Physiological and Biochemical Response of Winter Wheat (*Triticum aestivum* L.) to Ambient O$_3$ and the Antiozonant Chemical Ethylenediurea (EDU) in Jeddah, Saudi Arabia

**LAILA A. BAQASI**$^{1,2}$, **HUDA A. QARI**$^{1,2}$ and **IBRAHIM A. HASSAN**$^{2,3}$*

$^1$Department of Biology, Faculty of Science, King Abdulaziz University, Jeddah, Saudi Arabia.
$^2$Centre of Excellence in Environmental Studies (CEES), King Abdulaziz University, 21589 Jeddah, Saudi Arabia.
$^3$Department of Botany, Faculty of Science, Alexandria University, 21589 El Shatby, Alexandria, Egypt.
*Corresponding author E-mail: ihassan_eg@yahoo.com

http://dx.doi.org/10.13005/bpj/1346

(Received: February 07, 2018; accepted: March 09, 2018)

**ABSTRACT**

This study was conducted to investigate the use of ethylenediurea (EDU) as a possible tool to evaluate O$_3$ effects on wheat (*Triticum aestivum* L.) plants under field conditions in Jeddah. Wheat plants were exposed to ambient O$_3$ (AA) and the antiozonant chemical ethylenediurea (EDU) in closed fumigation chambers for the full growing season. Growth, yield and physiology were determined in response to O$_3$ and/or EDU. EDU-treated plants had higher photosynthetic rates (24%) and stomatal conductance (25%), which were reflected in higher growth and yield in terms of number of grains. The present study revealed that EDU could be used as a promising tool to mitigate damaging effects of O$_3$ on under field conditions. EDU protected wheat plants leading to increases in photosynthetic rates, growth and yield.

**Keywords:** Ethylenediurea (EDU), Growth, Ozone, Protection and Yield.

**INTRODUCTION**

Tropospheric O$_3$ is produced through photochemical reactions of its precursors (NOx and VOCs) and its concentrations varied spatially and temporally. Ozone (O$_3$) is a secondary phytotoxic pollutant causing changes in metabolic, biochemical and physiological processes leading to reductions in growth and yield of economic crops$^{1-4}$.

Ozone concentrations in Jeddah were recorded to be between 40 – 70 nl l$^{-1}$ $^{4,5}$. These concentrations are high enough to affect many plant processes, such as photosynthesis, transpiration, nutrient uptake, and senescence, resulting in significant effects on crop growth and yield.

Jeddah suffers from serious air pollution problems due to emissions from different sources. There is a very poor legislation regarding emissions from old cars in streets as well from factories which cause environmental hazards. Levels of heavy metals, gaseous air pollutants and particulate emissions far exceed internationally acceptable standards$^{4,6}$. 
The antiozonant chemical ethylenediurea (EDU) is extensively used to assess the crop loss due to O$_3$.

Wheat is a nutritious and versatile crop, it has been known to be sensitive to O$_3$ in the USA, Europe and other areas of the world. However, very little is known about its sensitivity in the Middle East. As yet, in Saudi Arabia, the information on impact of O$_3$ on plants are extremely scanty and fragmented.

The objective of the present investigation was to assess the response of local variety of wheat (Triticum aestivum L.) to ambient O$_3$ in terms of growth, photosynthesis, stomatal conductance, antioxidants and photosynthetic pigments and to assess the use of the antiozonant chemical ethylenediurea abbreviated as EDU as a reliable, effective and cheap tool to protect of O$_3$ effects on wheat plants under filed conditions in Jeddah, Saudi Arabia.

MATERIALS AND METHODS

Plant samples, growth conditions and Experimental design

Grains of wheat (Triticum aestivum L.) were hand-planted in 50 cm$^2$ plastic pots (10 seeds in each pot) filled with multipurpose in a controlled glasshouse. Plants were transferred to the field when second foliage leaf appeared on 3 Feb 2015 were sown. Experiments were conducted in the open field during Jan 2015 – April 2015. The treatments were: (a) ambient air (AA) and (b) AA + EDU (EDU). There were 12 pots/chamber. The crop matures in 70 - 80 days.

EDU application

EDU was applied as a soil drench; 200 ml of a 150 ppm EDU every 14 days. The control plants received tap water. There were 16 plastic pots (18 cm height x 16 cm diameter); 8 for AA and 8 for EDU.

Biomass and grain yield

Five plants per pot were harvested for determination of yield parameters at the final harvest during April 2015.

Gas exchange measurements and chlorophyll content

Net CO$_2$ photosynthetic rate (A) and stomatal conductance ($g_s$) were measured using a portable LICOR (IRGA-LICOR-6400, Lincoln, NE, USA). Measurements were carried out on ten attached leaves per treatment on weekly basis.

Biochemical assays and antioxidant enzymes assays

Leaves were weighed and ground at about 4°C in 50 m Tris-HCl buffer containing 3 mM MgCl$_2$, then the homogenates were centrifuged at 20 000 for 15 min (Centrifuge17 S/RS, Heraeus Sepatech). Enzyme assays were performed using the supernatants and the results were expressed on protein basis.

Four antioxidant enzymes were determined in the present investigation; namely; Superoxide dismutase (SOD), Catalase, Guaiacol peroxidase

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>AA</th>
<th>EDU</th>
</tr>
</thead>
<tbody>
<tr>
<td>SOD (mg$^{-1}$ protein)</td>
<td>65.6 ± 7.8$^b$</td>
<td>50.1 ± 6.3$^a$</td>
</tr>
<tr>
<td>APX (µmol min$^{-1}$ mg$^{-1}$ protein)</td>
<td>578.7 ± 76.8$^a$</td>
<td>498.1 ± 51.1$^b$</td>
</tr>
<tr>
<td>GPX (µmol min$^{-1}$ mg$^{-1}$ protein)</td>
<td>8.02 ± 1.01$^b$</td>
<td>7.3 ± 1.9$^a$</td>
</tr>
<tr>
<td>CAT (µmol min$^{-1}$ mg$^{-1}$ protein)</td>
<td>781.7 ± 97.5$^a$</td>
<td>614.1 ± 72.7$^b$</td>
</tr>
</tbody>
</table>

Each figure is a replicate of 8 reading ± 1SE. Mean not followed by the same letter are significantly different of P < 0.05.
(GPX), Ascorbate (APX) following the methods as given by Lee et al.\textsuperscript{18}.

**Data analysis**

ANOVA test was applied to data, differences between means were tested by Tuckey Test using SPSS statistical package, USA.

**RESULTS AND DISCUSSION**

The average temperatures were 33.2 °C and the average 8-h O\textsubscript{3} concentrations were 45.9 ppb. Accumulated ozone exposure above a threshold of 40 ppb (AOT40 was calculated to be 3.304 ppm).

EDU-treated plant had higher photosynthesis rate (A) and stomatal conductance ($g_s$) by 24%, increased by 25%, respectively, throughout the entire course of experiment (Fig. 1 A & B).

The rate of increase in photosynthetic rates due to application with EDU was steady for the first...
three weeks and increased thereafter and by the 8th week, it showed a slight decline due to senescence of the crop (Fig. 1A). On the other hand, the decline in stomatal conductance was more pronounced in the 8th week in both EDU-treated and ambient air plants (Fig. 1B).

Plant treated with EDU showed a positive impact on number and weight of grains. EDU caused increases in the grain number by 50% and grain weight by 82% (Fig. 2 A & B).

EDU did not alter shoot growth (P < 0.05), while it caused overall increase in root dry weight by 23% when compared to plants grown in ambient (AA) (Fig. 3).

Exposure to ambient O₃ caused increases in Superoxide Dismutase (SOD), Ascorbate

---

**Fig. 2**: Effect of EDU on wheat plants (*Triticum aestivum* L.) grain (A) number and (B) weight. Each figure is a mean of 109 readings ± 1SE

**Fig. 3**: Effect of EDU on shoot and root growth wheat plants (*Triticum aestivum* L.). (Means ± 1SE)
Peroxidase (APX) and Catalase (CAT) by 31, 16 and 27%, respectively (Table 1). Nevertheless, Guaiacol Peroxidase (GPX) showed insignificant increase 9.8%, (P > 0.05).

Jeddah is the second largest city in the Kingdom of Saudi Arabia, suffering from industrial and vehicular emissions which cause increases in tropospheric O$_3$. It is growing economically and industrially and having many anthropogenic activities; its population is about 3 million inhabitant.

The concentrations of O$_3$ recorded in Jeddah is between 40 – 50 ppb were close to those previously observed in several arid areas of Saudi Arabia. Until now, one investigation on impact of ambient O$_3$ and on crops was carried out in Saudi Arabia and the extent of economic losses of O$_3$ to crops in this country has been poorly evaluated.

The present investigation showed clearly that O$_3$ concentrations of Jeddah city cause negative effects on growth, biomass accumulation, and yield. Analyses of metabolic processes were performed in order to perceive O$_3$ toxicity at early stages.

Ambient O$_3$ significantly affected photosynthetic rates (A) values, in comparison to EDU-treated plants. This result is in agreement with our previous work. Moreover, EDU-treated plants had higher stomatal conductance ($g_s$) when compared to plants grown in ambient air.

There is a significant correlation between A and $g_s$ (data not shown), which indicates that reduction in stomatal conductance due to closing of stomatal could be the cause of the apparent reduction of plants exposed to ambient air "AA".

EDU has been extensively used to assess the impact of ambient O$_3$ in dry tropical/subtropical regions. However, there is only one study so far in the literature describing EDU application to crops in Saudi Arabia. EDU has no toxic effect per se. The positive effects of EDU were relevant on all variables of wheat plants in the present study, including O$_3$- caused visible injury, growth, yield, and physiological parameters.

EDU caused increases in the biometric parameters (growth parameters, grain number and weight) compared to plants exposed to ambient conditions, proving its effectiveness in alleviating toxic effects of O$_3$.

Plants can avoid and tolerate toxic effects of O$_3$ through induction of protective scavenging systems. Plants produce antioxidant enzymes (e.g. catalase "CAT", peroxidase "GPX", superoxide dismutase "SOD" and others) and ascorbate peroxidase "APX"; the enzymes of the ascorbate-glutathione cycle (Halliwell-Asada cycle) in order to alleviate accumulation of harmful concentrations of reactive oxygen species (ROS) caused by O$_3$.

Oxidative stress causes increases in activities of these enzymes, which indicate production of H$_2$O$_2$. SOD plays a key protective role against O$_3$ phytotoxicity, as it scavenges O$_2^-$ and catalyzes its dismutation to H$_2$O$_2$ and O$_2$. Both CAT and APX are scavengers of H$_2$O$_2$.

Results of the present study point out that decreased activities of antioxidant enzymes in EDU-treated wheat plants most likely increased resistance of plants to O$_3$. This protective mechanism of against harmful oxidative stress is reported in wheat plants in Saudi Arabia, in terms of levels of reactive oxygen species “ROS” and their analogous scavenging enzymes.

In conclusion, EDU conferred a significant protection to wheat plants leading to improvement in growth, yield, and photosynthetic performance.

ACKNOWLEDGEMENTS

This work was supported with a Grant from King Abdul Aziz City for Science & Technology (KACST). Grant #1498-37-TA.
REFERENCES


11. Lee EH, Upadhyaya A, Agrawal M, Rowland R.A. Mechanism of EDU-induced O3 protection: re-examination of free radical scavenger system in snap bean exposed to...


