

The Effect of Some Fruit Juices on Glimpiride Pharmacokinetic in Rat Plasma by using High Performance Liquid Chromatography-Mass Spectrometry

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ABSTRACT

Food-drug interactions may affect pharmacokinetics or pharmacodynamics of a drug. Failure to identify and properly manage drug-nutrient interactions may lead to serious consequences. In this study, we try to develop and validate a simple and sensitive analytical method for measuring of glimepiride in rat plasma and examine of any possible interactions of glimepiride with different fruit juices. A mixture of 80 % methanol and 20 % of 0.1% FA in water was used as a mobile phase and ACE 5 C18 (50 X 2.1 mm, 5 μ m) column were employed; and clarithromycin was used as internal standard. This method was confirmed in terms of linearity, precision, accuracy, stability and system suitability. The validated method was used to examine the effects of grapefruit and licorice of glimepiride pharmacokinetic in rat plasma. The kinetic data shows that glimepiride plasma level was high when combined with licorice, and at the same level when combined with grapefruit. The administration of grapefruit juice concomitantly with glimepiride shows insignificant effect on its C_{max} , elimination half-life and clearance ($P>0.05$). While for licorice juice, it has no significant effects on glimepiride pharmacokinetic parameters. A simple and sensitive analytical method for determination of glimepiride in rat plasma has been developed and validated by using HPLC-MS. Also, we found that fruit juices can alter glimepirides pharmacokinetic if they were taken simultaneously.

Keywords: HPLC-MS, glimepiride, pharmacokinetics, grapefruit, licorice.

INTRODUCTION

Glimepiride is an active, well-tolerated oral hypoglycemic drug¹. It is used chronically to maintain glucose level within normal range^{2,3}. It is different

from other sulfonylureas in which it will result in less hypoglycemic effect and weight loss⁴. It might be combined with other oral antidiabetic drugs or insulin, to improve the level of glucose in blood⁵.



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Glimepiride is metabolized by CYP2C9 (CYP450 isoenzyme)⁶. For that it should be considered when use it with any CYP 2C9 inducers (e.g. rifampicin) or inhibitors (e.g. fluconazole)⁷. Alcohol consumption may decrease the hypoglycemic effect of glimepiride in an unpredictable way⁸.

Drug-Food interactions may cause disturbances in the pharmacokinetics or pharmacodynamics of a drug or nutritional component or alteration in nutritional status because of using a drug⁹. The improperly management of drug-food interactions may cause a serious complication like reduction in certain oral drug absorption which leads to failure treatment¹⁰. In addition, nutrients may induce or inhibit certain enzymes in the gut that will alter the bioavailability of certain. For example, grapefruit juice is a selective intestinal CYP3A4 inhibitor¹¹. It was found that grapefruit may increases the bioavailability of certain drugs e.g. felodipine threefold than normal that may raise the risk of side effects¹². Also, long term using of drugs may lead to disturbances in GIT functions result in loss of body fluids and electrolytes. Due to that it is recommended to limit the drug prescription for necessary medications and use it for short time to lessen the adverse of drug-food interactions¹³.

Moreover, two clinical manifestations may result due to the interaction between drugs and nutrients, these are: first: elevation of drug levels in plasma which elevate the risk of side effects and

toxicity, second: reduction of drug plasma levels which cause failure of treatment^{14,15,16,17,18,19}.

After oral administration, glimepiride is almost completely absorbed from the GI tract and the C_{max} is reached within 2-3 hours. Taking glimepiride with meals, slightly increases the mean T_{max} and slightly decreases the mean C_{max} and AUC²⁰. It was found that after intravenous dosing in normal subjects, the volume of distribution was 113 ml/kg, the total body clearance (CL) was 47.8 ml/min and the protein binding was greater than 99.5%²¹.

Glimepiride is totally metabolized by the oxidative biotransformation after either IV or oral administration. The main metabolites are cyclohexyl hydroxyl methyl derivatives and carboxyl derivatives. CYP2C9 found to be included in the biotransformation of glimepiride to cyclohexyl hydroxyl methyl derivatives, which is extra metabolized to carboxyl derivatives by enzymes in cytoplasm. The cyclohexyl hydroxyl methyl derivatives possesses about one third of the pharmacological activity in comparison to its parent in an animal model. However, its hypoglycemic effect is not clear in humans²². These metabolites are excreted in feces and urine^{23,24}. The present work is focusing on development of a sensitive and simple chromatographic method for quantifying glimepiride in rat plasma, to be followed by examination of any possible interactions of glimepiride with different fruit juices (grapefruit and licorice) on animals experimental.

Table 1: Chromatographic conditions

HPLC conditions	Pump flow rate 0.6 ml/min	Autosampler injection volume 5µl	Autosampler temp. 10°C	Column oven temp. 40°C			
Chromatography	Mobile phase	Mixture of 80% methanol and 20% of 0.1% FA in water					
	Column type	ACE5C18 Column (50x2.1 mm,5µl)					
	Expected Retention times (minutes)	Glimepiride 0.3 - 0.5	Clarithromycin (IS) 0.25 - 0.4				
MRM detection conditions	Analytes	Q1 Mass	Q3Mass	DP	EP	CE	CXP
	Glimepiride	491.485	352.1	91	10	19	22
	Clarithromycin (IS)	748.200	158.1	76	10	30	4
MS conditions	CUR	Gas 1	Gas2	IS voltage	CAD	T	
	25	75	75	5500	5	550	

MATERIALS AND METHODS

Reagents

Rats plasma, (Harvested from animals from animal house in UOP). Methanol advanced gradient grade (Fisher scientific, B# 1155904), acetonitrile advanced gradient grade (Fisher scientific, B# 1156250), formic acid advanced gradient grade (GPR Rectapur, B# 07L210512), sodium hydroxide powder (VWR international), glimepiride raw material (JPM Co. B# WS/07/272), and clarithromycin raw material (JPM Co. B#0040310095)

Instrumentation

The used was an API Mass spectrometer which consists of: Degasser (Agilent 1260), solvent delivery systems pump (Agilent 1200), auto-sampler (Agilent 1200), thermostat column compartment (Agilent 1200), API 4000 mass spectrometer),

ACE 5, C18 (50 x 2.1 mm), 5 μ m, computer system, windows XP, SP3, data management software 1.5.1).

Bath sonicator Crest model-175T (Ultra Sonics CORP.), Sartorius balance BP 2215, Sartorius pH meter (Professional meter PP-25), and Centrifuge (eppendorf 5417C)

Chromatographic conditions

Mobile phase is composing of 80% of methanol and 20% of 0.1% formic acid in water. Column type ACE5C18 Column (50x2.1 mm, 5 μ l), clarithromycin was used as internal standard. Further details and mass parameters are illustrated in Table 1.

Stock and working solutions

Clarithromycin (IS) Solutions

A stock solution of clarithromycin (IS) at a level of 1 mg/ml was prepared by taking 10mg of

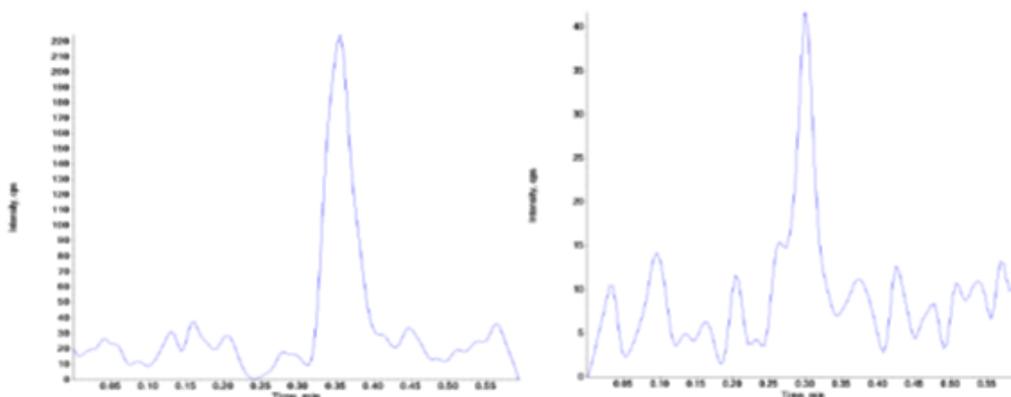


Fig. 1 : Glimepiride blank chromatogram

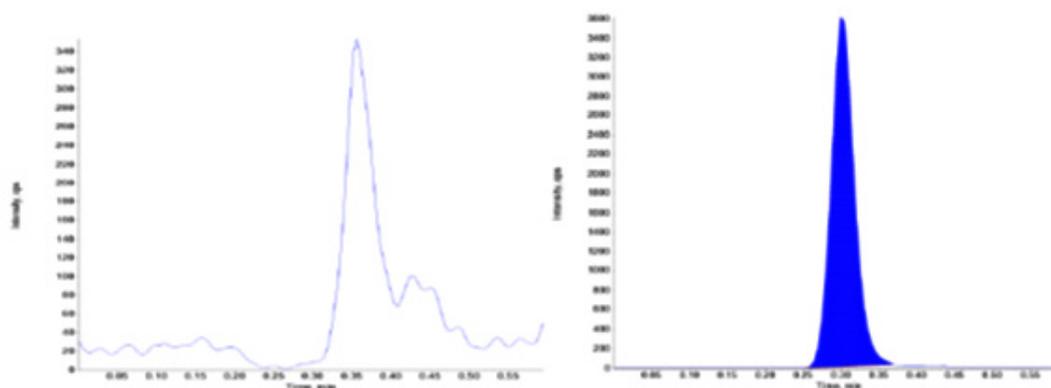


Fig. 2 : Glimepiride zero chromatogram

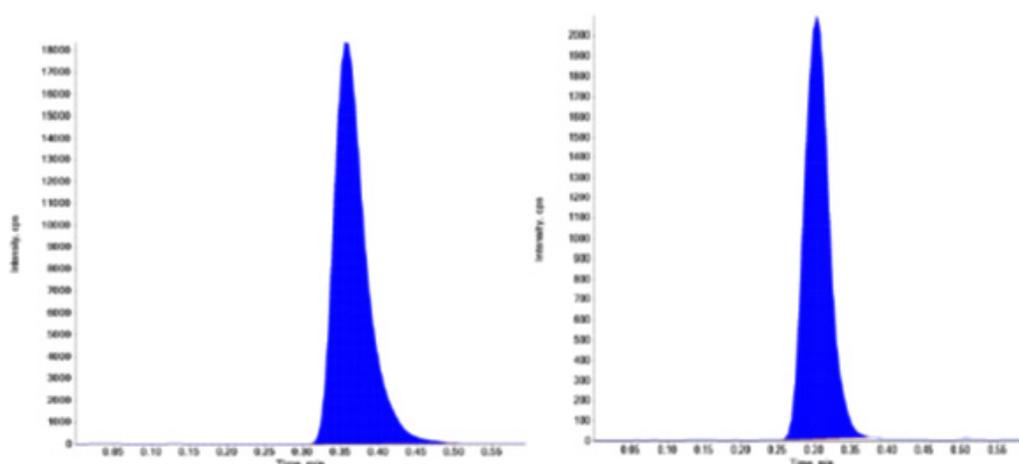


Fig. 3: Glimepiride rat unknown chromatogram at 1 hour

clarithromycin and dissolved it in 10ml methanol. Working solutions of IS were prepared by diluting 50.0 μ l from previous stock solution into 10.0 ml of methanol, to get a concentration of 5 μ g/ml, then further dilution was done by diluting 100 μ l (from standard solution of 5 μ g/ml) into 250.0 ml of methanol to get a final concentration of 2 ng/ml.

Glimepiride solutions

To prepare the stock solution of glimepiride 10.0 mg was taken and dissolved in 0.5 ml of DMSO; then the volume was completed to 10.0 ml by methanol to get a concentration of 1.0 mg/ml. Then, 200.0 μ l of the prepared solution was diluted into 10.0 ml of 1:1 water/methanol to obtain a concentration of 20.0 μ g/ml working solution.

Calibration curve and quality control (QC) samples in plasma

Calibration curve of seven standards of glimepiride were prepared by taking 100 μ l from each working solution and spiked into 10 ml of plasma to get concentrations of: 20, 50, 100, 200, 300, 400 and 600 ng/ml. Quality controls (QC) samples were: 60, 250 and 500ng/ml. Each plasma sample was divided to 25 μ l in 1.5 ml eppendorf tubes and stored at -30°C . Standard and quality control samples were used daily during analysis.

Method Validation

Precision and accuracy

Intra-day precision and accuracy

parameters were daily evaluated by analyzing of 6 duplicates of the lower limit of quantification (LLOQ) and QC samples. Three runs of LLOQ and QC were analyzed in three days to evaluate the inter-day variability. The ratios between standard deviation (SD) and the mean were used to find the relative standard deviation (RSD) values or CV%. Results were in percentages.

Accuracy was evaluated by calculating the ratio between the amounts recovered from the control samples with its values existing in the samples (theoretical values). The suitable values of intra-day and inter-day accuracy and precision were under 15%. For the LLOQ, the suitable value must be lower than 20% according to EMEA.

Linearity

Calibration curve of glimepiride was plotted between the ratio of drug peak area (PAR) and area of internal standard against drug concentration (C). The resulted equation: $\text{PAR} = \text{Slope} \times C + \text{Intercept}$ used to calculate the unknown concentrations of glimepiride. The nominal values of the drug and the calculated PAR used to determine the slope and intercept. Correlation coefficient (R^2) values used to determine the linearity of the plotted curve.

Specificity and selectivity

Six batches of plasma were used to determine the specificity of the method. Each batch was analyzed as blank and zero samples,

and compared to LLOQ to confirm the lack of endogenous peaks.

According to the EMEA 2011, six different batches of suitable blank medium that are independently evaluated must be used to verify the selectivity. Normally, the lack of interfering constituents is accepted where the response is lower than 20% of the LLOQ for the analyte and 5% for IS.

Sensitivity

In sensitivity, the chromatographic response of LLOQ must be e^5 times than the blank response with accuracy 80-120% and precision d^2 20%. Six replicates of LLOQ plasma samples were prepared along with the calibration curve.

Stability

Stability of glimepiride in rats' plasma is determined using low and high QC samples (blank plasma spiked with glimepiride at a level of a maximum of 3 times the LLOQ and to the ULOQ)

which are analyzed directly upon preparation and again after 24 hours at room temperature. The QC samples were evaluated by the calibration curve, which prepared from newly spiked calibration samples, and the resulted concentrations are compared to the nominal concentrations. The accepted levels of mean concentrations are $\pm 15\%$ of the nominal concentration.

Preclinical study

Preparation of glimepiride solution and fruit juices

Glimepiride solution to be given to rats in a dose of 115mg/kg (based on maximum daily dose of human) was prepared by weighing 0.015 g of glimepiride raw material then dissolved in 1.0 liter of 0.1 M sodium hydroxide solution.

Grapefruit; *Citrus paradise* (Rutaceae) was purchased from local market in Jordan, divided in half and directly squeezed by hand before used without any additions or treatments.

Table 2: Results of inter and intra- day precision and accuracy

Day 1 Target conc.	LLOQ 20 ng/ml	QC _{Low} 60 ng/ml	QC _{Mid} 250 ng/ml	QC _{High} 500 ng/ml
Calculated conc. \pm SD	19.78 \pm 0.65ng/ml	54.49 \pm 1.84ng/ml	266.94 \pm 9.12ng/ml	515.89 \pm 26.54
SE	0.27	0.75	3.73	10.84
Accuracy \pm SD	98.88 \pm 3.24	90.82 \pm 3.87	106.77 \pm 3.65	103.18 \pm 5.31
CV%	3.28	3.38	3.42	5.15
Range	19.4-20.4	51.9-56.6	250.4-275.1	487.5-548.6
Day 2				
Target conc.	20 ng/ml	60 ng/ml	250 ng/ml	500 ng/ml
Calculated conc. \pm SD	21.46 \pm 0.43	54.33 \pm 1.54	256.19 \pm 14.87	461.92 \pm 10.08
SE	0.18	0.63	6.07	4.12
Accuracy \pm SD	107.29 \pm 2.15	90.54 \pm 2.57	102.48 \pm 5.95	92.38 \pm 2.02
CV%	1.99	2.84	5.80	2.18
Range	21-21.9	52.1-56.8	236.3-274.95	452.7-478
Day 3				
Target conc.	20 ng/ml	60 ng/ml	250 ng/ml	500 ng/ml
Calculated conc. \pm SD	21.59 \pm 0.52	61.07 \pm 1.09	255.99 \pm 6.01	485.19 \pm 18.31
SE	0.21	0.44	2.45	7.48
Accuracy \pm SD	107.90 \pm 2.61	101.78 \pm 1.81	102.39 \pm 2.41	97.04 \pm 3.66
CV%	2.42	1.78	2.35	3.77
Range	20.8-22.1	59.6-62.8	247.8-262.9	461.2-504.1

Licorice roots from *Glycyrrhiza glabra* (Fabaceae) cultivated in Jordan, which were used to prepare the licorice juice. The roots were soaked in distilled water for 1 hour. The wet licorice paste then was taken in a clean white tissue and hold on a holder. Finally, the cold water was allowed to drizzle over the tied licorice cushion in a period of 2 hours. The prepared licorice juice has no additives and it was stored at 4°C and used within 48 hours.

Animal handling and study protocol

Study protocol was approved by ethical committee of the High Research Council, faculty of pharmacy and medical science, University of Petra, Amman, Jordan (No.17/2014). Adult male

and female Sprague Dawley laboratory rats were provided by the animal house of University of Petra with an average weight of 270.0 ±30g.

Rats were sited in controlled environment (20-25 °C) and uncovered to a photoperiod cycle (12 hours light/12hours' dark) daily. All rats fasted 24 hours before the experiment day.

Rats were divided into 14 groups, each of six rats. Four groups received glimepiride only and another five groups received glimepiride with grapefruit, while the last five groups received glimepiride with licorice. Then each rat received certain amount of a drug solution (per weights) orally

Table 3: Linearity and linear working range of six calibration curves of glimepiride data based on the measured concentration

Calibration Curve #	Measured concentration for each standard point (ng/ml)						
	20	50	100	200	300	400	600
1	19.07	53.33	98.58	208.85	289.87	379.89	620.42
2	19.30	51.62	103.92	205.13	284.30	376.46	629.26
3	19.84	48.92	105.38	209.18	284.09	374.34	628.25
4	18.04	50.40	109.52	210.93	292.94	380.27	607.90
5	18.42	50.62	108.46	204.27	297.34	381.46	609.43
6	18.03	52.17	103.50	211.73	297.85	388.80	597.93
Mean ± SD	18.78 ±0.74	51.18 ±1.54	104.89 ±3.93	208.35 ±3.03	291.07 ±6.08	380.20 ±4.98	615.53 ±12.49
CV%	3.93	3.01	3.75	1.46	2.09	1.31	2.03
Accuracy %	93.92	102.35	104.89	104.17	97.02	95.05	102.59

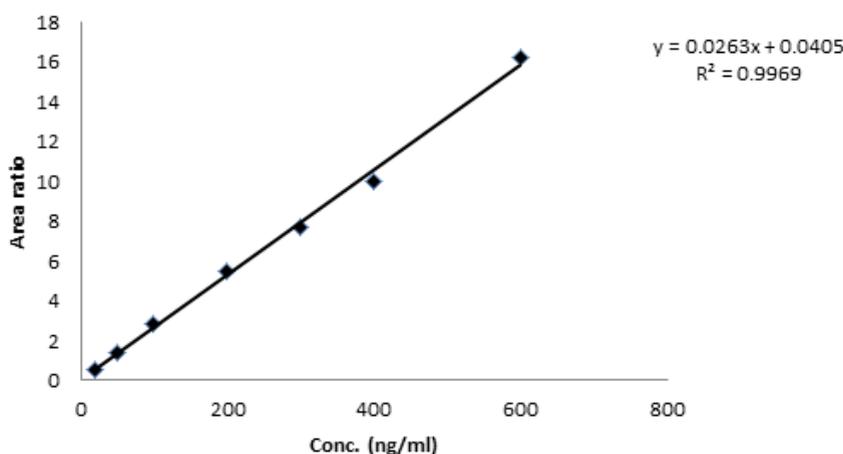


Fig. 4 : Plot of linearity for mean six calibration curves

by an oral gavage. Two milliliters of each fruit juices were given to the rats prior the administration of the drug, in addition by oral gavage.

Sample collection and processing

Blood collected by cutting the rats tail where few drops were taken and put in the Eppendorf tubes. Samples were pooled from each subgroup at the following time points: zero, 0.5, 1.0, 2.0, 3.0, 4.0, and 6.0 hours. Blood samples were collected in a micro-tubes containing Ethylenediaminetetraacetic acid (EDTA) and then after centrifugation at 5,000 RPM for 10 minutes; plasma was taken in labeled eppendorf tubes and stored at -30°C till the analysis.

For extraction 25 μ l of plasma sample and 200 μ l of internal standard (2 ng/ml clarithromycin in methanol) were taken in a 1.50 ml eppendorf tube and mixed for 1.0 min; then centrifuged for 15 min at 14000 rpm. The supernatant was then transferred into auto-sampler tubes; then analyzed under the chromatographic conditions as shown in Table 1. This

procedure was used for rat samples, calibration and quality control samples.

Pharmacokinetic study

Plasma level -time profile of glimepiride, glimepiride with liquorice, and glimepiride with grapefruit was constructed by plotting average plasma concentration in ng/ml versus time in hours.

Non-compartmental analysis was performed to calculate C_{max} , T_{max} , AUC_{-6} , MRT_{-6} , Clearance (Cl/F), elimination rate constant (K_{el}) and terminal elimination half-life ($t_{1/2}$) for glimepiride alone and another time when it was given with fruit juices using Microsoft Excel 2010.

Statistical analysis

SPSS version 21 computer software was used for statistical analysis. Paired t-test was used to detect significance in pharmacokinetic parameters using 95% confidence interval.

Table 4: Results of glimepiride QC low samples Autosampler stability at 10°C

Time	AUC Drug	AUC IS	Ratios	QC low (60mg/ml)		Accuracy %	Stability %
				Measured Conc.	Mean Measured		
0.0 hr	12281	7422	1.65	58.84	58.95	98.08	100.00
	11965	7245	1.65	58.72		97.88	
	11596	6960	1.66	59.27		98.80	
24.0 hr	11922	7278	1.63	59.31	58.66	98.85	99.52
	11000	6877	1.60	57.77		96.30	
	11466	7042	1.62	58.91		98.19	

Table 5: Results of glimepiride QC high samples Autosampler stability at 10°C

Time	AUC Drug	AUC IS	Ratios	QC low (500mg/ml)		Accuracy %	Stability %
				Measured Conc.	Mean Measured		
0.0 hr	73234	5466	13.39	497.87	497.64	99.57	100.00
	73043	5493	13.29	494.16		98.83	
	74519	5528	13.48	500.91		100.18	
24.0 hr	86069	6996	12.30	482.89	488.23	96.58	98.11
	86981	6945	12.52	491.69		98.34	
	87106	6978	12.48	490.11		98.02	

RESULTS AND DISCUSSION

Validation of HPLC-MS method

Validation of the present analytical method of glimepiride was conducted to evaluate the reliability of the method for the measuring the drug concentration in rat plasma. Figures 1-3 show the chromatogram of glimepiride blank, zero and rat plasma sample after 1 hour.

Accuracy and Precision

Results of inter-day accuracy over the tested concentration range (90 and 108%) in each

day are shown in table 2. In comparison to the accepted standards (85-115 %) for all concentrations and for LLOQ (80-120 %), the resulted accuracy was within the accepted ranges. Further, results of intra-day accuracy lied in the same accepted rang.

Inter and Intra-day precision (CV%) was less than 20% for LLOQ and 15% for the other concentrations. The CV% evaluation over the three days was less than 5.80% for (QC_{mid}), which indicates the closeness of the measurements (Table 2).

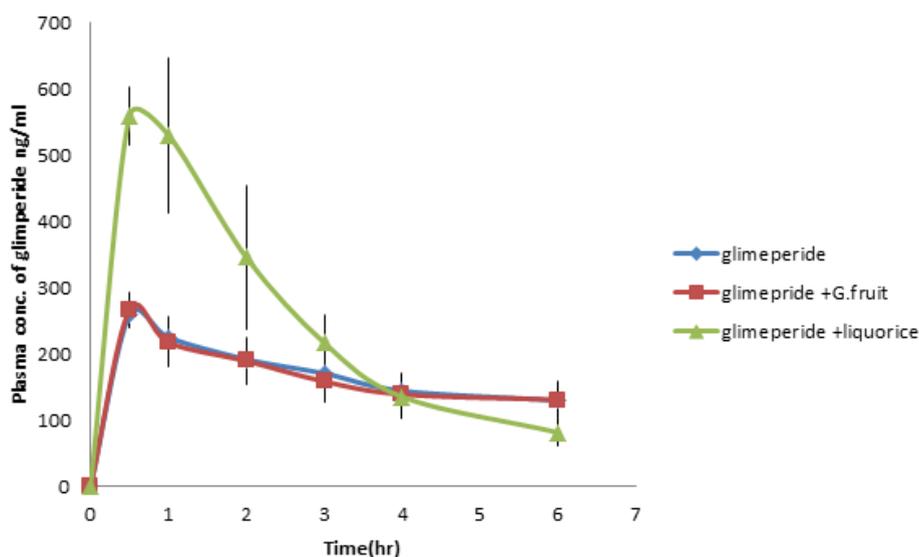


Fig. 5 : Plasma level-time profile of glimepiride when given alone and with liquorice and grape fruit

Table 6: Pharmacokinetic parameters of glimepiride alone and with liquorice and grapefruit juices

Parameters	Glimepiride alone	Glimepiride-grape fruit	Glimepiride- liquorice
C _{max} (ng/ml)	258.7±7.9	266.2±46.5	584.5±85.6*
T _{max} (hr)	0.500	0.500	0.500
AUC ₀₋₆ (ng.hr/L)	1008.58±22.65	984.9±30.43	1526.12±35.33*
AUMC ₀₋₆ (ng.hr ² /L)	1998.7±35.8	1921.98±4	2694.58±42.8*
MRT ₀₋₆ (hr)	1.98±0.4	1.95±0.5	1.76±0.5*
K _{el} (hr ⁻¹)	0.500±0.01	0.510±0.02	0.252±0.13*
Terminal T _{1/2} (hr)	1.38±0.3	1.35±0.29	2.75±0.3*
Cl/F (ml/hr)	29.7±8.4	30.45±10.2	19.6±6.3*

*Significant (p<0.05)

Linearity

Linear regression equation was applied to calculate the drug concentration at each validation day, using one unique target concentration for getting the "D area/ IS area" at each of the 3 days of validation and in stability testing.

All R^2 values were greater than 0.99 in all validation. Data of the standard curve with regards to correlation, slope, R^2 and intercept are shown in Table 3 and Figure 4. Data in the three validation days were within the accepted ranges in terms of linearity.

Stability

From Tables 4 and 5, it can be observed that autosampler stability test is within the accepted ranges where it doesn't exceed 15%.

Data of short term stability is specified by two QC samples (low and high) for glimepiride, after preparation method (autosampler stability), $T=10^\circ\text{C}$.

Specificity

Protein extraction procedure was specified and sensitive for glimepiride. Treated blank and zero samples from the six batches used clean chromatogram for specific method (Figures 1, 2 and 3)

Effect of juices on pharmacokinetic parameters of glimepiride

Glimepiride is metabolized by CYP450 and it is a substrate for p-GP OATP2B1 transporter in human²⁵. As shown in Figure 5, when glimepiride was administered alone, its serum level reached its maximum (258.711 ± 7.93 ng/ml) after half an hour and then gradually declined to get a minimum concentration of (130.2 ± 22.36 ng/ml) after 6 hours from the administration of glimepiride.

When glimepiride was administered in combination with grapefruit, the maximum serum concentration (266.226 ± 46.48 ng/ml) was reached after half an hour and then gradually declined to reach (131.06 ± 27.58 ng/ml).

On the other hand, combination of glimepiride and licorice showed an increasing in

serum concentration approximately twice as much as the concentrations when glimepiride is given alone. At the first half, an hour of administration, it reached its maximum serum concentration (559.585 ± 44.45 ng/ml) and then gradually declined to get a minimum concentration of (81.928 ± 20.84 ng/ml) at the end of follow up period (6 hours).

Licorice significantly ($p < 0.05$) increased the AUC_{0-6} , C_{max} and $t_{1/2}$ of glimepiride and decreased significantly ($p < 0.05$) clearance and elimination rate constant. But grapefruit juice had no significant ($p > 0.05$) effect on glimepiride pharmacokinetics. The calculated kinetic parameters are shown in Table 6.

A mechanism of this increasing may be due to the suppression of metabolic enzymes required for the metabolism of glimepiride expressed as elongation in elimination half-life and decrease total body clearance as well as increase in AUC_{0-6} . Significant change in elimination parameters suggests an interaction on elimination level. Many studies showed an inhibitory effect of liquorice on P-glycoprotein in the intestinal wall. Data of this study is not enough to characterize possible interaction on absorption level. However, t_{max} as an indicative parameter for absorption rate was the same when glimepiride was given alone and with liquorice.

While grape fruit juice did not affect any kinetic parameters of glimepiride. This interaction could be of clinical significance, because elevated glimepiride concentrations may increase the risk of hypoglycemia. Concomitant usage of licorice with glimepiride warrants close monitoring of blood glucose.

CONCLUSION

A validated bioanalytical procedure with high resolution and sensitivity was developed for the evaluation of glimepiride levels in rat's plasma. The effect of fresh liquorice beverage on the pharmacokinetic profile for glimepiride was shown to cause an increase in their bioavailability and decrease in its elimination. This is may be due to enzyme activation involved in its metabolism. While administration of grape fruit juice with glimepiride did not affect its pharmacokinetic profile.

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