among these tests, exclusively the isolation of the organism presents an absolute evidence of infection however bacteriological diagnosis is costly and even hazardous. Alternatively, serological tests are simpler to apply and a significant assistance in diagnosis. Direct agglutination, Rose Bengal



test, Coombs test and ELISA are the most desired methods for the serological diagnosis of brucellosis<sup>5,6,7</sup>.

Serological tests are commonly employed to perform several epidemiological researches as well as diagnostic applications, but there is no ideal serological test<sup>8</sup>. However, the diagnostic efficiency and discriminative capacity of a test are usually assessed by comparing the sensitivity and specificity of different tests analytically<sup>9</sup>. The diagnostic capability of a test could be evaluated by comparison with standard reference test and examined by gold standards<sup>10</sup>. Molecular diagnosis of brucellosis by PCR techniques has often been utilized as an additional alternative. Genus-specific PCR assays are relatively inexpensive diagnostic tests for screening with the possibility to recognize the causative agent even with minimal concentrations of DNA<sup>11</sup>. The objective of this study was to evaluate diagnostic effectiveness and discriminative power of Rose Bengal Plate Test (RBT), and in-house indirect enzyme linked immunosorbent assay (I-ELISA) tests utilized for diagnosis of bovine brucellosis in Egypt choosing real time PCR as the gold test. This study is amongst a type in the context of field diagnostic test evaluation for bovine brucellosis in Egypt which has considerable importance for disease surveillance and long term control plans.

## MATERIAL AND METHODS

### Ethical statement

The protocol for collection of animal materials was approved by districts veterinary and agricultural authorities. Sampling process was carried out during the daily slaughtering work on the slaughtering panel so that no IRB was required.

### Blood sample and Serum separation

A total of (230) blood sample were collected from the main abattoir of the region throughout a period of 15 months. Labeled sterile vacutainers were used to collect blood from animals at the time of slaughter (on the slaughtering panel) so no IRP was required. 5ml blood were collected in anticoagulant-free bottles which kept in slanted positions prior to being transported to the laboratory for serum separation. Clotted samples were centrifuged (4,000Xg for 15 min). Animal sera were stored at - 20 C° till assay application<sup>12</sup>

### Serological assays

#### Rose Bengal Test (RBT)

Rose Bengal Test was applied to serologically estimate the prevalence of brucellosis associated with Egyptian abattoirs. All serum samples were tested for agglutination against *Brucella* antigen using (PrioCHECK® *Brucella* Rose Bengal Test Kit, Prionics -Switzerland) by qualitative method according to the enclosed instructions. Briefly, equal volumes up to 30µl from both serum sample and *Brucella* antigen were mixed together and manually agitated for 2 minutes. Agglutination was considered a positive result and indicates the presence of specific antibodies to *Brucella* in the animal serum. While no agglutination was considered as a negative test result and indicates absence of specific antibodies to *Brucella*<sup>13</sup>. For increasing the test sensitivity, all positive sera were diluted by isotonic saline into 1/2, 1/4, 1/8, 1/16, 1/32, 1/64 dilutions and examined semi quantitatively<sup>14</sup>.

#### Modified in-house ELISA for bovine brucellosis diagnosis

Modified in-house ELISA assay was performed for rapid diagnosis of bovine brucellosis as described in<sup>15,16</sup>. 96 well ELISA plates (ICN Biomedicals, OH) were coated with antigens: *B. abortus* antigen (BD). Serum samples were diluted using the appropriated conjugating buffer to 1:160 and 1:320 dilutions. 100 µl from samples, Blank, positive and negative controls were introduced into the coated plates in two wells (duplicate), covered with a lid and incubated for one hour at 37°C in a humid chamber. Contents were discarded, washed three times and then loaded with the conjugate (Anti-Bovine IgG (H+L)°Peroxidase antibody produced in goat KPL). After that Plates were Incubated for 30 min at 37°C in a humid chamber. Then contents were discards and washed three times .100 µl of the prepared substrate (Sigma-Fast OPD tablets) solution were added to each well. Plates then were incubated for 30 min at room temperature (aprx 25°C-27°C) in the dark. 50 µl of stopping solution (1 M H<sub>2</sub>SO<sub>4</sub>) were added to each well. Developed color intensity ( optical density OD) were read at 492 nm. An OD cutoff point of 0.38 has been established as the cut-off of the assay. Any sample showed an OD value e" 0.38 was considered initially reactive at serum titer ≥ 1: 320 are considered reactive for the presence of

anti-*Brucella* antibodies. Samples showed OD  $\geq$  0.38 at 1:160 and  $<$  0.38 at 1:320 were considered equivocal. Sample show an OD  $<$  0.38 at 1:160 was considered non-reactive and was reported as negative for anti-*Brucella* antibodies.

#### Genomic DNA extraction

Total DNA was extracted from serum samples using a genomic DNA mini kit (Qiagen, QIAamp minprep, USA) according to the manufacturer's protocol. The extracted DNA was stored at  $-20^{\circ}\text{C}$  for further use. DNA concentration and purity of stock solutions were measured by with a Nano-Drop ND-1000 spectrophotometer at 260 and 280 nm.

#### Taqman real-time PCR assay

Serum samples were evaluated by Taqman real time assays for 11S7 gene amplification for identification of *Brucella* genus as described in<sup>17</sup>. The real-time PCR assays were carried out for *Brucella* identification using IS711 oligonucleotides pairs (forward: GCTTGAAGCTTGCGGACAGT) and (reverse GGCCTACCGCTGCGAAT and dual labeled FAM and BHQ probe (FAM-AAGCCAACACCCGGCCATTATGGT-BHQ-1) commercially purchased from ( Biosearch Technologies USA). Typical 25  $\mu\text{L}$  reaction contained: 12.5  $\mu\text{L}$  Taqman Universal PCR Master Mix (Applied Bio-systems, USA), 300 nM concentration of each forward and reverse primer, 200 nM concentration of the labeled probe and 2.5 ng of sample DNA. Taqman real-time PCR reactions were developed in (Step one plus - Applied Bio-systems equipment). The reaction mixture was initially incubated for 5 min at  $95^{\circ}\text{C}$ . Amplification was performed for 45 denaturation cycles at  $95^{\circ}\text{C}$  for 20 s, annealing and extension at  $60^{\circ}\text{C}$  for 1 min. Samples exhibited sigmoid amplification peaks with below 40 cycle thresholds (CTs) were considered as positive.

#### Statistical analysis

Statistical analysis was performed as described in<sup>18, 19</sup>. Considering the real time PCR the gold standard , the sensitivity and specificity of RBT and ELISA tests were evaluated using a total of 230 sera samples collected from two focal abattoirs . Each serum sample was subjected to the three tests and the results were entered into the computer. The reported data of the three tests' results was summarized in cross tabulation. Data were statistically analyzed using (Med-

Calc statistical software). sensitivity, specificity, positive likelihood ratio, negative likelihood ratio, prevalence, positive predictive value, negative predictive value and accuracy with relevance to 95% confidence intervals (CI) were calculated for serological tests in comparison to real time PCR as a gold test.

True positives = a Animals with real time positive and are test positive

True negatives = d Animals with real time negative and are test negative

False positives = b Animals with test positive and are real time negative

False negatives = c Animals with real time positive and are test negative

Identification of statistical variables ( Table 2,4)

## RESULTS

#### Prevalence of *Brucella* in male calves

A total of  $n=230$  serum sample were obtained from two main focal abattoirs in Egypt. All sera samples were tested blindly for the presences of *Brucella* using three assays ELISA , RBT and Taqman real time PCR. The prevalence of brucellosis varied according to the applied test (Table 1). The overall prevalence of brucellosis was  $n=124$  (53.9 %),  $n=173$  (75.2 %) and  $n=182$  (79.1 %) as determined by ELISA ,RBT and RT- PCR assays respectively.

#### Comparison of sensitivity and specificity of serological tests to IS711 Taqman real time PCR

In this study we compared the results of  $n=230$  serum sample obtained from serological tests to the results determined by Taqman real time PCR as a sensitive and gold standard test. Raw data including; positive , negative values and percentages for each test is indexed in (Table 2)

#### Comparison of rose Bengal test to IS711 Taqman real time PCR (Table 3 )

From ( $n=182$ ) positive RT-PCR up to ( $n=144$ ) were positive and ( $n=38$ ) sample were negative to RBT respectively. Using Med-Calc statistical software and by considering RT-PCR as the gold standard: the sensitivity , specificity and accuracy of RBT were calculated as 79.12% , 39.58 % and 70.87% respectively with a 95% confidence interval (CI) of (72.49% - 84.78%), (25.77% - 54.73%) and( 64.54% - 76.66%) respectively. Moreover, positive likelihood ratio (PLR) for

RBT test was 1.31 while negative likelihood ratio (NLR) was determined as 0.53. For rose Bengal Test, positive predictive value (PPV) 83.24% was reported as while the negative predictive value (NPV) was reported as 33.33 %.

#### Comparison of ELISA test to IS711 Taqman real time PCR (Table 5)

Regarding to ELISA assay only (n=101) sample from (n=182) positive RT-PCR were positive while (n=81) sample were negative. The

**Table 1.** Prevalence of brucellosis in asymptomatic male calves in the two slaughterhouse (abattoirs) by number and percentage of positive results per test used for detection

Sample( N)	ELISA	RBT	IS711 real time PCR
230	124 ( 53.9 %)	173 (75.2%)	182 (79.1)

**Table 2.** Raw data obtained from Rose Bengal Test (RBT) where true positives = (a) , true negatives= (d) , false positives= (b) and false negatives= (c)

Real time PCR	Brucella (RBT)		Total
	Positive	Negative	
Positive	(a) =144	(b) =38	(a+b) =182
Negative	(c) =29	(d) = 19	(c+d) =48
Total	(a+c) =173	(b+d) =57	n=230

test sensitivity was 55.49% with a 95% confidence interval (CI) of (47.96% - 62.85%) and the specificity was 52.08 % with a 95% confidence interval (CI) of (37.19% - 66.71%) in addition to an accuracy of 54.78% with a 95% confidence interval (CI) of (48.11% - 61.33%). The positive likelihood ratio for ELISA test was evaluated as 1.16 while negative likelihood ratio was determined as 0.85. The positive predictive value was reported as

**Table 3.** Statistical data of RBT for diagnosis of bovine brucellosis in comparison to real time PCR in a 95% confidence interval (CI)

Statistic parameter	Formula	Value	95% CI
Sensitivity	$\frac{a}{a+b}$	79.12%	72.49% to 84.78%
Specificity	$\frac{d}{c+d}$	39.58 %	25.77% to 54.73%
Positive Likelihood Ratio	$\frac{Sensitivity}{1 - Specificity}$	1.31	1.03 to 1.67
Negative Likelihood Ratio	$\frac{1 - Sensitivity}{Specificity}$	.053	0.34 to 0.83
Brucellosis prevalence	$\frac{a+b}{a+b+c+d}$	79.13%	73.30% to 84.19%
Positive Predictive Value	$\frac{a}{a+c}$	83.24%	79.60% to 86.33%
Negative Predictive Value	$\frac{d}{b+d}$	33.33 %	24.18% to 43.94%
Accuracy	$\frac{a+d}{a+b+c+d}$	70.87%	64.54% to 76.66%

81.45% while the negative predictive value was reported as 23.58 % %. Our results indicated that rose Bengal test is superior in sensitivity and specificity, followed by I-ELISA

## DISCUSSION

Screening and confirmatory diagnostic tests are the principal tool for efficient epidemiological analysis<sup>19</sup>. In Egypt although several papers were published to figure out the incidence of bovine brucellosis in various animal settings, only very few data are available on sensitivity and specificity of the serological tests.

Understanding the diagnostic sensitivity and specificity of an assay would contribute to minimize diagnostic problems in classifying diseased and non-diseased animals accurately in order to prevent unnecessary economical deficits

in cases the animals are incorrectly identified by the tests<sup>20</sup>. In accordance with our results rose Bengal test had the superior performance and then ELISA in descending order of sensitivity, specificity and accuracy. RBT continues to be the preferred screening tool owing to its fast result and cost availability<sup>21</sup>. A test with suitable positive predictivity (PPV) is necessary for ensuring the presence of disease, in addition to a test with good negative predictivity (NPV) to exclude the infection<sup>22</sup>. The purpose of diagnosis is usually to search for cases while maintaining false positives to a minimal level. One of the most important drawbacks of rose Bengal test is the considerable high numbers of false positive results which are commonly explained as a result of cross reactivity of the test with different types of bacteria. Considering the real time PCR the gold standard, the present study indicated that

**Table 5.** Statistical data of ELISA for diagnosis of bovine brucellosis in comparison to real time PCR in a 95% confidence interval (CI)

Statistic parameter	Formula	Value	95% CI
Sensitivity	$\frac{a}{a+b}$	55.49%	47.96% to 62.85%
Specificity	$\frac{d}{c+d}$	52.08 %	37.19% to 66.71%
Positive Likelihood Ratio	$\frac{\text{Sensitivity}}{1 - \text{Specificity}}$	1.16	0.84 to 1.60
Negative Likelihood Ratio	$\frac{1 - \text{Sensitivity}}{\text{Specificity}}$	0.85	0.62 to 1.17
Brucellosis prevalence	$\frac{a+b}{a+b+c+d}$	79.13%	73.30% to 84.19%
Positive Predictive Value	$\frac{a}{a+c}$	81.45%	76.08% to 85.84%
Negative Predictive Value	$\frac{d}{b+d}$	23.58 %	18.37% to 29.75%
Accuracy	$\frac{a+d}{a+b+c+d}$	54.78%	48.11% to 61.33%

the sensitivity of rose Bengal test is 79 %, while Cernyseva *et al*<sup>22</sup> found that the sensitivity of the test was 68.6%, Mesa *et al*<sup>23</sup> and Altwegg *et al*<sup>24</sup> place the quantity at 93.8 and 100%, respectively. The sensitivity of the RBT relies on the antigenic concentration and commercial antigenic formula utilized which provide a clarification for the differences of viewpoint present in the literature around the sensitivity of the RBT.

The sensitivity and specificity of ELISA in the present study (55.49% and 52.08 % respectively) are deviate from those experienced in other reports, for instance<sup>25</sup>. stated an ELISA sensitivity of 71.3% and a specificity of 100%. The lower sensitivity of ELISA in our study could be also due to the participation of animal with former or possibly chronic infection where the titer of antibodies is relatively low. As determined by the results acquired from the current study, the employment of real-time PCR approach along with one or more serological assay, particularly RBT is highly advisable for appropriate diagnosis of bovine brucellosis in developed and low resources regions.

## CONCLUSION

There is no individual serological test is convenient in all epidemiological conditions and all animal species. all tests have constraints especially when examining individual animals. Attention is required to be given to all variables that affect the relevance of the test procedure and test results to a certain diagnostic interpretation or utility. Therefore, carrying out test validation is extremely necessary to know the test properties and to decide the type of test we need to use for the study objective, epidemiological monitoring, or global trade. However, the assessment for the diagnostic test for diverse purposes not only depends on the accuracy, but also should take into account the capacity for the test throughput, technical intricacy, as well as affordability.

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