The Neuroprotective Effects of Alcoholic Extract of *Levisticum* Officinale on Alpha Motoneurons' Degeneration After Sciatic Nerve Compression in Male Rats

SALMAN MAHMOUDZEHI¹, GOL MOHAMMAD DORRAZEHI^{2*}, SIROUS JAMALZEHI³, AMIR HOSSEIN HEYDARI KHABBAZ⁴, FATEMEH GHORBANI¹, ABDOLLAH HOOTI⁵, ABDOLVAHED GHORBANI DADKANI¹ and MEHRDAD MAHMOUDI SOURAN¹

¹Department of Biology, Faculty of science, University of Sistan and Baluchistan, Zahedan, Iran. ²M.Sc in Genetic Engineering and Molecular Biology, Faculty of Biotechnology and Biomolecular Science, University Putra Malaysia (UPM), Selangore, Malaysia.

³School of Nursing and Midwifery, Iranshahr University of Medical Sciences, Iranshahr, Iran. ⁴Faculity of Veterinary, University of Zabol, Zabol, Iran.

⁵Induced Pluripotent Stem Cell Biotechnology Team, Stem Cells Department, National Institute of Genetic Engineering and Biotechnology, Tehran, Iran.

*Corresponding author E-mail: goldorrazehi@gmail.com

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ABSTRACT

As peripheral neurons are injured, several inflammatory factors will be released from the fiber of the injured nervous leading to morphological and biochemical changes in the injured point. This changes ultimately results in the Wallerian degeneration of the cell bodies of these neurons in the spinal cord which are involved in retrograde reaction. Levisticum officinale is one of the herbs that has been traditionally used for treatment of many diseases. Many studies have confirmed the anti-inflammatory and anti-oxidant effects of this plant. It is expected that the extract of Levisticum officinale would have a significant role in the restoration of nervous system injuries. The purpose of this study was to investigate the effects of the alcoholic extract of Levisticum officinale on the neuron protection of the sciatic nerves of injured rats. In this study, the ethanolic extract was obtained from Levisticum officinale. Then the rats were randomly divided into 5 groups, each containing 6 rats, including two control and compression groups and three treatment groups treated with 50, 75 and 100 mg/kg doses of the extract. Sciatic nerve in compression and treatment groups was exposed to compression for 60 seconds, and the ethanolic extract was injected intraperitoneally in the first, second and third weeks. The spinal cord of the rats were sampled and the neurons of the samples were counted using a dissector method after tissue passaging and staining. Meanwhile, the density of motoneurons was measured. T-test analysis was performed by using SPSS software. The density of a motoneurons in compression group decreased significantly as compared to the control group (p<0.001). The density of a motoneurons in all treatment groups with 50,75 and 100 extract doses was significantly increased as compared to the compression group (p<0.01). The highest neuron protection was observed in the treatment group with 100 mg/kg dose as compared to the compression group. Ethanolic extract of Levisticum officinale has neuron protective and restorative effects on the injuries of the peripheral nerves such as compression.

Keywords: Levisticum officinale, Neuron protection, á motoneurons, Wallerian degeneration.

INTRODUCTION

When the cell body of neurons is destroyed or detached from their respective axon due to illness,

injury or other factors, Wallerian degeneration occurs in the distal part of the axon, destroying the axons and their myelin sheath¹. When Wallerian degeneration occurs, the distal part of the axon becomes swollen and chaotic during a few days and neurofibrils rapidly disappear, followed by the breaking of axon into smaller pieces. in the early days after degeneration, myelin sheaths of nerve fibers break into small elliptical pieces and gradually blow apart, and the axon residues and myelin sheaths become destroyed by macrophages^{2,3}. If the damage is high, the effects of injuries, backward to the cell body, would cause central degeneration of the nerve and bodies break. This process is called chromatolysis. Furthermore, the nucleus shifts from the central position to the side position⁴. This process is faster in the peripheral nerves and all the steps are completed in a few weeks, while completion takes several months in the central nervous system. In addition, in an intact neuron cell body, healing of peripheral nerves progresses remarkably in contrast to the central nervous system^{3,5,21}.

Levisticum officinale is one of the most important herbs and medicinal plants that has been used for a long time. Roots, fruits and leaves of this plant have medicinal characteristics. Alcoholic extract of this plant is produced for treatment of kidney stones and urinary tract infections in the pharmaceutical industries. The most important compounds identified in different parts of the plant are: Z-ligustilide, â-Phellandrene, á-terpinyl acetate, E-ligustilide and flavonoids like Quercetin⁶. Flavonoids are effective in destroying free radicals. Due to the presence of Quercetin in Levisticum officinale, the antioxidant effect is obvious. One of the most important roles of antioxidant compounds is their anti-inflammatory effect. So far, the neuroprotective effect of the extract of this plant has not been studied. So with regard to anti-inflammatory effect of this plant, we decided to examine the effect of the ethanolic extract of this plant on Wallerian degeneration of the sciatic nerve^{6,} 7.

Axons in the peripheral nervous system (PNS) can ease the process of restoration but this process in the central nervous system (CNS) often fails. Regeneration of axons in the central nervous system is not completely successful and shows poor healing response to injury when the environment is harnessed towards the growth of axons, But peripheral neurons can easily follow the process of restoration^{3,8}. It is believed that motor neurons get trophic factors from the target organs and without

these factors the preservation is not possible. The loss of the protective effect of such factors because of denervation is likely to lead neurons towards exclusion from trophic factors and in most cases they lead to cell death^{9,10}. Related research shows that the degeneration of the pre ganglion may be a general phenomenon that occurs after peripheral nerve injury. Cutting dorsal root L4 - L6 of spinal nerves in rats begin the process of degeneration in the affected area and extend to the dorsal column at about 3 mm per hour¹¹.

Followed by sciatic nerve compression, many biological events occur in cellular and molecular level such as apoptosis, increase of the entry of calcium into neurons, release of excitatory neurotransmitters like glutamate, formation of free radicals and activation of inflammatory processes¹². Studies have shown that apoptosis occurs after sciatic compression and expression level of many genes increases (apaf1, BAX, Caspase 3,9). TNF-á is known as one of a Wallerian degeneration starters, which is an inflammatory cytokine that starts systemic inflammation and its concentration increases after peripheral nerve axon injury^{2,12}.

Flavonoids also inhibit production of inflammatory cytokines like IL6, IL1 and TNFá^{16,17}. Some of the flavonoids also inhibit NF-kB pathway. Inhibiting this pathway causes a decrease in TNF-á production^{18,19}. Quercetin is one of the flavonoids in this plant that has anti-inflammatory effect invivo. Quercetin inhibits inflammatory cytokines through weakening NF-kB and p38. This component also causes a decrease in miloperoxydase and oxidative stress and inhibits apoptosis^{16,20}. In a research, the amount of Quercetin was measured between 32 plants and the amount of Quercetin in Levisticum officinale was the highest level as compared to the 32 plants^{7,13}. Studies on Levisticum officinale have proven very important effects on several cases. In many of such studies, the ingredients of these plants have been investigated, for both therapeutic and functional applications.

In 2003, researchers succeeded in determining the composition and the amount of poliphenolic compounds in the different parts of *Levisticum officinale*¹³. The compounds found in this plant justifies the effects observed. In a study

conducted in 2007 in Iran, it became clear that the essential oil from Levisticum officinale was characterized by high quantity of monoterpens (98.3%). The main components in the oil were ²-Phellandrene (42.5%) and \pm -terpineol (27.9%)¹⁴. Mirjalili et al. in 2010 reported composition and antibacterial effects of the essential oil of this plant. The researchers in this study determined the amount of many materials in flowers and fruits at different developmental stages. The current study also showed that Levisticum officinale has a strong antibacterial effect. This was a very important result⁶. In 2011, Serkan Sertel et al. showed this plant has an antiproliferative activity. This study proved that this plant has an anticancer effect because of its antiinflammatory compounds¹⁵. Some studies showed anti-inflammatory effects of this plant on kidney inflammations. Flavonoids existing in Levisticum officinale can justify this anti-inflammatory effects on kidney inflammations²².

MATERIALS AND METHODS

In this study, Wistar albino rats were used. The rats at approximately 3 months and weighing 250 to 300 grams were prepared from Animals Research Center of Zahedan University of Medical Sciences, Zahedan, Iran. All the rats were kept for one week to adapt to new conditions in animals' room in University of Sistan and Balouchestan, Zahedan in the same condition as standard with a temperature of 21 to 25 ° C and 12 hours of light and 12 hours darkness light-cycle. Then the rates were randomly divided into five groups with 6 animals in each group:

Control group

In this group, surgery was performed and

only skin and muscle of the right thigh was opened but the compression of the sciatic nerve was not affected and only normal saline was injected.

Compression group

This group was subjected to surgery with severe compression of the sciatic nerve in the region of the right thigh (for 60 seconds with cord blood pence grade 2). The group was intraperitoneal injected with normal saline.

Treatment group A, B and C

In these groups, severe compression of the sciatic nerve in the region of the right thigh was performed (like the compression group) and ethanolic extract at a dose of 50, 75 and 100 mg/kg was injected to groups A, B and C respectively for three times (one injection for each week).

Twenty-eight days after the rats were injected with different doses of the extract, perfusion was performed. For this purpose, the animals' chest area was cut, as the triangle of the heart of animal was clearly visible. Then the needle of perfusion machine that contained the fixator (formalin 10 %) was entered into the aorta through left ventricle. In this way, the fixator flows the entire body of the animal and the animal organs can be fixed in the shortest time. The head and the skin of the animals were separated and the spinal cord segments related to sciatic nerve (L4-L5 and S1-S3) were sampled. Then the samples were entered into tissue passage stages. After that, trim operation was performed by a microtome machine and the samples were subjected to staining with Toluidin blue- erythrosin.

| Sample | Dissector Number (Σ frame) | Neurons Number ΣQ | Neuronal Density (ND) |
|------------|------------------------------------|----------------------|--------------------------|
| C1 | 30 | 63 | 1750 |
| C2 | 30 | 55 | 1527 |
| C3 | 30 | 62 | 1722 |
| C4 | 30 | 60 | 1666 |
| C5 | 30 | 58 | 1607 |
| C6 | 30 | 62 | 1722 |
| Means ± SE |) 30 | 60 | 1665.66±77.59 |

Table 1: Neuronal Density in Control group

Dissector method was used to count the density of neurons. This method consists of two parallel cutting that have a known distance. Neurons are counted in the frame of reference. For analysis of the raw data, some parameters such as \pounds Q) total neurons counted in a sample)_i \pounds Frame)Total number of sampled) V dissector (the volume of the sample) were calculated by the formula: V dissector = A Frame × H (A Frame as area of sampling frame and H as the distance between two slices or a thickness of each cut)) and Mean \pm SE are required. Numerical Density (ND was calculated by (: \pounds Qframe × dissector For statistical analysis, t-test and one way Anova test with significance level P<0.05 were used and were plotted using charts Excel software.

RESULTS AND DISCUSSION

The results of the study on effects of alcoholic extract of *Levisticum officinale* on neuronal

| Sample | Dissector Number (Σ frame) | Neurons Number ΣQ | Neuronal Density (ND) |
|------------|------------------------------------|----------------------|--------------------------|
| C1 | 30 | 26 | 722 |
| C2 | 30 | 30 | 833 |
| C3 | 30 | 28 | 777 |
| C4 | 30 | 28 | 777 |
| C5 | 30 | 25 | 690 |
| C6 | 30 | 26 | 722 |
| Means ± SD |) 30 | 27 | 753.5±47.33 |

Table 2: Neuronal Density in Compression group

Table 3: Neuronal Density in Treatment group A

| Sample | Dissector Number (Σ frame) | Neurons Number ΣQ | Neuronal Density (ND) |
|---------|------------------------------------|----------------------|--------------------------|
| C1 | 30 | 42 | 1167 |
| C2 | 30 | 39 | 1083 |
| C3 | 30 | 45 | 1250 |
| C4 | 30 | 39 | 1083 |
| C5 | 30 | 44 | 1222 |
| C6 | 30 | 43 | 1194 |
| Means ± | SD 30 | 42 | 1166.5±64.23 |

Table 4: Neuronal Density in Treatment group B

| Sample | Dissector Number (Σ frame) | Neurons Number ΣQ | Neuronal Density (ND) | |
|-----------|------------------------------------|----------------------|--------------------------|--|
| C1 | 30 | 49 | 1360 | |
| C2 | 30 | 52 | 1444 | |
| C3 | 30 | 55 | 1527 | |
| C4 | 30 | 54 | 1502 | |
| C5 | 30 | 50 | 1387 | |
| C6 | 30 | 52 | 1444 | |
| Means ± S | SD 30 | 52 | 1444±58.53 | |

density in the anterior horn of the spinal cord in treatments, compression and control groups as counting neurons are presented in the tables 1 to 6 as mean \pm standard deviation.

Neuronal density in compression group in comparison to the control group was significantly decreased because of the retrograde reaction (compression group 753.5±57.33, control group



Fig. 1: Density of alpha motoneurons of the anterior horn of the spinal cord of the control and compression groups

| Sample | Dissector Number (Σ frame) | Neurons Number ∑Q | Neuronal Density (ND) |
|------------|------------------------------------|----------------------|--------------------------|
| C1 | 30 | 54 | 1502 |
| C2 | 30 | 51 | 1416 |
| C3 | 30 | 51 | 1416 |
| C4 | 30 | 56 | 1556 |
| C5 | 30 | 57 | 1583 |
| C6 | 30 | 55 | 1527 |
| Means ± SE | 30 | 54 | 1500±64.37 |

Table 5: Neuronal Density in Treatment group C

Table 6: Comparing of Neuronal Density in all groups

| Sample | Control | Compression | Treatment A | Treatment B | Treatment C |
|------------|---------------|--------------|--------------|-------------|-------------|
| | group | group | | | |
| C1 | 1750 | 722 | 1167 | 1360 | 1502 |
| C2 | 1527 | 833 | 1083 | 1444 | 1416 |
| C3 | 1722 | 777 | 1250 | 1527 | 1416 |
| C4 | 1666 | 777 | 1083 | 1502 | 1556 |
| C5 | 1607 | 690 | 1222 | 1387 | 1583 |
| C6 | 1722 | 722 | 1194 | 1444 | 1527 |
| Means ± SD | 1665.66±77.59 | ±47.33 753.5 | 1166.5±64.23 | 1444±58.53 | 1500±64.38 |



Fig. 2: Neuronal densities of the control, compression and different treatment groups

1665.66±77.59, P<0.001) (figure 1). Neuronal density in treatment group A with a dose of 50 mg/ kg alcoholic extract (1166.5±64.23) as compared to the compression group showed a significant increase (753.5±47.33, P<0.05). Density of alpha motoneurons of the anterior horn in treatment group B, with alcoholic extract of the plant at a dose of 75 mg/ kg (1444 ± 58.53) as compared to the compression group (753.5±47.33) showed a significant increase (P<0.01). The most neuroprotective effect was observed in treatment group C with 100 mg/kg alcoholic extract as compared to both of the other treatment groups, and showed a significant increase as compared to the compression group (1500±64.38, 753.5±47.33, P<0.01) (figure 2).

According to previous related studies, this plant is a rich source of anti-inflammatory compounds that has therapeutic effects on some organs' inflammation and also *Levisticum officinale* has strong anti-bacterial and anti-cancer therapeutic effect. This study was designed, based on the results of the previous studies conudcted. based on the anti-inflammatory effects of this herb that has been demonstrated in previous studies, the results obtained in this study is in sufficient agreement with its reported medicinal potential. Existing antiinflammatory compounds like Quercetin cause inflammation and suppression of nerve cells. However, recovery of the cell body and the fibrils of neurons in peripheral nervous system is repairable. This study demonstrated that this plant has a potential capacity of neurons protection.

CONCLUSION

According to this study, neuronal density in compression group as compared to the control group had a significant decrease. It means that the animal sciatic nerve compression leads to the creation of retrograde central degeneration to the cell bodies of motor neurons in the anterior horn of the spinal cord. Based on the results, the treatment groups with alcoholic (ethanolic) extract showed more neuronal density than the compression group and also injected dose-dependent effects were observed significantly. Additionally, the dose of 100 mg/kg had the highest effect, dose of 75 mg/kg had average effects and dose of 50 mg/kg had the least effect on restoration of nervous system injuries. Therefore, the injection of alcoholic (ethanolic) extract of Levisticum officinale has both repair and restoration effects on peripheral nerves (such as sciatic nerve) injuries. Further studies by increasing the number of samples and increasing the number of injections will generate more accurate results and provide better evaluation of the effects of plant extract. Study on neuroprotective effects of the methanolic extract of this plant and also monitoring the behavioral tests in rats helps to examine the applicability of Levisticum officinale extracts for animal healthcare.

REFERENCES

- Stoll G, Jander S, Myers RR: Degeneration and regeneration of the peripheral nervous system: from Augustus Waller's observations to neuro inflammation. *J Peripher Nerv Syst*, 7:13-27, (2002).
- Galiano M, Liu ZQ, Kalla R, Bohatschek M, Koppius A, Gschwendtner A, Xu S, Werner A, Kloss CU, Jones LL, Bluethmann H & Raivich G. Interleukin-6 (IL6) and cellular response to facial nerve injury: effects on lymphocyte recruitment, early microglial activation and axonal outgrowth in IL6-deficient mice. *Eur J Neurosci;* 14: 327–341, (2001).
- Vargas ME, Barres BA. Why is Wallerian degeneration in the CNS so slow? *Annu Rev Neurosci*, **30**:153-179, (2007).
- Mason MR, Lieberman AR, Grenningloh G & Anderson PN. Transcriptional upregulation of SCG10 and CAP-23 is correlated with regeneration of the axons of peripheral and central neurons invivo. *Mol Cell Neurosci*, 20: 595–615, (2002).
- Martini R, Fischer S, Lopez-Vales R, David S. Interactions between Schwann cells and macrophages in injury and inherited demyelinating disease. *Glia*, 56: 1566-1577 (2008).
- Mirjalili.M, Negadabrahi.S, yousefzadi.M, Hadian.G, Salehi.P and Sonboli.A. The composition and antibacterial activity of the essential oil of Levisticum officinale Koch flowers and fruits at different developmental stages. J Serb. Chem. Soc., 75(12): 1661– 1669 (2010).
- Wojdylo.A,. Oszmianski. J. Czmerys. R. Antioxidant activity and phenolic compounds in 32 selected herbs. *Food Chemistry;* 105: pp. 940-949 (2007).
- Aguayo AJ, David S, Richardson P, Bray G. Axon elongation in peripheral and central nervous system transplants. In: Advances in Cellular Neurobiology. Fedoroff S, Hertz L (Eds). Academic Press, New York, 3: 215–234 (1982).
- Sunderland S. Nerve and nerve injuries.
 2nd edition. Edinburgh (United Kingdom): Churchill Livingstone.35-9; (1978).
- 10. Beirowski B, Adalbert R, Wagner D, Grumme

D, Addicks K, Ribchester R, *et al*: The progressive nature of Wallerian degeneration in wild-type and slow Wallerian degeneration (WIdS) nerves. *BMC Neuroscience*, :6 (2005).

- 11. Arvidsson J, Ygge J, Grant G . Cell loss in lumbar dorsal root ganglia and transganglionic degeneration after sciatic nerve resection in the rat.*BrainRes*, **14**; 373(1-2):15-21 (1986).
- Oppenhheim. JJ, Feldman. M. Introduction to the role of cytokines in innate and defense and adaptive immunity. In: Oppenhheim JJ, Feldman M, editors. Cytokine reference. New York: Academic, 3–20 (2001).
- Agnieszka Najda, Tadeusz Wolski, Jan Dyduch, Tomasz Baj. Determination of quantitative composition of poliphenolic compounds occur in anatomically different parts of Levisticum officinale Koch, *Electronic Journal of Polish Agricultural Universities;* 6(1) (2003).
- 14. Verdian Rizi Mohammad Reza, Hadjiakhoondi Abbas. The essential oil composition of Levisticum officinale from Iran, *Asian journal* of biochemistry, **2**(2), 161-163 (2007).
- 15. Serkan Sertel, Tolga Eichhorn, Peter k.Plinkert and Thomas Efferth. Chemical composition and antiproliferative activity of essential oil from the leaves of a medicinal herb, Levisticum officinale, against UMSCC1 head and neck squamous carcinoma cells, *Anticancer Research*, **31**: 185-192 (2011).
- Brewer MS. Natural antioxidants: sources, compounds, mechanisms of action, and potential applications . *Compr Rev Food Sci F*; **10**(4): 221 (2011).
- Oh H, Kim DH, Cho JH, Kim YC. Hepatoprotective and free radical scavenging activities of phenolic petrosins and flavonoids isolated from Equisetum ravens. *J Ethnopharmacol.;* 95(2-3):421-4 (2004).
- Kerschensteiner M, Schwab ME, Lichtman JW, Misgeld T. In vivo imaging of axonal degeneration and regeneration in the injured degeneration and regeneration in the injured, 100: 1520-29 (2000).
- 19. Heuman R .Korschinh a .Bandlow.c .chengo of nerve growth factor synthesis non normal

cell in respon to sciatic nerve ternsection *.j cell boil.* **104**: 1653.1631 (1987).

- 20. Camara-Lemarroy CR, Guzman-de la Garza F, Fernandez-Garza NE: Molecular Inflammatory Mediators in Peripheral Nerve Degeneration and Regeneration. *Neuro immune modulation*, **17**:314-324 (2010).
- 21. Behnam Rasouli, M Nikravesh. M Mhadavi

Shahri, N Tehranipour M. Post operative timeeffects after sciatic nerve crush on the number of alpha motoneurons, using a sterologicalcounting method (disector). *Iran Biomed J*, **4**(1): pp.45-49 (2000).

 Wendell Combest, Marian Newton, Austin Combest, June Hannay Kosier. Effects of herbal supplements on the kidney. *Urologic nursing*, 25(5); (2005).